



# Post hoc analysis of two clinical trials to compare the immunogenicity and safety of different polio immunization schedules in Chinese infants

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**Background:** A comparative analysis of the immunogenicity and safety of different poliovirus immunization schedules in Chinese infants is imperative to guide the administration of efficient strategies for the eradication of poliomyelitis.

**Methods:** A *post hoc* analysis was conducted with the data from two poliovirus vaccine clinical trials involving a combined total of 2,400 infants aged 60–90 days. Trivalent oral poliovirus vaccine (tOPV), bivalent oral poliovirus vaccine (bOPV), Sabin strain-based inactivated poliovirus vaccine (sIPV), and conventional inactivated poliovirus vaccine (cIPV) were used in different schedules, the immunogenicity and safety of which were compared 28 days after the last of three doses.

**Results:** In a per-protocol set analysis, the tOPV-tOPV-tOPV schedule induced seroconversion in 99.1%, 98.2%, and 96.0% of the inoculated infants for poliovirus type I, II, and III, respectively. The seroconversions for poliovirus types I and III were each almost 100% after immunization with the cIPV-bOPV-bOPV, sIPV-sIPV-bOPV, cIPV-cIPV-bOPV, sIPV-sIPV-tOPV, cIPV-cIPV-tOPV, or sIPV-bOPV-bOPV schedule. However, the schedules that used one IPV dose followed by two (poliovirus type I and III) bOPV doses failed to induce high-level immunity against type II poliovirus. IPV-related schedules were associated with a slightly higher incidence of adverse events (AEs).

**Conclusions:** If the capacity of IPV can be increased, two or more doses of IPV should be administered before vaccination with bOPV in a sequential schedule to improve immunity against type II poliovirus.

**Keywords:** Poliovirus sequential vaccination schedule; oral poliovirus vaccine (OPV); inactivated poliovirus vaccine (IPV); immunogenicity; safety

Submitted Mar 16, 2020. Accepted for publication Oct 12, 2020.

doi: 10.21037/atm-20-2537

View this article at: <http://dx.doi.org/10.21037/atm-20-2537>

## Introduction

The worldwide use of attenuated poliovirus vaccine (Sabin strains) has contributed greatly to the eradication of infections with wild polioviruses in children (1), but the existence of vaccine-derived polioviruses (VDPVs) and vaccine-associated paralytic poliomyelitis (VAPP) remains an obstacle in the final stage of poliomyelitis eradication (2,3). The switch from oral poliovirus vaccine (OPV) to inactivated poliovirus vaccine (IPV) was proposed as an important strategy for realizing the goal of eradicating poliomyelitis (4). However, although the implementation of this strategy has resulted in fewer cases caused by wild poliovirus strains, the existence of VDPVs still negatively impacts poliomyelitis eradication efforts (5,6). In this context, the World Health Organization (WHO) proposed a strategy of implementing a phased withdrawal of trivalent OPV (tOPV) and introducing IPV; this plan includes one or two doses of IPV to induce basic immunity against poliovirus, especially type II poliovirus, followed by one or two doses of bivalent OPV (bOPV) as a boost (2). It is designed to further decrease the number of poliomyelitis cases induced by wild poliovirus strains and reduce the likelihood of infection with VDPVs. This transitional strategy is expected to produce a suitable environment for the complete switch from immunization schedules that incorporate OPV to an IPV-only immunization schedule (7).

The experts and policy makers in China have been concerned about the immune effect of using immunization schedules that combine IPV and OPV during the switch from a tOPV schedule to an IPV-bOPV schedule. Sabin strain-based inactivated poliovirus vaccine (sIPV) was recently licensed in China in 2015, so there are fewer data about the use of sIPV in sequential immunization. Two clinical trials have been performed to investigate the efficacy and safety of sequential immunization with sIPV and bOPV in China. Because altering the poliovirus vaccination approach from using only tOPV to using a sequential schedule that combines IPV and bOPV will result in changes to immunity against poliovirus in infants, we performed a *post hoc* analysis of two clinical trials to observe these changes. The resulting data will provide guidance for the application of sequential poliovirus immunization in China.

The *post hoc* analysis performed here was designed to compare and evaluate the immunogenicity and safety of different poliovirus immunization schedules based on the results from two clinical trials. These trials both investigated the immune effects of a three-tOPV-dose schedule and of

a schedule composed of one or two IPV doses followed by two or one bOPV doses in children aged 2, 3, and 4 months. The seroconversion rates and neutralizing antibody titers in the vaccinated infants were compared to evaluate poliovirus vaccine performance during the switch from a tOPV immunization schedule to an IPV-bOPV immunization schedule in China.

## Methods

### Study design

This *post hoc* analysis included two clinical trials. One trial was conducted in Guangxi Province, China from 2011–2012 to evaluate the safety and immunogenicity of the live attenuated OPV (human diploid cell) (ClinicalTrials.gov number: NCT02231632). The other trial was carried out from 2015–2016 in Guangxi Province, China to examine the immunogenicity and safety of sequential immunization schedules of IPV+bOPV (ClinicalTrials.gov number: NCT03614702). Both trials were sponsored by the Institute of Medical Biology, Chinese Academy of Medical Sciences (IMBCAMS), and Guangxi Center for Disease Prevention and Control. Both clinical studies were conducted in accordance with the Declaration of Helsinki (as revised in 2013) and were approved by the Ethical Committee of Guangxi Zhuang Autonomous Region (approval numbers: 2009L07791 and GXIRB2015-0024-01). Informed consent was obtained for all included participants.

### Participants

The inclusion and exclusion criteria in both clinical trials were similar. The inclusion criteria were as follows: (I) infant is younger than 90 days but older than 60 days; (II) guardian provides written informed consent; (III) the infant's guardian and family follow the requirements of the clinical trial protocol; (IV) infant has no immune globulin immunization history after birth (except hepatitis B immune globulin) and no history of other live vaccination in the 28 days before vaccination; and (V) infant has an axillary temperature of <37.1 °C. The exclusion criteria were as follows: (I) infant has a personal or family history of allergy, convulsions, epilepsy, encephalopathy, or psychosis; (II) infant has an allergy to neomycin, streptomycin, or polymyxin B; (III) infant has an immunodeficiency or is receiving immunosuppressors; (IV) infant has a history of poliomyelitis; (V) infant has an acute febrile disease or infectious disease; (VI) infant experiences an abnormal stage

of labor, has a history of asphyxiation, or has a congenital malformation, developmental disorder, or severe chronic disease; (VII) infant has exhibited severe anaphylactic reactions following any previous vaccination; (VIII) infant has received oral steroids for 14 consecutive days within 1 month before the trial; (IX) infant has had a fever (axillary temperature of  $\geq 38.0$  °C) in the previous 3 days; (X) infant has had diarrhea (defecation frequency of  $\geq 3$  times/day) within the previous week; (XI) infant is participating in other clinical drug trials; and (XII) infant has any other condition that might influence the evaluation. The clinical trial that was conducted to examine the immunogenicity and safety of sequential immunization schedules composed of IPV and bOPV had one more inclusion criterion: infant had no history of immunization with an inactivated vaccine in the 14 days before vaccination.

The guardians and families of the participants voluntarily complied with the requirements of the clinical trial protocol. An informed consent form was signed by both the guardians and the study doctor of each participant prior to initiation of the clinical trial. Participants were permitted to voluntarily withdraw at any time during the trial. Participants could be withdrawn from the study in cases of failure to adhere to the follow-up visits, violation of or deviation from the trial protocol, or the appearance of other abnormal symptoms that interfered with the trial.

### Vaccines

In clinical trial NCT02231632, the administered tOPV was derived from the human embryonic lung diploid cell line KMB-17 or the monkey kidney cell line Vero (provided by IMBCAMS). The total amount of live poliovirus was no less than 6.15 lgCCID<sub>50</sub> (50% cell culture infective dose); each dose of tOPV included type I ( $\geq 6.0$  lgCCID<sub>50</sub>), type II ( $\geq 5.0$  lgCCID<sub>50</sub>), and type III ( $\geq 5.5$  lgCCID<sub>50</sub>) poliovirus.

In clinical trial NCT03614702, the sIPV was provided by IMBCAMS. Each dose (0.5 mL) contained 30, 32, and 45 D-antigen units of poliovirus types I, II, and III, respectively. The conventional inactivated poliovirus vaccine (cIPV) was purchased from Sanofi Pasteur. Each dose (0.5 mL) contained 40, 8, and 32 D-antigen units of poliovirus types I, II, and III, respectively. The bOPV was provided by IMBCAMS. Each bOPV dose contained at least 5.92 lgCCID<sub>50</sub> of live poliovirus, comprising type I ( $\geq 5.8$  lgCCID<sub>50</sub>) and type III ( $\geq 5.3$  lgCCID<sub>50</sub>) polioviruses. The tOPV was derived from the human embryonic lung diploid cell line KMB-17 as in clinical trial NCT02231632.

### Procedures

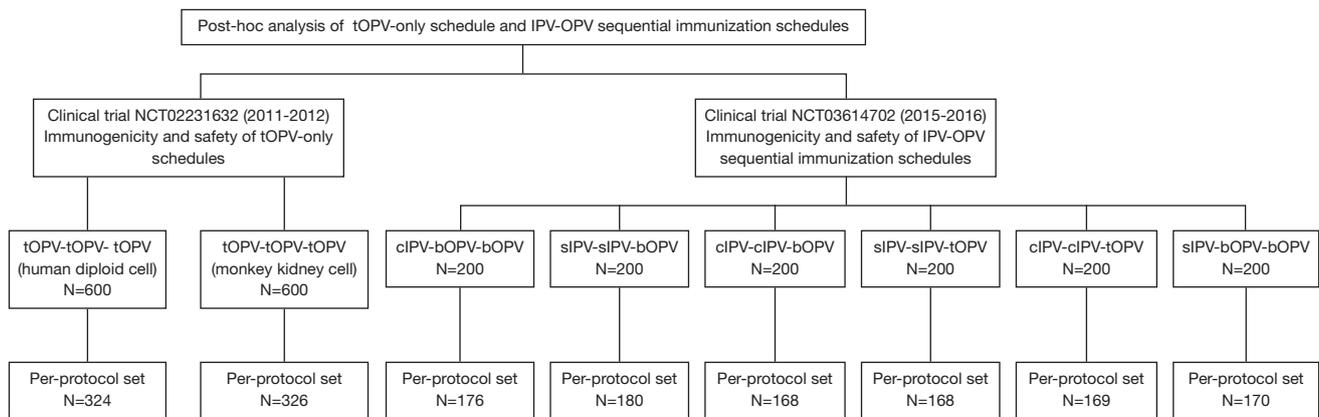
In clinical trial NCT02231632, the eligible infants (N=1,200) randomly received three doses of either the tOPV from human diploid cells or the tOPV from monkey kidney cells at the age of 2, 3, and 4 months in accordance with the routine poliovirus vaccine inoculation procedure in China.

In clinical trial NCT03614702, 1,200 participants were randomized to receive one of six different sequential poliovirus immunization schedules (sIPV-bOPV-bOPV, sIPV-sIPV-bOPV, sIPV-sIPV-tOPV, cIPV-bOPV-bOPV, cIPV-cIPV-bOPV, or cIPV-cIPV-tOPV; 200 participants per group). The IPV was administered via an intramuscular injection, whereas the OPV was administered orally. The three vaccine doses were administered at ages 2, 3, and 4 months old, respectively.

In accordance with the approved protocol, participants were observed onsite for 30 min after vaccination. In clinical trial NCT02231632, any adverse events (AEs) occurring up to 30 days after vaccination were recorded. In clinical trial NCT03614702, AEs were recorded through 28 days post-inoculation, and any serious AEs (SAEs) that occurred between 28 days and 6 months after full vaccination were followed up. Venous blood samples were collected before the first dose and at 28 days after the third dose and used by the National Institutes for Food and Drug Control to test the titer and seroconversion rate of antibodies following the protocol recommended by the WHO. Seroconversion was defined as a neutralizing antibody titer of  $<1:8$  before vaccination and of  $>1:8$  against poliovirus types I, II, and III poliovirus after three doses of vaccine. A neutralizing antibody titer of  $\geq 1:8$  before vaccination was considered to indicate maternally-transferred antibodies; for these cases, seroconversion was defined as a four-fold increase of polio-specific antibody response after three doses of vaccine. The changes in antibody titer after vaccination were analyzed. Both the maximum dilution and the maximum reported titer were 16,384; in cases where the actual titer was greater than 16,384, the value used in calculations was 16,384.

### Statistical analysis

The seroconversion rates against poliovirus types I, II, and III were calculated in terms of the poliovirus immunization schedule. The seroconversion rates and titers of antibodies against poliovirus types I, II, and III were compared between the groups of infants vaccinated under different



**Figure 1** Data flow chart of the two clinical trials included in this post hoc analysis. bOPV, bivalent (types I and III) oral polio vaccine; IPV, inactivated polio vaccine; tOPV, trivalent oral polio vaccine; sIPV; Sabin strain-based IPV; cIPV, conventional IPV, also known as Salk strain-based IPV.

schedules, and the significance of each difference was tested with a two-sided 95% confidence interval. A chi-squared test was used to compare the differences in the titer of poliovirus type II-specific antibodies between groups.

The post-vaccination geometric mean titer (GMT) and the fold changes of antibodies against poliovirus types I, II, and III in all groups are presented as the geometric means and 95% confidence intervals. The differences between groups were compared by performing an analysis of variance after logarithmic transformation of the data.

## Results

### *Baseline characteristics of the vaccinated infants*

This *post hoc* analysis was based on the data from two clinical trials conducted in Guangxi Province, China (Figure 1). The baseline characteristics of the infants in the per protocol set (PPS) analysis were similar between groups (Table 1, Table S1).

### *Seroconversion rates induced by poliovirus immunization schedules*

The seroconversion rates against poliovirus types I, II, and III were 99.1%, 98.2%, and 96.0%, respectively, in the infants who received three tOPV doses (Table 2). Notably, the positive rates of antibodies against poliovirus types I, II, and III before vaccination were 49.2%, 30.3%, and 24.0%, respectively; these positive cases are likely due to maternal antibodies (Table 1) (8). For the combined sequential immunization programs, the IPV-bOPV-bOPV

immunization program had a lower seroconversion rate against poliovirus type II than did the IPV-IPV-bOPV immunization program, regardless of whether the IPV used in the immunization program was sIPV or cIPV (Table 2, Tables S2,S3). However, the seroconversion rates for poliovirus type I and III were both almost 100% for all sequential schedule groups. The modestly higher seroconversion rates for poliovirus type III in the IPV-bOPV-bOPV groups and IPV-IPV-bOPV groups suggest that bOPV can induce a better immune response to poliovirus type III compared with tOPV because it lacks the competition from poliovirus type II. The seroconversion rates against poliovirus type II were 51.8% in the sIPV-bOPV-bOPV group and 60.2% in the cIPV-bOPV-bOPV group, suggesting that these schedules confer insufficient clinical protection against wild poliovirus or VDPV strains (Table 2). However, compared with the schedules containing one IPV dose, the vaccination schedules containing two IPV doses (sIPV or cIPV) followed by one bOPV dose were capable of eliciting a higher seroconversion rate to poliovirus type II (79.4% of vaccinated infants for sIPV-sIPV-bOPV and 81.0% for cIPV-cIPV-bOPV), suggesting that a single antigenic stimulation by the poliovirus type II vaccine is unable to elicit adequate immune protection. Thus, two or more IPV doses should be required, especially for infants at a high risk of infection with type II VDPV.

### *Neutralizing antibody titers*

The effectiveness of the immunologic barrier against

**Table 1** Baseline characteristics of the vaccinated infants in terms of polio immunization schedule in per protocol set in 2012 and 2016

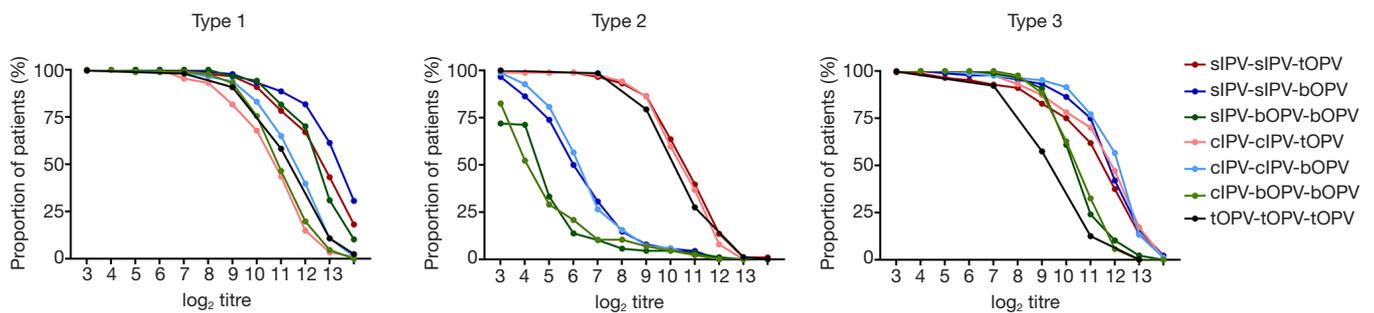
Parameter	tOPV-tOPV-tOPV (N=650) <sup>†</sup>	cIPV-bOPV-bOPV (N=176)	sIPV-sIPV-bOPV (N=180)	cIPV-cIPV-bOPV (N=168)	sIPV-sIPV-tOPV (N=168)	cIPV-cIPV-tOPV (N=169)	sIPV-bOPV-bOPV (N=170)	P value
Age (days), mean ± SD	72.5±8.7	70.5±7.8	70.4±7.6	70.6±7.7	71.7±8.0	70.3±7.5	70.5±7.3	
Sex, n (%)								
Male	380 (58.5)	88 (50.0)	97 (53.9)	98 (58.3)	83 (49.4)	89 (52.7)	94 (55.3)	1.00
Female	270 (41.5)	88 (50.0)	83 (46.1)	70 (41.7)	85 (50.6)	80 (47.3)	76 (44.7)	1.00
Type 1 polio strain								
Seropositive, n (%; 95% CI)	320 (49.2, 45.4–53.1)	78 (44.3, 36.9–52.0)	69 (38.3, 31.2–45.9)	73 (43.5, 35.8–51.3)	69 (41.1, 33.6–48.9)	73 (43.2, 35.6–51.0)	73 (42.9, 35.4–50.7)	0.14
GMT, geometric mean (95% CI)	9.8 (9.0–10.7)	8.3 (7.2–9.6)	8.1 (6.1–6.6)	8.4 (7.1–10.0)	8.7 (7.2–10.4)	8.5 (7.3–9.9)	8.4 (7.2–9.9)	0.23
Type 2 polio strain								
Seropositive, n (%; 95% CI)	197 (30.3, 26.8–33.8)	64 (36.4, 29.3–43.9)	64 (35.6, 28.6–43.0)	62 (36.9, 29.6–44.7)	59 (35.1, 27.9–42.9)	56 (33.1, 26.1–40.8)	54 (31.8, 24.8–39.3)	1.00
GMT, geometric mean (95% CI)	6.6 (6.2–7.1)	7.3 (6.3–8.4)	7.2 (6.2–8.3)	7.1 (6.2–8.1)	7.0 (6.0–8.0)	6.4 (5.7–7.3)	7.1 (6.1–8.2)	0.76
Type 3 polio strain								
Seropositive, n (%; 95% CI)	156 (24.0, 20.7–27.3)	39 (22.2, 16.3–29.0)	33 (18.3, 13.0–24.8)	27 (16.1, 10.9–22.5)	29 (17.3, 11.9–23.8)	34 (20.1, 14.4–27.0)	30 (17.7, 12.2–24.2)	1.00
GMT, geometric mean (95% CI)	5.9 (5.5–6.2)	5.5 (4.9–6.1)	5.1 (4.7–5.6)	5.4 (4.8–6.1)	5.3 (4.8–5.9)	5.4 (4.9–6.0)	5.4 (4.8–6.2)	0.38

The sequential schedules of cIPV-bOPV-bOPV, sIPV-sIPV-bOPV, cIPV-cIPV-bOPV, sIPV-sIPV-tOPV, and sIPV-bOPV-bOPV were compared in a clinical trial conducted between 2015 and 2016 in China. <sup>†</sup>, pooled data for the tOPVs derived either from human diploid cell or monkey kidney cell in the clinical trial conducted between 2011 and 2012 in China. tOPV, trivalent oral polio vaccine; cIPV, conventional inactivated poliovirus vaccine; bOPV, bivalent oral polio vaccine; sIPV, Sabin strain-based inactivated poliovirus vaccine; SD, standard deviation; CI, confidence interval; GMT, geometric mean titer.

**Table 2** Seroconversion rate and geometric mean titer against poliovirus types I, II, and III 28 days after three doses of different immunization schedules in per protocol set analysis in 2012 and 2016

Immunogenicity indicator	TOPV-tOPV-tOPV (N=650) <sup>†</sup>	cIPV-bOPV-bOPV (N=176)	sIPV-sIPV-bOPV (N=180)	cIPV-cIPV-bOPV (N=168)	sIPV-sIPV-tOPV (N=168)	cIPV-cIPV-tOPV (N=169)	sIPV-bOPV-bOPV (N=170)	P value
<b>Type 1 polio strain</b>								
Seropositive, n (%), 95% CI)	644 (99.1, 98.3-99.8)	174 (98.9, 96.0-99.9)	178 (98.9, 90.0-99.9)	166 (98.8, 95.8-99.9)	165 (98.2, 94.9-99.6)	165 (97.6, 94.1-99.4)	169 (99.4, 96.8-100.0)	0.72
GMT, geometric mean (95% CI)	4,108 (3,788.0-4,454.0)	2,066.1 (1,777.9-2,400.9)	7,359.6 (6,379.2-8,490.6)	2,330.4 (1,940.2-2,799.0)	4,316.7 (3,640.0-5,119.3)	1,294.8 (1,073.2-1,562.2)	5,303.9 (4,610.5-6,101.5)	<0.001
<b>Type 2 polio strain</b>								
Seropositive, n (%), 95% CI)	638 (98.2, 97.1-99.2)	106 (60.2, 52.6-67.5)	143 (79.4, 72.8-85.1)	136 (81.0, 74.2-86.6)	166 (98.8, 95.8-99.9)	167 (98.8, 95.8-99.9)	88 (51.8, 44.0-59.5)	<0.001
GMT, geometric mean (95% CI)	609.4 (565.3-657)	18.2 (14.5-22.7)	73.4 (60.1-89.7)	68.0 (56.7-81.4)	1,446.5 (1,239.5-1,688.1)	1,219.7 (1,029.9-1,430.7)	11.7 (9.8-14.0)	<0.001
<b>Type 3 polio strain</b>								
Seropositive, n (%), 95% CI)	624 (96.0, 94.5-97.5)	175 (99.4, 96.9-100.0)	179 (99.4, 96.9-100.0)	166 (98.8, 95.8-99.9)	164 (97.6, 94.0-99.4)	167 (98.8, 95.8-99.9)	169 (99.4, 96.8-100.0)	<0.05
GMT, geometric mean (95% CI)	486.1 (440.3-536.7)	1,274.9 (1,132.3-1,435.5)	2,941.1 (2,493.7-3,468.9)	2,783.4 (2,345.7-3,302.8)	1,771.3 (1,396.4-2,246.7)	1,716.6 (1,359.0-2,168.3)	1,285.7 (1,124.2-1,470.4)	<0.001

The sequential schedules of cIPV-bOPV-bOPV, sIPV-sIPV-bOPV, cIPV-cIPV-tOPV, sIPV-sIPV-tOPV, and sIPV-bOPV-bOPV were compared in a clinical trial conducted between 2015 and 2016 in China. <sup>†</sup>, pooled data for the TOPVs derived either from human diploid cell or monkey kidney cell from the clinical trial conducted between 2011 and 2012 in China. TOPV, trivalent oral polio vaccine; cIPV, conventional inactivated poliovirus vaccine; bOPV, bivalent oral polio vaccine; sIPV, Sabin strain-based inactivated poliovirus vaccine; CI, confidence interval; GMT, geometric mean titer.



**Figure 2** Reverse cumulative distribution curves showing the titers of serum antibodies neutralizing poliovirus types I, II and III.

infection by wild poliovirus or VDPV strains largely depends on the level of neutralizing antibodies in the sera of an individual (9,10). We observed high GMTs for all three types of poliovirus after immunization with tOPV-tOPV-tOPV, which is consistent with a previous report (11). The IPV-bOPV-bOPV and IPV-IPV-bOPV schedules, regardless of whether sIPV or cIPV was used, were capable of eliciting high GMTs against poliovirus types I and III (Table 2, Figure 2). However, the IPV-bOPV-bOPV schedule induced a lower level of antibody response against type II poliovirus, regardless of whether sIPV or cIPV was used. Although the IPV-IPV-bOPV group exhibited a relatively high level of antibodies against type II poliovirus, this level was still lower than those in the IPV-IPV-tOPV groups (Table 2, Table S4, Figure 2).

### Safety evaluation

The safety of the tOPV-tOPV-tOPV schedule has been demonstrated over its >50 years of use in Chinese children (12); the primary safety concern for this approach is the rare cases of VAPP (6,13). As part of new poliovirus eradication strategy, the final goal of which is to adopt an IPV-only immunization schedule and eradicate all wild polioviruses and VDPVs, a switch from tOPV to bOPV is necessary, and the safety of this change must be evaluated. One of the included clinical trials reported a total of 951 AEs, and the other reported 918 AEs (Table 3). The incidences of fever were 28.0% (335/1,195) in the tOPV-tOPV-tOPV schedule and 49.9% (598/1,198) for all sequential schedules combined. Additionally, the incidences of abnormal irritability were 5.9% and 20.9%, the incidences of local reactions (e.g., pain, redness, swelling, and induration) were 0% and 1.3%, and the incidences of gastrointestinal symptoms were 10.0% and 6.7% in the

tOPV-tOPV-tOPV schedule and for all sequential schedules combined, respectively. SAEs were reported in 5.9% (71/1,195) of participants in the tOPV-tOPV-tOPV group and in 5.0% (60/1,198) of participants who were vaccinated with a sequential schedule (Table 3, Table S5). The injected IPV was associated with a relatively high incidence of AEs, even though most of the reported AEs were identified as being unrelated to inoculation with poliovirus vaccine.

### Discussion

The sequential removal of Sabin poliovirus strains leading up to a full replacement of OPV with IPV is a goal established by the Global Polio Eradication Initiative [2015] for the eradication of poliomyelitis caused by all wild polioviruses and VDPVs in children worldwide. As the first step towards this goal, the strategy of using sequential immunization schedules combining bOPV with IPV by removing Sabin type II should maintain an adequate immune protective effect against any possible infection by wild poliovirus strains or VDPVs (14).

From 2011 to 2012, we conducted a clinical trial to evaluate the safety and immunogenicity of the live attenuated OPV (human diploid cell; ClinicalTrials.gov number: NCT02231632). Vaccination with three doses of tOPV administered at 2, 3, and 4 months of age, respectively, was the immunization strategy against poliomyelitis in mainland China until April 2016 (15). Since then, global immunization strategies for poliovirus have changed. Clinical trial NCT03614702 was carried out from 2015 to 2016 in Guangxi Province, China to examine the immunogenicity and safety of sequential immunization schedules with sIPV+bOPV. The present study is a comprehensive analysis of these two clinical trials in which the changes induced by switching the immunization strategy

**Table 3** Incidence of adverse events in the infants after immunization with different polio vaccination schedules in safety set analysis

Adverse event	TOPV-tOPV-tOPV (N=1,195) <sup>†</sup>	cIPV-bOPV-bOPV (N=199)	sIPV-sIPV-bOPV (N=199)	cIPV-cIPV-bOPV (N=200)	sIPV-sIPV-tOPV (N=200)	cIPV-cIPV-tOPV (N=200)	sIPV-bOPV-bOPV (N=200)	P value
Any adverse event	951 (79.6)	150 (75.0)	153 (79.8)	148 (73.0)	156 (80.0)	156 (78.0)	155 (79.0)	0.60
Serious adverse events	71 (5.9)	10 (5.0)	10 (5.0)	11 (5.5)	9 (4.5)	9 (4.5)	11 (5.5)	0.97
Unsolicited adverse events	835 (69.9)	87 (43.7)	94 (47.2)	90 (45.0)	96 (48.0)	97 (48.5)	91 (45.5)	<0.001
Solicited adverse events	468 (39.2)	129 (64.8)	131 (65.8)	122 (61.0)	119 (59.5)	127 (63.5)	133 (66.5)	<0.001
Fever	335 (28.0)	106 (53.3)	103 (51.8)	93 (46.5)	93 (46.5)	96 (48.0)	107 (53.5)	<0.001
Abnormal irritability	71 (5.9)	39 (19.6)	45 (22.6)	31 (15.5)	43 (21.5)	49 (24.5)	43 (21.5)	<0.001
Drowsiness	11 (0.9)	6 (3.0)	10 (5.0)	8 (4.0)	17 (8.5)	10 (5.0)	16 (8.0)	<0.001
Gastrointestinal symptoms <sup>‡</sup>	120 (10.0)	12 (6.0)	17 (8.5)	14 (8.5)	8 (7.0)	10 (4.0)	19 (5.0)	<0.05
Diarrhea	135 (11.3)	24 (12.1)	31 (15.6)	22 (11.0)	27 (13.5)	20 (10.0)	21 (10.5)	0.59
Allergy	17 (1.4)	8 (4.0)	3 (1.5)	5 (2.5)	8 (4.0)	3 (1.5)	4 (2.0)	0.09
Local reactions <sup>§</sup>	0	1 (0.5)	2 (1.0)	1 (0.5)	4 (2.0)	3 (1.5)	5 (2.5)	<0.001

The data are expressed as n (%). <sup>†</sup>, pooled data for the tOPVs derived either from human diploid cell or monkey kidney cell from the clinical trial conducted between 2011 and 2012 in China. <sup>‡</sup>, including nausea, vomiting, and anorexia. <sup>§</sup>, including pain, redness, swelling, and induration. tOPV, trivalent oral polio vaccine; cIPV, conventional inactivated poliovirus vaccine; bOPV, bivalent oral polio vaccine; sIPV, Sabin strain-based inactivated poliovirus vaccine. The sequential schedules of cIPV-bOPV-bOPV, sIPV-sIPV-bOPV, cIPV-cIPV-bOPV, sIPV-sIPV-tOPV, cIPV-cIPV-tOPV, and sIPV-bOPV-bOPV were compared in a clinical trial conducted between 2015 and 2016 in China.

from only tOPV to IPV+bOPV can be observed. Although some published reports describe other clinical trials on sequential immunization with IPV and OPV, most of the previous studies on sequential poliovirus immunization used conventional IPV and did not incorporate the use of sIPV. Our study supplements the existing data by addressing the use of sIPV in sequential immunization.

This *post hoc* analysis compared the immune effects induced by a tOPV-tOPV-tOPV schedule with those induced by various sequential schedules using IPV (sIPV or cIPV) followed by bOPV. Vaccine-induced immunity to poliovirus must be maintained at an adequate level in Chinese children during the transition from IPV-bOPV combination schedules to the complete cessation of OPV administration and the sustained use of an IPV-only schedule. Our results confirm that, similar to the tOPV-tOPV-tOPV schedule, schedules containing one or two IPV doses (sIPV or cIPV) followed by one or two doses of bOPV could induce high seroconversion rates against poliovirus type I and III, associated with high neutralizing antibody titers. However, the administration of one IPV dose followed by two doses of bOPV failed to induce an adequate level of seroconversion or neutralizing antibody titer against type II poliovirus. Thus, the application of a schedule with a single IPV dose during the transition period may lead to an increased risk of infection with type II circulating vaccine-derived poliovirus (cVDPV). In contrast, the administration of two IPV doses followed by one dose of bOPV elicit higher levels of seroconversion and antibody titers against type II poliovirus.

There are still pandemics of infection with wild poliovirus strains in at least two countries that border China (5,16). To improve immunity against type II poliovirus, two or more doses of IPV should be administered before the administration of bOPV in a sequential schedule. A previous study of IPV also reported that one dose of IPV induced seroconversion rates of 19–46%, 32–63%, and 28–54% for poliovirus types I, II, and III, respectively (17). A boost with poliovirus vaccine is important for the establishment of effective immunity against poliovirus (18,19). Importantly, all these data suggest that a schedule using one dose of IPV followed by two doses of bOPV is not suitable for poliovirus immunization during the transition period, especially in developing countries where tOPV had been used alone for a long time and where there is still a risk of the spread of wild poliovirus strains and VDPVs (20,21). At this stage, using at least two doses of IPV will be a better option for a routine poliovirus immunization schedule to maintain an adequate

level of population immunity to all poliovirus types and avoid the emergence of epidemic cases of paralysis induced by wild poliovirus strains or VDPVs.

The safety evaluation results suggest that sequential immunization schedules combining OPV with IPV are associated with a higher incidence of some AEs, such as fever and local reactions, which may be due to the inclusion of one or two injections in these schedules. Although both included clinical trials were completed in the same region, they were not conducted during the same period, and there were also some small differences in their methods of AE collection, so the AE differences between poliovirus vaccination schedules may be caused by factors other than the immunization procedure.

In conclusion, during the transition from a tOPV-only immunization schedule to an IPV-bOPV immunization schedule, sequential schedules containing two IPV doses are a reasonable option that align with current WHO recommendations. Our findings may help decision-makers in developing countries to better optimize poliovirus vaccination policies.

## Acknowledgments

We are very grateful to the staff of the Centers for Disease Control and Prevention of Guangxi Province and Liujiang City and all the onsite sub-investigators for their contributions. We are also indebted to the staff of National Institutes for Food and Drug Control for providing the serum neutralization antibody assay, to Simurike Pharmaceutical Information Consulting Co., Ltd. for supervising the study and providing data management services, and to the experts from the Center for Drug Evaluation, China Food and Drug Administration and National Center for Disease Control and Prevention for their valuable opinions and suggestions. We thank Katie Oakley, PhD, from Liwen Bianji, Edanz Editing China ([www.liwenbianji.cn/ac](http://www.liwenbianji.cn/ac)), for editing the English text of a draft of this manuscript.

*Funding:* This work was supported by the State Project for Essential Drug Research and Development (2015ZX09101031) and Yunnan Province Project (2016BC002).

## Footnote

*Data Sharing Statement:* Available at <http://dx.doi.org/10.21037/atm-20-2537>

*Peer Review File:* Available at <http://dx.doi.org/10.21037/atm-20-2537>

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/atm-20-2537>). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The two included clinical studies were each conducted in accordance with the Declaration of Helsinki (as revised in 2013). These two clinical studies were approved by the Ethical Committee of Guangxi Zhuang Autonomous Region (approval numbers: 2009L07791 and GXIRB2015-0024-01), and informed consent was obtained for all the participants.

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## References

1. Thompson KM, Tebbens RJ. Current polio global eradication and control policy options: perspectives from modeling and prerequisites for oral poliovirus vaccine cessation. *Expert Rev Vaccines* 2012;11:449-59.
2. Patel M, Zipursky S, Orenstein W, et al. Polio endgame: the global introduction of inactivated polio vaccine. *Expert Rev Vaccines* 2015;14:749-62.
3. Duintjer Tebbens RJ, Pallansch MA, Kim JH, et al. Oral poliovirus vaccine evolution and insights relevant to modeling the risks of circulating vaccine-derived polioviruses (cVDPVs). *Risk Anal* 2013;33:680-702.
4. Garon J, Seib K, Orenstein WA, et al. Polio endgame: the global switch from tOPV to bOPV. *Expert Rev Vaccines* 2016;15:693-708.
5. Initiative GPE. List of wild poliovirus by country and year. 2019. Available online: <http://www.polioeradication.org/Dataandmonitoring/Poliothisweek/Wildpolioviruslist.aspx>. Accessed 1 Jul 2019.
6. Wang HB, Luo HM, Li L, et al. Vaccine-derived poliovirus surveillance in China during 2001-2013: the potential challenge for maintaining polio free status. *BMC Infect Dis* 2017;17:742.
7. Initiative GPE. Polio Eradication & Endgame Strategic Plan 2013-2018. 2016. Available online: [http://www.polioeradication.org/Portals/0/Document/Resources/StrategyWork/PEESP\\_EN\\_US.pdf](http://www.polioeradication.org/Portals/0/Document/Resources/StrategyWork/PEESP_EN_US.pdf). Accessed 12 Jul 2019.
8. Gaensbauer JT, Gast C, Bandyopadhyay AS, et al. Impact of Maternal Antibody on the Immunogenicity of Inactivated Polio Vaccine in Infants Immunized With Bivalent Oral Polio Vaccine: Implications for the Polio Eradication Endgame. *Clin Infect Dis* 2018;67:S57-S65.
9. Bandyopadhyay AS, Garon J, Seib K, et al. Polio vaccination: past, present and future. *Future Microbiol* 2015;10:791-808.
10. Norrby E, Uhnöo I, Brytting M, et al. Polio close to eradication. *Lakartidningen* 2017;114:EPDT.
11. Lu L, Li X, Zhang H, et al. Immunogenicity and persistence from different 3-dose schedules of live and inactivated polio vaccines in Chinese infants. *Vaccine* 2015;33:4653-8.
12. Yu WZ, Wen N, Zhang Y, et al. Poliomyelitis eradication in China: 1953-2012. *J Infect Dis* 2014;210 Suppl 1:S268-74.
13. Yan D, Zhang Y, Zhu S, et al. Limited and localized outbreak of newly emergent type 2 vaccine-derived poliovirus in Sichuan, China. *Clin Vaccine Immunol* 2014;21:1012-8.
14. Platt LR, Estivariz CF, Sutter RW. Vaccine-associated paralytic poliomyelitis: a review of the epidemiology and estimation of the global burden. *J Infect Dis* 2014;210 Suppl 1:S380-9.
15. Prevention CCfDCa. Decision to Implement New Polio Vaccine Immunization Strategy. 2016. Available online: <http://www.chinacdc.cn/jkzt/crb/zl/jshzy/>. Accessed 5 Jul 2019.
16. Yan D, Wang D, Zhu S, et al. Immunogenicity of oral polio vaccine and Salk inactive polio vaccine against Xinjiang imported type 1 wild poliovirus. *Clin Infect Dis* 2020;70:1980-4.
17. Estivariz CF, Pallansch MA, Anand A, et al. Poliovirus vaccination options for achieving eradication and securing the endgame. *Curr Opin Virol* 2013;3:309-15.
18. O'Ryan M, Bandyopadhyay AS, Villena R, et al. Inactivated poliovirus vaccine given alone or in a sequential schedule with bivalent oral poliovirus vaccine in Chilean infants: a randomised, controlled, open-label, phase 4, non-

- inferiority study. *Lancet Infect Dis* 2015;15:1273-82.
19. Taniuchi M, Famulare M, Zaman K, et al. Community transmission of type 2 poliovirus after cessation of trivalent oral polio vaccine in Bangladesh: an open-label cluster-randomised trial and modelling study. *Lancet Infect Dis* 2017;17:1069-79.
  20. Lewis I, Ottosen A, Rubin J, et al. A Supply and Demand Management Perspective on the Accelerated Global Introductions of Inactivated Poliovirus Vaccine in a Constrained Supply Market. *J Infect Dis* 2017;216:S33-9.
  21. Duintjer Tebbens RJ, Hampton LM, Thompson KM. Implementation of coordinated global serotype 2 oral poliovirus vaccine cessation: risks of potential non-synchronous cessation. *BMC Infect Dis* 2016;16:231.

**Cite this article as:** Zhao T, Mo Z, Ying Z, Huang T, Che Y, Li G, Yang X, Sun M, Jiang L, Shi L, Ye H, Zhao Z, Liu X, Li J, Li Y, Li R, Jiang R, Wang J, Fu Y, Ma R, Shi H, Yang H, Li C, Yang J, Li Q. *Post hoc* analysis of two clinical trials to compare the immunogenicity and safety of different polio immunization schedules in Chinese infants. *Ann Transl Med* 2021;9(3):253. doi: 10.21037/atm-20-2537

**Table S1** Statistical analysis of baseline characteristics of the vaccinated infants in terms of polio immunization schedule in per protocol set

Parameter	Test method	P value
Age (days), mean $\pm$ SD		
Sex, n (%)		
Male	Pearson CHI-SQUARE	1.00
Female	Pearson CHI-SQUARE	1.00
Type 1 polio strain		
Seropositive, n (% , 95% CI)	Pearson CHI-SQUARE	0.14
GMT, geometric mean (95% CI)	ANOVA	0.23
Type 2 polio strain		
Seropositive, n (% , 95% CI)	Pearson CHI-SQUARE	1.00
GMT, geometric mean (95% CI)	ANOVA	0.76
Type 3 polio strain		
Seropositive, n (% , 95% CI)	Pearson CHI-SQUARE	1.00
GMT, geometric mean (95% CI)	ANOVA	0.38

SD, standard deviation; CI, confidence interval; GMT, geometric mean titer; ANOVA, analysis of variance.

**Table S2** Statistical analysis of seroconversion rate and geometric mean titer against poliovirus (types 1, 2, and 3) 28 days after three doses of different immunization schedules in per protocol set analysis

Immunogenicity indicator	Test method	P value
Type 1 polio strain		
Seropositive, n (% , 95% CI)	Fisher exact test	0.72
GMT, geometric mean (95% CI)	ANOVA	<0.001
Type 2 polio strain		
Seropositive, n (% , 95% CI)	Pearson CHI-SQUARE	<0.001
GMT, geometric mean (95% CI)	ANOVA	<0.001
Type 3 polio strain		
Seropositive, n (% , 95% CI)	Fisher exact test	<0.05
GMT, geometric mean (95% CI)	ANOVA	<0.001

CI, confidence interval; GMT, geometric mean titer; ANOVA, analysis of variance.

**Table S3** Multiple comparisons of seroconversion rate against poliovirus (types 1, 2, and 3) 28 days after three doses of different immunization schedules

Immunogenicity indicator	Group	vs.	Group	Test method	P value
Type2 polio strain	cIPV-bOPV-bOPV	vs.	sIPV-sIPV-tOPV	Pearson CHI-SQUARE	<0.001
			sIPV-sIPV-bOPV	Pearson CHI-SQUARE	<0.001
			cIPV-cIPV-tOPV	Pearson CHI-SQUARE	<0.001
			cIPV-cIPV-bOPV	Pearson CHI-SQUARE	<0.001
			sIPV-bOPV- bOPV	Pearson CHI-SQUARE	0.13
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.001
	sIPV-sIPV-tOPV	vs.	sIPV-sIPV-bOPV	Pearson CHI-SQUARE	<0.001
			cIPV-cIPV-tOPV	Fisher exact test	1.00
			cIPV-cIPV-bOPV	Pearson CHI-SQUARE	<0.001
			sIPV-bOPV- bOPV	Pearson CHI-SQUARE	<0.001
			tOPV-tOPV-tOPV	Fisher exact test	0.75
	sIPV-sIPV-bOPV	vs.	cIPV-cIPV-tOPV	Pearson CHI-SQUARE	<0.001
			cIPV-cIPV-bOPV	Pearson CHI-SQUARE	0.79
			sIPV-bOPV- bOPV	Pearson CHI-SQUARE	<0.001
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.001
	cIPV-cIPV-tOPV	vs.	cIPV-cIPV-bOPV	Pearson CHI-SQUARE	<0.001
			sIPV-bOPV- bOPV	Pearson CHI-SQUARE	<0.001
			tOPV-tOPV-tOPV	Fisher exact test	0.75
	cIPV-cIPV-bOPV	vs.	sIPV-bOPV- bOPV	Pearson CHI-SQUARE	<0.001
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.001
sIPV-bOPV- bOPV	vs.	tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.001	
Type3 polio strain	cIPV-bOPV-bOPV	vs.	sIPV-sIPV-tOPV	Fisher exact test	0.21
			sIPV-sIPV-bOPV	Fisher exact test	1
			cIPV-cIPV-tOPV	Fisher exact test	0.62
			cIPV-cIPV-bOPV	Fisher exact test	0.62
			sIPV-bOPV- bOPV	Pearson CHI-SQUARE	1
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.05
	sIPV-sIPV-tOPV	vs.	sIPV-sIPV-bOPV	Fisher exact test	0.20
			cIPV-cIPV-tOPV	Fisher exact test	0.45
			cIPV-cIPV-bOPV	Fisher exact test	0.69
			sIPV-bOPV- bOPV	Fisher exact test	0.21
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	0.49
	sIPV-sIPV-bOPV	vs.	cIPV-cIPV-tOPV	Fisher exact test	0.61
			cIPV-cIPV-bOPV	Fisher exact test	0.61
			sIPV-bOPV- bOPV	Fisher exact test	1
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.05
	cIPV-cIPV-tOPV	vs.	cIPV-cIPV-bOPV	Fisher exact test	1
			sIPV-bOPV- bOPV	Fisher exact test	0.62
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	0.09
	cIPV-cIPV-bOPV	vs.	sIPV-bOPV- bOPV	Fisher exact test	0.62
			tOPV-tOPV-tOPV	Pearson CHI-SQUARE	0.09
sIPV-bOPV- bOPV	vs.	tOPV-tOPV-tOPV	Pearson CHI-SQUARE	<0.05	

tOPV, trivalent oral polio vaccine; cIPV, conventional inactivated poliovirus vaccine; bOPV, bivalent oral polio vaccine; sIPV, Sabin strain-based inactivated poliovirus vaccine.

**Table S4** Multiple comparisons of antibody geometric mean titer against poliovirus (types 1, 2, and 3) 28 days after three doses of different immunization schedules

Immunogenicity indicator	Group	vs.	Group	Test method	P value
Type1 polio strain	cIPV-bOPV-bOPV	vs.	sIPV-sIPV-bOPV	T test	<0.001
			cIPV-cIPV-bOPV	T test	0.30
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
	sIPV-sIPV-bOPV	vs.	cIPV-cIPV-bOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.01
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
	cIPV-cIPV-bOPV	vs.	sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
	sIPV-sIPV-tOPV	vs.	cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	0.08
			tOPV-tOPV-tOPV	T test	0.59
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
cIPV-cIPV-tOPV	vs.	sIPV-bOPV- bOPV	T test	<0.001	
		tOPV-tOPV-tOPV	T test	<0.001	
		sIPV-sIPV-tOPV	T test	<0.001	
		cIPV-cIPV-tOPV	T test	<0.001	
		sIPV-bOPV- bOPV	T test	<0.01	
		tOPV-tOPV-tOPV	T test	<0.01	
Type2 polio strain	cIPV-bOPV-bOPV	vs.	sIPV-sIPV-bOPV	T test	<0.001
			cIPV-cIPV-bOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
	sIPV-sIPV-bOPV	vs.	cIPV-cIPV-bOPV	T test	0.53
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
	cIPV-cIPV-bOPV	vs.	sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	0.17
			cIPV-cIPV-tOPV	T test	<0.001
	sIPV-sIPV-tOPV	vs.	sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
cIPV-cIPV-tOPV	vs.	sIPV-bOPV- bOPV	T test	<0.001	
		tOPV-tOPV-tOPV	T test	<0.001	
		sIPV-sIPV-tOPV	T test	<0.001	
		cIPV-cIPV-tOPV	T test	<0.001	
		sIPV-bOPV- bOPV	T test	<0.001	
		tOPV-tOPV-tOPV	T test	<0.001	
Type3 polio strain	cIPV-bOPV-bOPV	vs.	sIPV-sIPV-bOPV	T test	<0.001
			cIPV-cIPV-bOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.05
			cIPV-cIPV-tOPV	T test	<0.05
			sIPV-bOPV- bOPV	T test	0.95
			tOPV-tOPV-tOPV	T test	<0.001
	sIPV-sIPV-bOPV	vs.	cIPV-cIPV-bOPV	T test	0.68
			sIPV-sIPV-tOPV	T test	<0.001
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.001
	cIPV-cIPV-bOPV	vs.	sIPV-sIPV-tOPV	T test	<0.01
			cIPV-cIPV-tOPV	T test	<0.001
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	0.82
			cIPV-cIPV-tOPV	T test	<0.001
	sIPV-sIPV-tOPV	vs.	sIPV-bOPV- bOPV	T test	<0.05
			tOPV-tOPV-tOPV	T test	<0.001
			sIPV-sIPV-tOPV	T test	<0.05
			cIPV-cIPV-tOPV	T test	<0.05
			sIPV-bOPV- bOPV	T test	<0.001
			tOPV-tOPV-tOPV	T test	<0.001
cIPV-cIPV-tOPV	vs.	sIPV-bOPV- bOPV	T test	<0.05	
		tOPV-tOPV-tOPV	T test	<0.001	
		sIPV-sIPV-tOPV	T test	<0.001	
		cIPV-cIPV-tOPV	T test	<0.001	
		sIPV-bOPV- bOPV	T test	<0.001	
		tOPV-tOPV-tOPV	T test	<0.001	
sIPV-bOPV- bOPV	vs.	tOPV-tOPV-tOPV	T test	<0.001	

tOPV, trivalent oral polio vaccine; cIPV, conventional inactivated poliovirus vaccine; bOPV, bivalent oral polio vaccine; sIPV, Sabin strain-based inactivated poliovirus vaccine.

**Table S5** Statistical analysis of incidence of adverse events in the infants after immunization with different polio vaccination schedules in safety set analysis

Adverse event	Test method	P value
Any adverse event	Pearson CHI-SQUARE	0.60
Serious adverse events	Pearson CHI-SQUARE	0.97
Unsolicited adverse events	Pearson CHI-SQUARE	<0.001
Solicited adverse events	Pearson CHI-SQUARE	<0.001
Fever	Pearson CHI-SQUARE	<0.001
Abnormal irritability	Pearson CHI-SQUARE	<0.001
Drowsiness	Pearson CHI-SQUARE	<0.001
Gastrointestinal symptoms	Pearson CHI-SQUARE	<0.05
Diarrhea	Pearson CHI-SQUARE	0.59
Allergy	Fisher exact test	0.09
Local reactions <sup>c</sup>	Pearson CHI-SQUARE	<0.001

<sup>c</sup>, including pain, redness, swelling, and induration.