Current practices on genetic testing in ovarian cancer

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Abstract: Epithelial ovarian cancer (EOC) is probably the tumor type with the highest percentage of hereditary cases observed, irrespectively of selection criteria. A fourth to a fifth of unselected epithelial EOC patients carry pathogenic variants (PVs) in a number of genes, the majority of which encode for proteins involved in DNA repair pathways. BRCA1 and BRCA2 predisposing PVs were the first to be associated to ovarian cancer, with the advent in DNA sequencing technologies leading to the discovery and association of additional genes which compromise the homologous recombination (HR) pathway. In addition, PVs genes involved in mismatch repair (MMR) pathway, account for 10–15% of hereditary EOC. The identification of women with HR deficient ovarian cancers has significant clinical implications concerning chemotherapy regimen planning and development and use of targeted therapies as well. More specifically, in patients with BRCA1/2 PVs or HR deficiency maintenance treatment with poly(ADP-ribose) polymerase (PARP) inhibitors, either in the first line setting or in recurrent disease, improves the progression-free survival. But also patients with HR proficient tumors show a benefit. Therefore, genetic testing in ovarian cancer has a prognostic and predictive value. In this review, we discuss which ovarian cancer patients should be referred for genetic counseling and how to perform genetic testing. We also discuss the timing of genetic testing and its clinical relevance to BRCA status.

Keywords: Genetic testing, BRCA mutations, homologous recombination hereditary ovarian cancer, poly(ADP-ribose) polymerase inhibitors (PARP inhibitors)

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Introduction

Epithelial ovarian cancer (EOC) is the most lethal gynecological cancer. It represents the seventh most commonly diagnosed cancer among women in the world with a 5-year survival rate of 46%. Annually worldwide, 295,000 women will be diagnosed and 184,000 will die. In 2018 in the United States, there were approximately 22,240 new cases of ovarian cancer diagnosed and 14,070 ovarian cancer deaths (1-3). One of the main factors contributing to the high death-to-incidence rate is the advanced stage of the disease at the time of diagnosis. Therefore, improving prevention and early detection is a research priority because disease diagnosed at an early stage has a 5-year relative survival rate that exceeds 90% (4,5). EOC encompasses a heterogenous group of malignancies that vary in etiology, molecular biology, and numerous other characteristics and is divided according to histologic subtypes: high-grade serous (the most frequent), low-grade serous, clear cell, endometrioid, and mucinous (6). Each histologic subtype is associated with a distinct clinical behavior (response to chemotherapy, pattern of metastases, survival) but has historically been treated as one entity (6-9).

The majority of hereditary ovarian cancer can be
attributed to germline BRCA1 & BRCA2 pathogenic variants. A number of additional variants, in genes beyond BRCA1/2, have been identified and are suspected to play a significant role in ovarian carcinogenesis. As the complexity of genetic testing increases, the evolution of next generation sequencing (NGS) allows rapid evaluation of variants in multiple cancer susceptibility genes at similar costs to single gene sequencing.

**Molecular landscape**

**Homologous recombination (HR) repair pathway**

DNA double-strand breaks (DSBs) are one of the most common and most cytotoxic types of DNA damage that can consequently result in significant genomic aberrations, which if left unrepaired or improperly repaired, will ultimately result in cell death (10). In humans, DSBs are repaired by a number of repair pathways, the most important of which involve non-homologous end joining (NHEJ) and HR (11). Of these, HR is the repair mechanism which is performed with high fidelity.

The process of HR repair, in which a number of proteins are involved, starts with DSBs recognition by the MRN complex that consists of three proteins: MRE11, RAD50, and NBS1. During the initial recruitment, MRN complex binds through its MRE11 component to DSB. Subsequently, RAD50 arms undergo structural changes, thereby allowing the bridging of both DNA ends. In a next step, signaling cascade is activated with the activation of the ATM kinase. The ATM kinase phosphorylates many other DNA damage response proteins, including members of the MRN complex (12).

One crucial step in HR repair is the formation of single-strand DNA overhangs at the sites of DNA ends. This process requires resection of 5’ termini to generate 3’ single-strand DNA overhangs (tails) and is initiated by the MRN complex in conjunction with CtIP and BRCA1 (10,13,14). More specifically, the bifunctional endo/exonuclease MRE11 can initiate DNA-end resection in 5’-3’ direction and begins by removing ~200–300 nucleotides away from the DSB site (15). This initial resection results in relatively short single-strand DNA overhangs and is followed by mass resection of DNA by either the 5’-3’ exonuclease 1 or DNA 2 in conjunction with the helicase Sgs1, thereby generating extensive single-strand DNA stretches (11,14,15). Subsequently, single-strand DNA is coated with replication protein A (RPA) complexes. In parallel, BRCA2 is recruited in a BRCA1- and PALB2-dependent manner to ultimately recruit recombinase RAD51, a DNA-dependent ATPase, to the single-strand DNA overhangs. Loading of RAD51 into single-strand DNA is a key step in HR repair. RAD51 replaces RPA and forms nucleoprotein filaments in single-strand DNA, which are essential for the homology search in sister chromatid and strand exchange (10,15,16).

Intermediary metabolic products of streptomycetes, including bleomycin, neocarzinostatin, and related compounds are effective cytotoxic agents. They directly induce DSB by attacking specific carbons in deoxyribose, leaving non-standard end-groups (17-19). Furthermore, topoisomerases inhibitors have DNA-based anti-tumor efficacy. Topoisomerases types I and II are enzymes that open and close one and both strands of DNA, respectively. Both types involve a covalent DNA-protein bond in their catalytic cycle (20,21). This transition state can be stabilized by topoisomerase inhibitors leading to DSB for type II inhibitors (epipodophyllotoxins) or single strand break (SSB) for type I inhibitors (camptothecins). Topoisomerases are covalently attached to the 5’-ends (bacterial type I, bacterial and eukaryotic type II) or 3’-ends (eukaryotic type I) of the broken DNA (22,23).

**Poly(ADP-ribose) polymerase (PARP): mechanism of action**

The PARPs are a family of enzymes capable of catalyzing the transfer of ADP-ribose to target proteins. To date, 17 members of the PARP family have been identified. PARP-1 is the first and best characterized member and accounts for ~90% of the total PARP activity. It plays a main role in DNA base excision repair (BER) and repair of DNA SSBs. Human PARP-1 has a multi-domain architecture composed of six domains. PARP-1 binding to the ends of an SSB is achieved through the coordinated action of two zinc finger domains located at the N-terminus of the protein. The catalytic domain is composed of an ADP-riboseyl transferase fold and a helical subdomain (10,24,25). PARP-1 binding to SSBs is followed by the polymerization of ADP-ribose (PARylation) with NAD+ as a substrate, resulting in the production of variable-sized polymers of ADP-ribose (PAR) (11). Successive addition of ADP-ribose units to form long and branched poly(ADP-ribose) chains, covalently linked to receptor proteins, including PARP-1, histone and other DNA repair proteins, leads to the formation of polymers adjacent to the DNA breaks. These highly negatively charged polymers form a scaffold and

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Mismatch repair (MMR)

The MMR system, an evolutionarily conserved mechanism, plays an important role in maintaining genomic stability. It recognizes and corrects biosynthetic errors that arise during DNA replication as well as mis paired bases that are generated during recombination or caused by oxidative DNA damage. Therefore, MMR reduces DNA errors 100–1,000 fold, and prevents them from becoming fixed mutations during cellular proliferation (33–37). Seven MMR genes, MLH1, MLH3, MSH2, MSH3, MSH6, PMS1, and PMS2 are involved in human MMR function.

Hereditary nonpolyposis colorectal cancer (HNPPC) or Lynch syndrome is caused by mutations in the genes that are responsible for MMR and is associated with a high risk of colon cancer, as well as other cancers including endometrial cancer, ovary, stomach, hepatobiliary tract, small intestine, upper urinary tract, brain, and skin. In ovarian cancer, MMR deficiency is the most common cause of hereditary ovarian cancer after HR deficiency accounting for another 10–15% of hereditary ovarian carcinomas (36,38). Pooled data from four large hereditary cancer registries in Europe and the United States determined a 6.7% lifetime risk for ovarian cancer in proven or probable MSH2 and MLH1 mutation carriers from Lynch syndrome families. Overall, women with HNPCC syndrome have approximately a 12–15% lifetime risk of developing ovarian cancer (39). In addition to the inherited gene mutations, other mechanisms of gene inactivation (such as promoter hypermethylation) leading to loss of expression of one of the main MMR genes occurs in up to 29% of cases (36).

Furthermore, there is published evidence that high mRNA levels of MSH6, MLH1, and PMS2 were associated with a favorable overall survival (OS) in ovarian cancer, suggesting these MMR genes may serve as potential positive prognostic indicators in ovarian cancer patients treated with platinum-based chemotherapy (40).

Susceptibility to EOC

Prevalence of pathogenic sequence variants (PVs) in cancer predisposing genes

Ovarian cancer is probably the tumor type with the highest percentage of hereditary cases observed, irrespective of selection criteria, such as early age at onset or family history. More specifically, a fourth to a fifth of unselected EOC patients carry PVs in a number of genes, the majority of which, encode for proteins involved in DNA repair pathways (41). BRCA1 and BRCA2 predisposing variants were the first to be associated to ovarian cancer, with the advent in DNA sequencing technologies leading to the discovery and association of additional genes. PVs of many
of the genes which encode for proteins involved in HR have been associated with increased risk of ovarian cancer, due to the significant role of HR in ovarian carcinogenesis (1,42-44).

The Cancer Genome Atlas (TCGA) found HR to be defective in approximately half of 489 women with stage II to IV high-grade serous ovarian cancers (HGSOCs) (45) attributed to germline variants in BRCA1 (in 8.5% of tumors) or BRCA2 (6.3%), somatic variants in BRCA1 (3.2%) or BRCA2 (2.9%) or epigenetic inactivation of BRCA1 (10.8%) due to promoter methylation resulting in transcriptional silencing. The prevalence of epigenetic/genetic mechanisms of BRCA1 aberration have been noted to vary between ethnicities, with PVs predominating in White Europeans, and methylation in people of African descent (46). Alsop et al. reported germline BRCA1 or BRCA2 PVs in approximately 15% of EOC patients, and 23% of patients with HGSOC (47).

In addition to the most commonly occurring alterations in BRCA1/2 genes, genetic aberrations in other genes compromise the HR pathway, including the Fanconi anemia genes (PALB2, PRIB1), the core RAD genes (RAD51C, RAD51D) and genes involved in HR pathway either directly (CHEK2, BARD1, NBN, ATM) or indirectly (such as CDK12, that encodes a cyclin-dependent kinase involved in the transcription regulation of several HR genes) (48-51). PVs in many of these genes confer increased, but variable, risks for ovarian cancer development. In addition, as previously mentioned, PVs genes involved in MMR pathway, account for 10–15% of hereditary EOC and is typically associated with tumors of endometrioid or clear-cell histopathology (1). Furthermore, PTEN downregulation (6.7%) has been associated with RAD51 transcriptional downregulation, while EMSY overexpression and amplification (7.9%) has been reported as another mechanism of HR deficiency in women with HGSOCs (1,50-52).

The majority of the aforementioned genes are included and analyzed in multi-gene panels utilized for hereditary cancer predisposition. HR deficient EOC has greater sensitivity to DNA-damaging agents that crosslink DNA, such as cisplatin, because HR is required for the repair of these lesions. Therefore, the identification of women with HR deficient ovarian cancers has significant clinical implications concerning chemotherapy regimen planning and development and use of targeted therapies as well. Cascade testing should also be offered to blood relatives of PV carriers.

Germline predisposing variants (Figure 1)

BRCA1 and BRCA2

PVs in BRCA1 and BRCA2 genes are detected in 10–15% (47,53-55) of unselected EOC patients, accounting for the majority of hereditary cases. BRCA1 and BRCA2 PVs confer 44% and 17% lifetime risks for ovarian cancer diagnosis, respectively (56), in contrast to ~1% risk for diagnosis in the general population. The vast majority of women carrying BRCA1 and BRCA2 PVs will be diagnosed with ovarian cancer, specifically with high-grade serous histology, with a median age of onset at ~51 and ~61 years, respectively(41,53,57). On the contrary, the largest prospective study performed to date, showed that the risk for ovarian cancer diagnoses was low up to the age of 40 and 50 years for BRCA1 and BRCA2 carriers, respectively (56). Ovarian cancers that bare pathogenic BRCA1 & BRCA2 variants are characterized by distinct clinical behaviors, illustrated by improved survival, visceral disease distribution, higher response rates to platinum and certain non-platinum chemotherapy agents, and sensitivity to PARPi.

Identification of BRCA1 and BRCA2 PVs can have major impact on appropriate clinical management of cancer patients through the choice of tailored treatments. Beyond the proven clinical benefit observed on the patients themselves, the genetic information can be used as a measure of primary prevention for ovarian cancer and is therefore critical for their family relatives. Asymptomatic carriers, that can be identified through cascade testing, and are at high risk, can be offered risk-reducing options that can ultimately lead to reduced morbidity and mortality. Simultaneously, through testing, all family relatives that can ultimately lead to reduced morbidity and mortality. Simultaneously, through testing, all family relatives that will test negative, can be assured that they have a lifetime ovarian risk, similar to the general population.

The proposed primary prevention measure for ovarian
cancer risk reduction among BRCA1 and BRCA2 carriers is bilateral salpingo-oophorectomy (BSO), as regular surveillance has been shown to be inadequate to detect tumors at an early stage. BSO, which can reduce the risk for ovarian cancer diagnosis up to 96% (57,58), should be considered after extensive counseling and discussion with the individual and of course, after completion of childbearing. Ideally, for BRCA1 carriers the procedure should be performed after the age of 35 and before the age of 40 years. For BRCA2 carriers, taking into consideration the lower penetrance and later onset of diagnosis, BSO can be performed after the age of 40 and before the age of 45 years (59,60). Keeping this age limits for BSO, the vast majority of ovarian cancers arising in the context of BRCA1 and BRCA2 predisposing variants will be avoided.

RAD51C and RAD51D

One of the co-localizing molecules with BRCA1 and BRCA2 in the repair of double-strand DNA breaks is RAD51C. RAD51C loss-of-function variants are rare among ovarian cancer patients, with their prevalence varying between 0.3–1.1% (61), with the upper end corresponding to studies enriched with cases with breast and ovarian cancer family history. The lifetime risk for ovarian cancer diagnosis among RAD51C carriers is ~5% (51,61). PVs in an additional member of the RAD51 family are also associated with increased ovarian cancer risk and are more frequently identified in familial ovarian cancer cases, reaching a prevalence of 5.1% in families with at least three ovarian cancer cases (62). Damaging RAD51D variants are marginally less frequent among ovarian cancer patients, compared to RAD51C PVs, observed in ~0.2% of unselected ovarian cancer patients (61,63) up to 0.9% of individuals having strong family history (62). RAD51D loss-of-function variants are associated with at least a six-fold increased risk for ovarian cancer, therefore identification of such genetic defects can have clinical utility (61,62).

Both RAD51C and RAD51D are ovarian cancer susceptibility genes, showing a reduced magnitude, when compared to BRCA1 and BRCA2, while identification of causative variants in females can lead informed decisions on preventative measures. PVs in any of these genes confer sufficient risk to guide risk-reducing BSO, as proposed in the National Comprehensive Cancer Network (NCCN) guidelines (64). Furthermore, it is plausible that genetic defects in these genes can facilitate as biomarkers for PARP sensitivity. Interestingly, there have been developed decision analytic models for the identification of the ovarian cancer risk thresholds, appropriate for risk reducing salpingo-oophorectomy-based primary prevention. Cost-effectiveness analysis has been highlighted by the National Institute of Health and Care Excellence (NICE) as the favored form of economic assessment to compare relative costs and health outcomes in decision modeling (65).

BRIP1

Heterozygous BRIP1 (BRCA1-interacting protein C-terminal helicase-1) damaging variants are associated to ovarian cancer susceptibility. The prevalence of PVs among familial ovarian cancer patients can be as high as 2.8% (66) or ~1% among unselected patients (51,61). The cumulative lifetime risk for EOC diagnosis among BRIP1 carriers has been estimated as 5–5.8% (61,67), with the most cases being diagnosed after menopause. Among the studies reporting the largest series of BRIP1 carriers, the mean age at diagnosis was 61–64 years (51,66,67). The elevated risk for EOC diagnosis justifies recommendation for risk-reducing removal of salpinges and ovaries among asymptomatic carriers and should be guided based on family history and individual’s choice, which can be offered at a later age, when compared to BRCA1 and BRCA2 carriers.

PALB2

Clinical testing for PALB2 (partner and localizer of BRCA2) has become a standard of practice, when evaluating breast cancer predisposition, while encoding for a key protein that works closely with BRCA1 and BRCA2 to facilitate DNA repair through HR. Nevertheless, the association of PALB2 PVs to ovarian cancer susceptibility is still conflicting, with the majority of studies reporting relative risks that ranged from 0.9 to 5.5 and lacked statistical power (52,67,68). Through monitoring 524 families known to carry PALB2 PVs, the risk to EOC predisposition was estimated to reach 5% by the age of 80 years (69). For PALB2 carriers, discussion and recommendation for risk-reducing surgery should take place for cases with strong family history for ovarian cancer. It should also be taken under consideration, that PALB2 PVs are indeed quite rare among ovarian cancer patients, identified in less than 0.5% of cases interrogated (51,61,67). A quite important observation, which provides promising therapeutic interventions, involves the sensitivity of PALB2 associated tumors to platinum-based chemotherapy and PARPis (46,70).

MLH1, MSH2, MSH6 and PMS2

Loss-of-function variants in the MMR genes, MLH1,
**Tumor testing in ovarian cancer**

The need to increase the number of ovarian cancer patients that can potentially benefit from treatment with targeted therapies, such as PARPis, has significantly increased the need for tumor testing through which additional genetic changes, which can predict sensitivity to PARP inhibition, can be identified. Indeed, an additional 4–7% of *BRCA1* and *BRCA2* PVs can be identified through tumor testing in EOC patients previously tested negative for germline loss-of-function *BRCA1* and *BRCA2* variants, while ~3% can bear somatic variants in other genes that are implicated in HR (78,79).

Hypothetically, through tumor testing, both germline and somatic genetic alterations can be detected, whereas germline testing alone cannot identify somatic mutations. Although the trend for tumor testing alone seems appealing, there are a number of factors that can influence the accuracy of such testing, while it can’t be considered as a gold standard for germline variant detection. Most importantly, in most cases the starting material for tumor testing is DNA extracted from formalin-fixed paraffin-embedded (FFPE) tissues, which can be technically challenging to amplify and to provide accurate results. In order to obtain tumor DNA, tumor microdissection is required. This means that in practical terms, a small diagnostic biopsy is likely to be unsuitable for further analysis. Of note, poor fixation can result in fragmented DNA, while the formalin itself can induce C/U deamination, leading to sequencing errors and the risk of false mutation calls. Another important aspect that should be taken into account is tumor heterogeneity and the actual percentage of tumor cells that are included in the tumor specimen, which can be challenging in FFPE DNA (80), while the phenomenon of genetic reversion of the initial *BRCA1* or *BRCA2* variant that
can occur most frequently as a result of chemotherapy, or even as an adaptive resistance mechanism (81). Given these considerations, although tumor testing can be proven of high importance during therapeutic decision-making, it certainly cannot replace traditional germline testing for assessing predisposition to hereditary cancer.

**Testing for HR deficiency**

It is evident that the majority of ovarian tumors presented with HR deficiency will derive from individuals harboring germline PVs in *BRCA1* and *BRCA2*. However, a number of tumors can have impaired HR pathway, as a result of alternative genetic defects, occurring either in the germline or in the soma, as previously described. Indeed, based on the data from TCGA, as much as half of the high-grade ovarian cancers harbor PVs in genes involved in the HR pathway (45). Of course, it has to be clarified that not all genetic alterations in these genes will result in HR deficiency.

Identification of HR deficiency in a tumor can be reflected through additional assays which can determine the genomic instability and can provide an HR deficiency score by measuring: the generic loss of heterozygosity (LOH), telomeric allelic imbalance and large-scale state transitions. Such functional assays aim to determine the genomic scars that are created following accumulation of genomic alterations and have been mainly developed and patented by commercial companies. Both LOH and HR deficiency score have been implemented as biomarkers, by a number of trials testing the efficiency of different PARPis, such as niraparib (82) and rucaparib (83). Interestingly, in the latter ARIEL2 study, 34% of *BRCA* wild type patients with LOH low archival specimen had an LOH high pretreatment specimen highlighting a change in biomarker status over time. In contrast, there was no case in which the classification changed from LOH high to LOH low between the archival and pretreatment specimen. Based on the variability and the complexity of the different genetic phenomena that can occur in an ovarian tumor, the available tests should provide higher levels of accuracy when ordered and used in a complementary manner.

**The role of PARPis in EOC and their therapeutic potential in patients with HR deficiency (Table 1)**

PARPis have transformed treatment for ovarian cancer. Olaparib received FDA approval based on the results from Study 19. This phase II study randomized women with relapsed platinum-sensitive HGSOC who had received at least two prior platinum-based chemotherapy regimens and were in complete or partial response to their most recent regimen to olaparib maintenance treatment or placebo. Olaparib treatment improved progression-free survival (PFS) in the overall population (hazard ratio 0.35, P<0.0001). In the *BRCA* mutant population, the benefit of olaparib versus placebo was even greater than in the *BRCA* wild-type population (90). The SOLO2 phase III trial was designed to confirm the efficacy of olaparib maintenance in patients with *BRCA* mutated, relapsed EOC who were in complete or partial response to their most recent platinum-based regimen. Olaparib maintenance treatment provided a significant PFS improvement with no detrimental effect on quality of life in these patients (85). Another phase III trial (SOLO1), evaluated the efficacy of olaparib as maintenance therapy in patients with newly diagnosed advanced EOC with a mutation in *BRCA1*, *BRCA2*, or both who had a complete or partial clinical response after first-line platinum-based chemotherapy. The risk of disease progression or death was 70% lower with olaparib than with placebo (84). Finally, in the phase III SOLO3 trial patients with platinum sensitive, relapsed EOC with germline *BRCA* mutations were randomized to olaparib or non-platinum chemotherapy of physician’s choice. The study demonstrated that treatment with the PARPi resulted in better outcomes [overall response rate (ORR) and PFS] than non-platinum chemotherapy in these women (86).

Trials with two other PARPi, niraparib and rucaparib, have confirmed the efficacy of these drugs as maintenance therapy. More specifically, the phase III ENGOT-OV16/NOVA trial evaluated the efficacy of niraparib versus placebo as maintenance treatment for patients with platinum-sensitive, recurrent ovarian cancer. Patients in the niraparib group had a significantly longer median PFS than did those in the placebo group (21.0 versus 5.5 months in the germline *BRCA* mutant cohort, 12.9 versus 3.8 months in the non-germline *BRCA* mutant cohort with HR deficiency, and 9.3 versus 3.9 months in the overall non-germline *BRCA* mutant cohort) (82). Niraparib was also evaluated in women with newly diagnosed advanced EOC who had a response to platinum-based chemotherapy (PRIMA/ENGOT-OV26/GOG-3012 trial). Among 373 (50.9%) patients with HR deficiency, the median PFS was 21.9 months in the niraparib group and 10.4 months in the placebo group (hazard ratio 0.43, P<0.001). In the entire population, the corresponding PFS was 13.8 and 8.2 months, respectively (hazard ratio 0.62, P<0.001). At
Table 1  Selected phase III trials with PARP inhibitors for epithelial ovarian cancer

<table>
<thead>
<tr>
<th>Study</th>
<th>Population</th>
<th>Treatment arms</th>
<th>Median PFS</th>
<th>HR</th>
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<tr>
<td>SOLO 1, (84)</td>
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<td>SOLO 2, (85)</td>
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<td>Olaparib maintenance versus placebo</td>
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<td>0.3</td>
<td>&lt;0.0001</td>
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<td>0.27</td>
<td>&lt;0.001</td>
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<td>Non-gBRCAm cohort with HRD*: 12.9 versus 3.8 months</td>
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<td>Overall non-gBRCAm cohort: 9.3 versus 3.9 months</td>
<td>0.45</td>
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<td>Overall population: 13.8 versus 8.2 months</td>
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<td>PRIMA, (87)</td>
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<td>Patients with HRD: 21.9 versus 10.4 months</td>
<td>0.43</td>
<td>&lt;0.001</td>
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<td>Overall population: 13.8 versus 8.2 months</td>
<td>0.62</td>
<td>&lt;0.001</td>
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<td>ARIEL3, (88)</td>
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<td>0.23</td>
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<td>Patients with HRD: 13.6 versus 5.4 months</td>
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<td>ITT* population: 10.8 versus 5.4 months</td>
<td>0.36</td>
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<td>0.59</td>
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<td>Patients with HRD including those with BRCA mutations: 37.2 versus 17.7 months</td>
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<td>&lt;0.001</td>
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<td>Patients with HRD without BRCA mutations: 28.1 versus 16.6 months</td>
<td>0.43</td>
<td>&lt;0.001</td>
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*4, epithelial ovarian cancer; *b, germline BRCA mutated; *c, homologous recombination deficiency; *d, intent-to-treat. PARPi, poly(ADP-ribose) polymerase; PFS, progression-free survival; HR, hazard ratio; EOC, epithelial ovarian cancer; HRD, homologous recombination deficiency.

24 months, the rate of overall survival was 84% in the niraparib group and 77% in the placebo group (87).

The phase III ARIEL3 trial, assessed rucaparib versus placebo after response to second-line or later platinum-based chemotherapy in patients with high-grade, recurrent, platinum-sensitive ovarian carcinoma. Median PFS in patients with a BRCA4-mutant carcinoma (germline or somatic) was 16.6 months in the rucaparib group versus 5.4 months (hazard ratio 0.23, P<0.0001). In patients with a HR deficient carcinoma it was 13.6 versus 5.4 months (hazard ratio 0.32, P<0.0001), while in the intention-to-treat population, it was 10.8 versus 5.4 months (hazard ratio 0.36, P<0.0001). Thus, ARIEL3 provided further evidence that use of a PARPi in the maintenance treatment setting could be considered a new standard of care for women with platinum-sensitive EOC following a complete or partial response to second-line or later platinum-based chemotherapy (88). These trials clearly indicate that the relationship between sensitivity to PARPi and HR repair deficiency is likely to be more similar to a continuous than to a fixed and discrete variable. As such, PARPi have the potential to be beneficial in a much wider proportion of EOC patients than was originally proposed (91).

More recently, because PARP inhibition has immuno-
regulatory effects (92,93), combination therapy of PARP and immune checkpoint inhibitors is being developed. A phase II study (MEDIOLA) of olaparib and the PD-L1 inhibitor durvalumab in patients with relapsed, platinum-sensitive, BRCA1-mutated ovarian cancer showed an ORR of 72% (94). The phase I/II study TOPACIO investigated the combination of niraparib and pembrolizumab in patients with platinum-resistant/refractory ovarian cancer (95). The ORR was 18% and the disease control rate was 65%. There was no difference in response by BRCA and homologous recombination deficiency (HRD) status. Currently, several studies that combine PARPi with olaparib, pembrolizumab, and pembrolizumab and cediranib, with olaparib, pembrolizumab and pembrolizumab of combination of niraparib plus pembrolizumab in patients with platinum-resistant/refractory ovarian cancer (96). The ORR was 18% and the disease control rate was 65%. There was no difference in response by BRCA and homologous recombination deficiency (HRD) status. Currently, several studies that combine PARPi and immune checkpoint inhibitors are being carried out. Among them, two interesting clinical trials should be mentioned: ENGOT-OV4+/FIRST study compares the efficacy and safety of standard of care platinum-based therapy ± dostarlimab (an anti-PD-1 humanized monoclonal antibody that has shown clinical activity as monotherapy in early phase trials) followed by niraparib ± dostarlimab maintenance as first-line treatment of stage III or IV EOC (96); ATHENA (GOG-3020/ENGOT-OV45) is a randomized, double-blind, placebo-controlled, phase 3 study of rucaparib + nivolumab following frontline platinum-based chemotherapy (97).

Because PARPi and anti-angiogenic inhibitors are also synergistic (92,98), the combination of these drugs also has been studied. In the phase 3 trial PAOLA-1, patients with newly diagnosed, advanced, high-grade EOC having a response after first-line platinum-taxane chemotherapy plus bevacizumab were randomly assigned in a 2:1 ratio to receive olaparib or placebo for up to 24 months and bevacizumab for up to 15 months in total. Maintenance with olaparib provided a significant PFS benefit, which was substantial in patients with HRD-positive tumors, including those without a BRCA mutation (89).

Beyond PAOLA-1, several other trials are examining the role of anti-angiogenesis, in combination with PARPi in the different treatment lines of EOC, and different subpopulations as well. In most studies, the combination of cediranib [an oral, highly potent, VEGF receptor (VEGFR) tyrosine kinase inhibitor with activity against all three VEGFRs] with olaparib is tested. The randomized, phase II AVANOVA2 trial compared niraparib and bevacizumab versus niraparib alone as definitive treatment for platinum-sensitive recurrent ovarian cancer (99). Niraparib plus bevacizumab significantly improved PFS compared with niraparib alone (median PFS 11.9 versus 5.5 months, respectively; adjusted hazard ratio 0.35, P<0.0001). Following the encouraging results of a randomized phase II trial (NCT01116648) which showed significant improvement in PFS with cediranib + olaparib versus olaparib monotherapy in recurrent platinum-sensitive EOC, notably in non-germline BRCA mutated patients (100), the phase IIIb CONCERTO study was conducted. This study investigated the combination of cediranib plus olaparib in non-germline BRCA mutated patients with recurrent platinum-resistant EOC who had received ≥3 previous lines of therapy for advanced disease. The combination showed evidence of antitumor activity in this heavily pretreated subpopulation, while toxicity was manageable with dose modifications (100). In contrast to the previous study, the randomized, phase II OCTOVA trial, investigates the benefit of single agent olaparib compared to olaparib plus cediranib or weekly paclitaxel in women with BRCA-mutated platinum-resistant ovarian cancer (101). Also, a randomized phase II/III study (COCOS), compares of the combination of cediranib and olaparib to cediranib or olaparib alone, or standard of care chemotherapy in women with recurrent platinum-resistant or -refractory ovarian, fallopian tube, or primary peritoneal cancer (102). Finally, the randomized phase III ICON 9 trial, assesses the efficacy, safety and tolerability of maintenance olaparib in combination with cediranib compared to maintenance olaparib alone following a response to platinum-based chemotherapy in women with relapsed platinum-sensitive ovarian, fallopian tube or peritoneal cancer (103).

In clinical practice, beyond anti-angiogenics, and immune checkpoint inhibitors, we aim to reduce overlapping toxicities by combining PARPi separately with several other agents, such as phosphoinositide 3-kinase (PI3K), protein kinase B (AKT), mammalian target of rapamycin (mTOR), WEE1, mitogen-activated protein kinase (MEK), and cyclin dependent kinase (CDK) 4/6 inhibitors (104).

**Recommendations on testing for EOC**

Summarizing the available evidence on germline and tumor testing for EOC patients, and having as a goal to target both the hereditary and therapeutic component of ovarian cancer, germline genetic testing should be offered to all women diagnosed with EOC. This referral should be done irrespectively of their age at diagnosis or relevant family history, while the test ordered should be able to detect damaging variants in all genes associated to ovarian cancer susceptibility and not be restricted to BRCA1 and BRCA2 testing. Subsequently, it is recommended to order tumor
testing, at least for BRCA1 and BRCA2 genes, in all women tested negative for germline PVs. It would be of the greatest benefit of the patients that the test(s) is offered at the time of initial diagnosis (105).

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