



# LncRNA-TCONS\_00034812 is upregulated in atherosclerosis and upregulates miR-21 through methylation in vascular smooth muscle cells

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**Background:** LncRNA-TCONS\_00034812 is a critical player in the proliferation of aortic smooth muscle cells. It is known that artery injury plays an important role in atherosclerosis. However, the potential implication of LncRNA-TCONS\_00034812 in atherosclerosis remains unclear. In this study, we collected artery specimens from patients with atherosclerosis and healthy controls to investigate the involvement of LncRNA-TCONS\_00034812 in atherosclerosis.

**Methods:** Sixty patients with atherosclerosis and 60 controls, admitted at The First Hospital of Changsha (Changsha, China), between March 2017 and March 2019, were included. An artery biopsy was performed on all participants to obtain the artery specimens. Real-time quantitative PCR were performed to quantify the relative expression level of LncRNA-TCONS\_00034812. Its role in atherosclerotic lesion was evaluated in (high fat diet) HFD-induced ApoE<sup>-/-</sup> mice. Moreover, human aortic smooth muscle cells (HAOSMCs) was employed to study functional role of LncRNA-TCONS\_00034812 overexpression and knockdown by methylation-specific PCR and cell proliferation assay.

**Results:** Overexpression of TCONS\_00034812 resulted in miR-21 upregulation and a decrease of miR-21 gene methylation. In contrast, silencing of TCONS\_00034812 caused miR-21 downregulation and an increase of miR-21 gene methylation. Cell proliferation analysis indicated that the overexpression of TCONS\_00034812 and miR-21 promoted cell proliferation, while silencing of TCONS\_00034812 played an opposite role. Moreover, miR-21 overexpression weakened the effects of silencing TCONS\_00034812 on cell proliferation.

**Conclusions:** In summary, LncRNA-TCONS\_00034812 is upregulated in atherosclerotic samples, and its overexpression upregulates miR-21 through methylation in human aortic smooth muscle cells (HAOSMCs). Our study indicates that LncRNA-TCONS\_00034812 could serve as a potential biomarker for diagnosis of atherosclerosis.

**Keywords:** LncRNA-TCONS\_00034812; atherosclerosis; miR-21; methylation; aortic smooth muscle cells

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## Introduction

Atherosclerosis is caused by the formation of plaques inside arteries, which are the result of an accumulation of cholesterol, fat, calcium, and other substances (1). Although atherosclerosis is not lethal, it increases the risk of other lethal diseases of the heart, such as coronary heart disease (2). At present, cholesterol-lowering therapy is widely used to treat early-stage atherosclerosis, which is reversible (3,4). However, advanced atherosclerosis is not reversible, and cholesterol-lowering therapy is not effective for such cases (3,4). Smoking, high blood pressure, and high cholesterol are the main risk factors for atherosclerosis (5). However, the molecular pathogenesis of this disease is still largely unknown (6), which limits the development of effective therapies.

DNA methylation is a crucial mechanism modulating genes in cell differentiation and pathological processes. Aberrant DNA methylation such as hypermethylation of DNA in the promoter region of tumor suppressor genes, is a well-studied epigenetic marker in various cancers (7). Methylated DNA immunoprecipitation (MeDIP), methylated CpG island amplification (MCA) as well as whole genome bisulfite sequencing have been used to profile methylation pattern changes in atherosclerotic samples (8-10). Microarray and RNA-seq have been successfully applied to profile the transcriptomic changes in the pathogenesis of atherosclerosis (11,12). Many atherosclerosis-related genes have been found to be regulated through DNA methylation mechanism (13). Several recent studies demonstrated that patients with atherosclerosis display heightened level of DNA methylation in aortic samples and blood samples (14,15), and the level of DNA methylation seems to correlate with atherosclerotic lesion progression (16). Altered methylation pattern modulates the expression changes of genes in inflammation, aging and oxidative stresses (13). For example, hypermethylation of the estrogen receptor gene ( $ER\alpha$ ) is associated with aging and atherosclerosis, which may function to regulate smooth muscle cell proliferation (17). Genes associated with atherosclerosis, such as inflammatory markers and coagulation markers, can be used as biomarkers for cardiovascular risk prediction by the receiver operating characteristic (ROC) curve method (18).

Non-coding RNAs (ncRNAs) include miRNAs and long (>200 nt) ncRNAs (lncRNAs). NcRNAs participate in diverse pathological and physiological processes mainly by regulating gene expression (19). A large number of differentially expressed ncRNAs have been found in atherosclerosis and some of them are critical players in the regulation of vascular

smooth muscle cells (VSMCs), which have important roles in the occurrence and development of atherosclerosis (20,21). Atherosclerosis is initiated by endothelial dysfunction and neovascularization is a prominent feature of atherosclerotic lesions (22). The hyperplasia of the vasa vasorum in the artery wall is followed by neovascularization within the atherosclerotic plaque, and plaque hemorrhage is a feature of unstable plaques frequently happening in intimal neovessels (23). LncRNA-TCONS\_00034812 was showed to regulate cell proliferation and apoptosis of pulmonary artery smooth muscle cells and its mechanism (24). However, its role in the neovascularization and plaque formation in atherosclerotic lesions is unclear. In addition, miR-21 targets tropomyosin 1 to promote the proliferation of VSMCs (25). In this study, we compared LncRNA-TCONS\_00034812 and miR-21 expression in intima samples from healthy controls and atherosclerotic patients. We found both LncRNA-TCONS\_00034812 and miR-21 expression was significantly upregulated in atherosclerotic samples, and atherosclerosis lesions were ameliorated by TCONS\_00034812 knockdown in HFD-induced ApoE<sup>-/-</sup> atherosclerosis mouse model. The overexpression of LncRNA-TCONS\_00034812 and miR-21 also promoted the proliferation of human aortic smooth muscle cells (HAOSMCs) expression. These results suggest that LncRNA-TCONS\_00034812 and miR-21 may serve as a biomarker and therapeutic target for atherosclerosis. We present the following article in accordance with the MDAR reporting checklist (available at <https://dx.doi.org/10.21037/atm-21-2632>).

## Methods

### Participants

Sixty patients with atherosclerosis and 60 controls, admitted at the Second Xiangya Hospital of Central South University between March 2017 and March 2019, were included. The clinical and demographic information for patients and controls is listed in *Table 1*. There were 33 males and 27 females among the patients with atherosclerosis with an age range of 23–35 years (mean age 29.1±4.3 years). The 60 controls included 33 males and 27 females with an age range of 23–34 years (mean age 29.0±4.1 years). Histological analysis was performed on all patients to confirm atherosclerosis. All patients were at first-time diagnosis. Patients with other clinical complications or who had received other therapies in the previous 6 months were excluded. All procedures performed in this study involving human participants were in accordance with the Declaration of Helsinki (as revised in 2013). The

**Table 1** Clinical data of patients and controls

Variable	Atherosclerosis (n=60)	Controls (n=60)
Age	29.1±4.3	29.0±4.1
Gender		
Male	33	33
Female	37	37
Smoking	29	28
hs-CRP (mg/L)	4.67±0.23	3.94±0.48
TG (mmol/L)	1.72±1.11	1.49±0.56
TC (mmol/L)	4.19±0.38	4.40±0.41
LDL-C (mmol/L)	2.30±0.30	2.27±0.33
HDL-C (mmol/L)	1.23±0.40	1.71±0.29

hs-CRP, high-sensitivity C-reactive protein; TG, triglycerides; LDL-C, low-density lipoprotein cholesterol; HDL-C, high-density lipoprotein cholesterol.

study was approved by medical ethics board of the Second Xiangya Hospital of Central South University, Changsha, China (No.: PJ-KY-2020-128) and informed consent was taken from all the patients.

### Artery specimens

An artery biopsy was performed on all participants to obtain the artery specimens. The artery biopsy was collected from the atherosclerosis-affected sites of the atherosclerosis patients. The 60 controls were diagnosed with suspected artery lesions, but lesions were subsequently excluded following biopsy. All specimens were from the temporal artery of the control subjects. Intima, the inner layer of the artery specimens, was separated and maintained in liquid nitrogen before use.

### Human aortic smooth muscle cells (HAOSMCs)

HAOSMCs (354-05A, Sigma-Aldrich, , Saint Louis, Missouri, USA) were used in this study. The HAOSMCs were cultivated under the conditions described in the protocol from Sigma-Aldrich. HAOSMCs were harvested at passage 3 to 5 to perform the subsequent experiments.

### Cell transfections

As a backbone, the pcDNA3.1 vector was used to construct

the TCONS\_00034812 (10) expression vector. MiR-21 mimic and negative control (NC) miR-21 were acquired from Sigma-Aldrich. HAOSMCs were collected at about 80% confluence and were counted. Lipofectamine 2000 (a kind of Invitrogen, Sigma-Aldrich, Saint Louis, Missouri, USA) was applied to transfect  $10^6$  cells with 10 nM vector, 40 nM siRNA, or 40 nM miRNA. The cells that were not transfected were adopted as the control (C) cells. The cells transfected by the empty vector, NC siRNA, or NC miRNA served as NC cells. Subsequent experiments were carried out 48 h after the transfection.

### RNA preparation and real-time quantitative PCR (RT-qPCR)

The extraction of total RNAs from the intima samples and HAOSMCs was completed with Trizol (Sigma-Aldrich, Saint Louis, Missouri, USA). Eighty-five percent ethanol was applied to precipitate and wash the RNA samples, in order to harvest miRNAs. The extracted RNA samples were treated with a gDNA eraser (Takara), to digest the genomic DNAs. RNA concentrations were measured by a NanoDrop™ 2000 spectrophotometer (Thermo Fisher, Waltham, MA, USA). Reverse transcription was performed with a QuantiTect Reverse Transcription Kit (QIAGEN, Düsseldorf, Germany) with total RNA as the template to complete the synthesis of the cDNA samples. PCR reactions were prepared with a KAPA SYBR® FAST qPCR Master Mix (Kapa Biosystems, Wilmington, Massachusetts, USA). The TCONS\_00034812 expression was estimated with GAPDH as the endogenous control. The MiR-21 expression was calculated by using the All-in-One™ miRNA qRT-PCR Reagent Kit (Genecopoeia, Rockville, Maryland, USA). The expression levels of miR-21 were normalized to the endogenous control miR-21. The researchers repeated all experiments three times. The mean values were counted by the  $2^{-\Delta\Delta CT}$  method to determine the fold changes of the gene expression levels.

### Animal protocol

Male ApoE<sup>-/-</sup> mice were acquired from Beijing HFK Bioscience Co., Ltd. (Beijing, China). The 6-week-old mice weighed 28–32 g. An atherogenic diet was fed to the ApoE<sup>-/-</sup> mice for 4 weeks to create an atherosclerosis model. The atherogenic diet included 2% cholate, 10% cholesterol, 10% lard, and 78% basal feed. After the establishment of

the model, the blood samples gathered via cardiac puncture were centrifuged at 1,500  $\times$ g at 4 °C for 10 min to acquire the serum. Subsequently, the mice were sacrificed. The aortic sinuses of the mice were cut out and stored in liquid nitrogen before use. Experiments were performed under a project license (No.: 20201115S0310211[308]) granted animal use ethics board of the Second Xiangya Hospital of Central South University, Changsha, China, in compliance with institutional Animal Care and Use Guidelines for the care and use of animals.

#### *Atherosclerotic lesion analysis by hematoxylin and eosin staining*

After the dissection, the aortic sinuses were put in a 4% paraformaldehyde solution dehydrated with alcohol. Then the aortic sinuses were embedded in paraffin. Serial paraffin sections were made with a thickness of 5  $\mu$ m, then stained with hematoxylin and eosin (H&E) (Histoserv, Germantown, MD, USA). The necrotic core quantification and lesion areas were examined. An Olympus fluorescent microscope (DP80; Olympus Corp., Tokyo, Japan) was used to observe the morphological changes.

#### *Methylation-specific PCR (MSP)*

At 48 h after transfection, the Genomic DNA Isolation Kit (ab65358, Abcam, Cambridge, UK) was used to extract the genomic DNAs. DNA samples were alternated by the EZ DNA Methylation-Gold™ kit (ZYMO, Irvine, California, USA). Routine PCR reactions were performed with the Taq 2X Master Mix (NEB, Beijing, China) to detect gene methylation.

#### *Cell proliferation assay*

A CCK-8 assay (Dojindo, Japan) was performed to analyze the proliferation of HAOSMCs (harvested at 48 h post-transfection). The 3,000 cells in a 0.1-mL cell culture medium were shifted to all the wells of a 96-well cell culture plate. At 4 h before cell collection, the cell cultivation was kept at 37 °C. Then CCK-8 solution was mixed into each well to achieve the ultimate concentration of 10%. Cells were harvested every 24 h up to 96 h. The OD value measurement was undertaken at 450 nm to report cell proliferation.

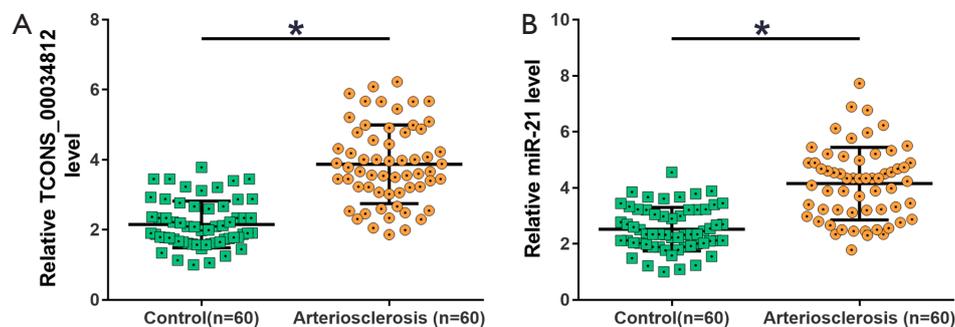
#### *Statistical analysis*

The above experiments were conducted in three independent biological replicates with the mean values calculated. The between-group differences were compared by unpaired *t*-tests, ANOVA (one-way), and the Tukey test. Correlations were analyzed by Pearson's correlation coefficient. A *P* value <0.05 was considered statistically significant.

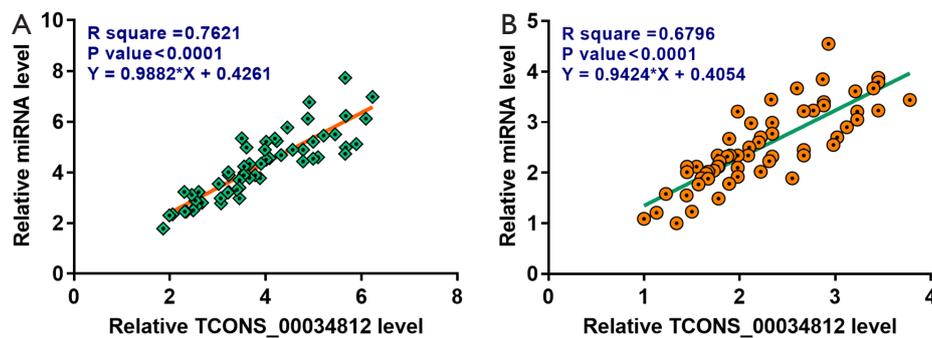
## Results

#### *TCONS\_00034812 and miR-21 were upregulated in the intima samples from atherosclerosis patients*

The expression levels of TCONS\_00034812 and miR-21 were measured from the intima samples of atherosclerosis patients (*n*=60) and controls (*n*=60). TCONS\_00034812 expression was visibly increased in the atherosclerosis group compared with the control group (Figure 1A, *P*<0.05), and MiR-21 expression was also markedly higher in the



**Figure 1** qPCR analysis revealed the upregulation of TCONS\_00034812 and miR-21 were upregulated in intima samples from atherosclerosis patients. Expression levels of TCONS\_00034812 (A) and miR-21 (B) in intima samples of atherosclerosis patients and controls were measured by performing RT-qPCR. \*, *P*<0.05. RT-qPCR, real-time quantitative PCR.



**Figure 2** TCONS\_00034812 and miR-21 were inversely associated across intima samples from atherosclerosis patients in a significant degree. The association between TCONS\_00034812 expression and miR-21 expression across intima samples in atherosclerosis patients (A) and controls (B) were explored by performing Pearson's correlation coefficient.

atherosclerosis group than in the control group (*Figure 1B*,  $P < 0.05$ ).

#### ***TCONS\_00034812 and miR-21 were positively associated in the intima samples***

The association of the expression levels between TCONS\_00034812 and miR-21 was explored by performing Pearson's correlation coefficient. Results demonstrated that TCONS\_00034812 expression and miR-21 expression were distinctly and positively associated across the intima samples of the atherosclerosis patients (*Figure 2A*). Furthermore, a positive and significant association between TCONS\_00034812 and miR-21 was also reflected in the intima samples of the controls (*Figure 2B*).

#### ***Atherosclerosis lesions were ameliorated by TCONS\_00034812 knockdown in HFD-induced ApoE<sup>-/-</sup> mice***

The atherosclerotic lesion areas were defined by H&E staining to further evaluate the effect of TCONS\_00034812 knockdown on atherosclerotic development in ApoE<sup>-/-</sup> mice. Results showed that injection with si-TCONS\_00034812 led to a more significant decrease in the atherosclerotic lesion area than in the si-NC group (*Figure 3*).

#### ***TCONS\_00034812 positively regulates the expression of miR-21 in HAOSMCs through methylation***

HAOSMCs were transfected by using the TCONS\_00034812 expression vector, TCONS\_00034812 siRNA or miR-21 mimic, to analyze the interaction between them. Transfections

were confirmed at 48 h post-transfection by performing RT-qPCR (*Figure 4A*,  $P < 0.05$ ). Compared with the NC and C groups, TCONS\_00034812 overexpression caused the upregulation of miR-21 ( $P < 0.05$ ) and decreased the methylation of the miR-21 gene (*Figure 4B*). In contrast, silencing of TCONS\_00034812 resulted in downregulation of miR-21 ( $P < 0.05$ ) and increased miR-21 gene methylation (*Figure 4C*), while miR-21 overexpression did not have any effect on TCONS\_00034812 expression (*Figure 4D*).

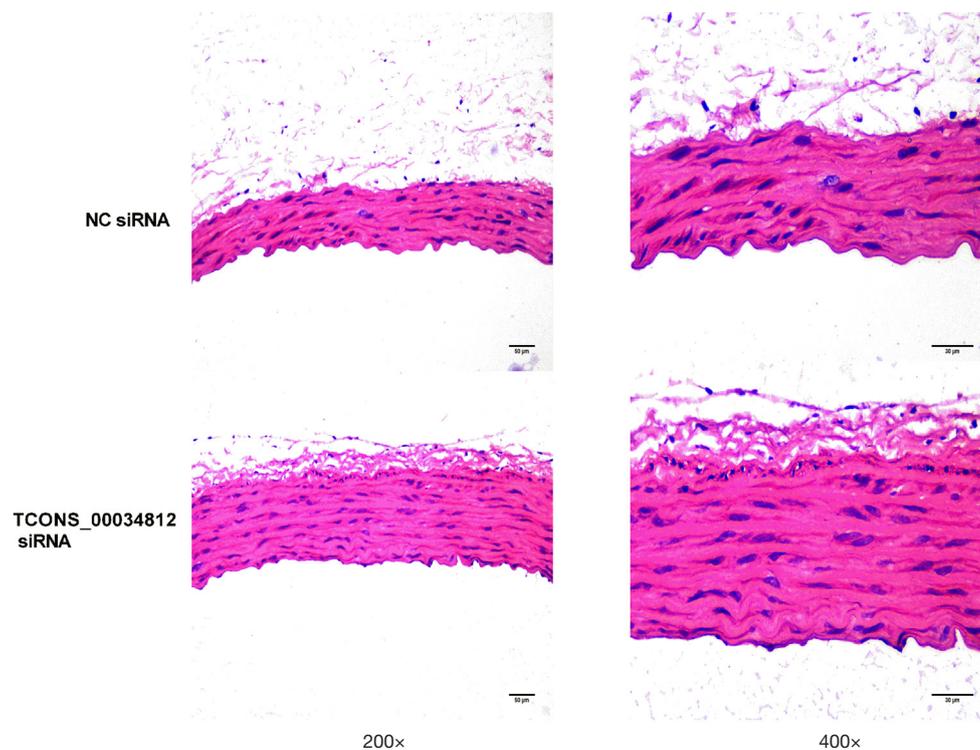
#### ***TCONS\_00034812 regulated miR-21 to promote the proliferation of HAOSMCs***

The HAOSMC proliferation affected by the transfections was analyzed by a CCK-8 assay. Overexpression of TCONS\_00034812 and miR-21 promoted cell proliferation but silencing of TCONS\_00034812 gave rise to a negative influence. In addition, miR-21 overexpression attenuated the consequences of silencing of TCONS\_00034812 (*Figure 5*,  $P < 0.05$ ).

## **Discussion**

The involvement of TCONS\_00034812 in atherosclerosis was established in the current study. Our study showed that TCONS\_00034812 was overexpressed in atherosclerosis and that it interacted with miR-21 through methylation to regulate the proliferation of HAOSMCs.

VSMCs play different roles at different stages of atherosclerosis (26-28). The increased proliferation rate of VSMCs can cause the formation of plaques, which then induce the development of atherosclerosis (26). In contrast, the aberrant proliferation of VSMCs in advanced



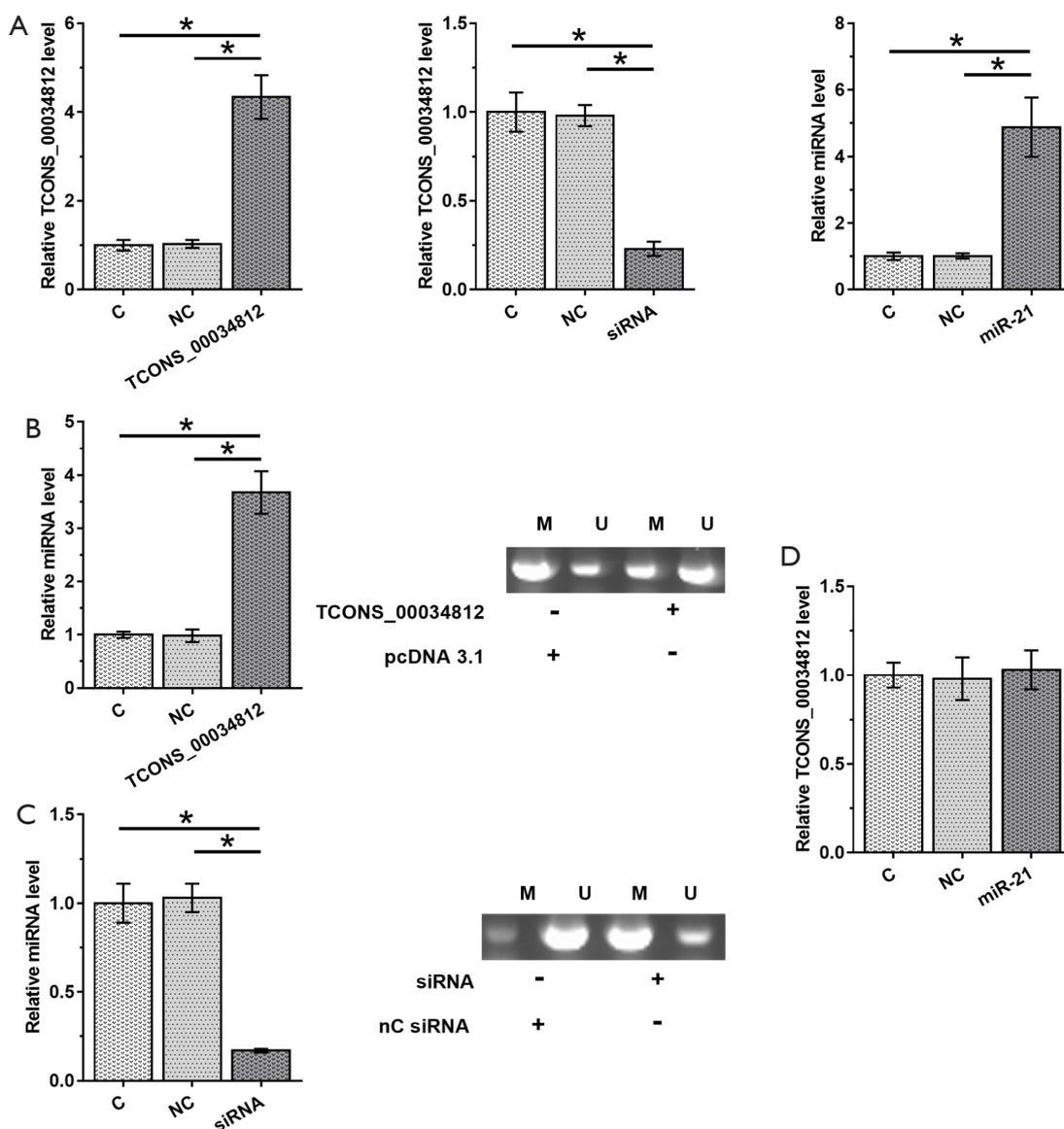
**Figure 3** Detected by HE, atherosclerotic lesion was improved by TCONS\_00034812 knockdown in HFD-induced ApoE<sup>-/-</sup> mice. Aortic sinus lesion area was showed to be smaller in si-TCONS\_00034812-injected mice than in si-NC-injected mice, \*, P<0.05. NC siRNA, small interfering RNA control.

stage atherosclerosis plays a protective role by preventing fibrous cap rupture (26). A recent study reported that TCONS\_00034812 was upregulated in a rat model of pulmonary artery hypertension, and its downregulation led to an increased proliferation rate of hypoxia in pulmonary artery smooth muscle cells (24). Interestingly, we also observed the upregulation of TCONS\_00034812 in intima samples from atherosclerosis patients. We also observed the enhancing effects of TCONS\_00034812 on the proliferation of HAOSMCs. The opposite results observed in this study might be due to the different types of VSMCs used in this study. More studies are necessary to investigate the influences of TCONS\_00034812 in the proliferation of other types of VSMCs.

A growing body of evidence indicates that dysregulation of lncRNA expression may contribute to the pathogenesis of atherosclerosis and vascular injury (29). For example, lncRNA MIAT is found to be significantly upregulated in atherosclerotic lesions, resulting in a sponging effect on miR-149-5p and the inhibition of efferocytosis (30). lncRNA-H19 is proposed to augment angiogenesis and

accelerates the development of atherosclerosis (31). Different lncRNAs are implicated in a wide spectrum of roles in atherosclerosis by recruiting leukocytes and immune cells, regulating lipid homeostasis and remodeling local vascular structures (29).

It has been reported that miR-21 is overexpressed in atherosclerosis and can target tropomyosin 1 to accelerate the proliferation of VSMCs (25). The same study also revealed the upregulation of miR-21 in atherosclerosis and its enhancing effects on the proliferation of VSMCs (25). Other research has reported that the methylation of miRNAs may be regulated by lncRNAs (32). Liu *et al.* showed that TCONS\_00034812 could negatively regulate miR-21 gene methylation to positively adjust the expression of miR-21, thereby participating in the proliferation of HAOSMCs (24). However, there may also be other mechanisms involved, and this would require more research to confirm. It is known that TCONS\_00034812 may regulate the proliferation and apoptosis of pulmonary artery smooth muscle cells by interacting with MAPK signaling, which plays a critical role in DNA hypomethylation (33).

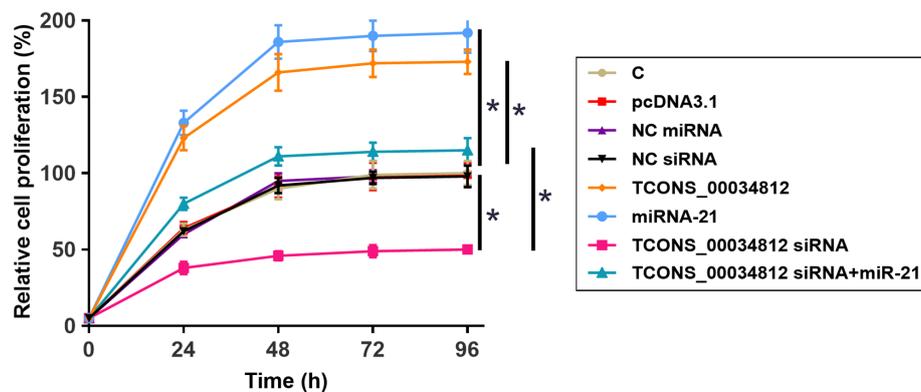


**Figure 4** TCONS\_00034812 positively regulate the expression of miR-21 in HAOSMCs through methylation. HAOSMCs were transfected with TCONS\_00034812 expression vector, TCONS\_00034812 siRNA or miR-21 mimic to analyze the interaction between them. Transfections were completed at 48 h after the transfection by proceeding RT-qPCR (A). The effects of overexpression of TCONS\_00034812 (B) and silencing of TCONS\_00034812 (C) on miR-21 expression and miR-21 gene methylation were evaluated by RT-qPCR and MSP. The effects of miR-21 overexpression on TCONS\_00034812 were estimated by RT-qPCR (D). \*, P<0.05. HAOSMCs, human aortic smooth muscle cells; C, untreated control; NC, miRNA control; NC siRNA, small interfering RNA control.

Therefore, MAPK signaling may mediate the regulation of miR-21 methylation by TCONS\_00034812.

Recent studies also highlighted a heightened level of DNA methylation in aortic samples and blood samples of patients with atherosclerosis (13,14), and the level of DNA methylation seems to correlate with atherosclerotic

lesion progression (15). Several factors could contribute to the development of atherosclerosis by modulating DNA methylation. For example, inflammation in cardiovascular diseases is reported to alter DNA methylation and proposed as a risk factor for atherosclerosis (34). Aging could lead to gene-specific hypermethylation, which may impact on genes



**Figure 5** TCONS\_00034812 regulated miR-21 to promote HAOSMCs proliferation. The effects of transfections on HAOSMCs proliferation were analyzed by CCK-8 assay. \*,  $P < 0.05$ . HAOSMCs, human aortic smooth muscle cells; C, untreated control; NC miRNA, miRNA control; NC siRNA, small interfering RNA control.

implicated in the atherosclerosis pathogenesis (17). On the other hand, chronic oxidative stress could also deregulate methylation signature of genes involved in the pathological development of atherosclerosis (35).

One limitation of this study is its small sample size. Also, our animal model experiments did not include an exploration of the interaction between TCONS\_00034812 and miR-21 in atherosclerosis. Our future studies will attempt to overcome these problems. Due to the increased incidence rate of atherosclerosis in young people in China, this study only included patients younger than 35 years. As it is known that atherosclerosis mainly affects older people, our future studies will include older patients to further explore the association between TCONS\_00034812 and miR-21 in this disease. Finally, our control group consisted of people with suspected artery lesions, and therefore the temporal artery samples obtained were not truly normal arteries. This is due to the difficulty of obtaining normal arteries. Therefore, our conclusions remain to be further validated.

In conclusion, TCONS\_00034812 is overexpressed in atherosclerosis and upregulates miR-21 through methylation in HAOSMCs, which affects their proliferation.

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## Footnote

**Reporting Checklist:** The authors have completed the

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**Data Sharing Statement:** Available at <https://dx.doi.org/10.21037/atm-21-2632>

**Conflicts of Interest:** All authors have completed the ICMJE uniform disclosure form (available at <https://dx.doi.org/10.21037/atm-21-2632>). The authors have no conflicts of interest to declare.

**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All procedures for human sample collection performed in this study involving human participants were in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by ethics board of the Second Xiangya Hospital of Central South University, Changsha, China (No.: PJ-KY-2020-128) and informed consent was taken from all the patients. All animal experiments were performed under a project license (No.: 20201115S0310211[308]) granted animal use ethics board of the Second Xiangya Hospital of Central South University, Changsha, China, in compliance with institutional Animal Care and Use Guidelines for the care and use of animals.

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