



Analysis of preoperative circulating tumor cells for recurrence in patients with hepatocellular carcinoma after liver transplantation

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Background: Liver transplantation (LT) is an effective option for hepatocellular carcinoma (HCC) and end-stage liver cirrhosis. However, organ shortage and tumor recurrence are the main obstacles limiting its popularization and application in HCC patients. Testing for circulating tumor cells (CTCs) would be a valuable strategy to predict the recurrence and metastasis of HCC after LT. Various CTCs detection methods have different advantages and disadvantages. This study aims to investigate the predictive value of testing for CTCs based on immunofluorescence in situ hybridization of peripheral blood cells in patients with HCC after LT.

Methods: A total of 50 patients who received testing for CTCs and then underwent LT were enrolled in this study. Negative enrichment and immunofluorescence in situ hybridization (imFISH) methods were introduced to detect CTCs.

Results: Twenty-six (52%) patients were CTC-positive, and 24 (48%) patients were CTC-negative. The results showed that CTCs result was correlated with tumor size ($\chi^2=5.773$, $P=0.016$), AFP level ($\chi^2=5.454$, $P=0.020$), tumor grade ($\chi^2=6.478$, $P=0.039$) and Recurrence ($\chi^2=6.211$, $P=0.013$). Twelve patients had recurrence after LT within one year. The results showed that the CTCs result ($P=0.034$) was the only independent factor impacting long-term survival. The 1-year disease-free survival rates of CTC-negative and CTC-positive patients were 91.6% and 61.5%, respectively ($P=0.020$). The 1-year overall survival of CTC-positive patients and CTC-negative was 88.5% and 91.7%, respectively ($P=0.751$).

Conclusions: CTCs result was closely related to the early recurrence of patients with HCC after LT. CTC-positive patients had a worse prognosis after LT than the CTC-negative group.

Keywords: Circulating tumor cell (CTC); hepatocellular carcinoma (HCC); liver transplantation (LT)

Submitted Mar 22, 2020. Accepted for publication Jul 27, 2020.

doi: 10.21037/atm-20-2751

View this article at: <http://dx.doi.org/10.21037/atm-20-2751>

Introduction

Hepatocellular carcinoma (HCC) is a malignant disease with high morbidity and mortality. In the world, HCC is the fifth most common and third most deadly cancer, and it is the third leading cause of cancer-related death in China (1). Hepatectomy was once the standard treatment to cure HCC worldwide. However, according to some studies, even with radical resection, the overall 5-year recurrence rates of large hepatocellular carcinoma (LHC) and small hepatocellular carcinoma (SHC) are 80% and 43.5%, respectively (2). The main cause of recurrence is tumor cell dissemination via blood vessel infiltration. Circulating tumor cells (CTCs) are likely to spread before or after operation without a definite detection method. In addition, squeezing and destroying tumors during operation is also an important factor leading to hematogenous dissemination of tumor cells.

CTCs were first discovered in the blood of a patient with metastatic cancer and described by Ashworth *et al.* in 1869 (3). With nearly 100 years of improvements, the predictive value of CTCs has been emphasized. In 1998, Racila *et al.* introduced a new method to detect CTCs and suggested that CTCs appeared early and were associated with tumor progression, creating the theoretical basis for the clinical use of CTCs in current cancer research and therapy (4). Vona *et al.* first investigated the prognostic value of blood CTCs in patients with HCC and showed a close association with prognosis and recurrence in patients with HCC (5). Multifarious approaches were developed for detection, and various CTC methods result in different results. At present, the existing CTC detection methods are as follows: Immunoaffinity-based positive enrichment (6) (antibodies target tumor cell-associated antigens and then adhere to magnetic particles), immunoaffinity-based negative enrichment (7) (antibodies target unwanted background cells and then adhere to magnetic particles), biophysical separation methods [cells are separated by biophysical characteristics such as density (8)], direct imaging modalities (a series of novel methods developed to replace enrichment), functional assays (enrichment is based on cell bioactivity), and so on.

Liver transplantation (LT) is an effective treatment option for both tumors and cirrhosis (9). Patients who meet the Milan criteria and the University of California San Francisco (UCSF) criteria can achieve encouraging survival outcomes after LT. Saposochin *et al.* and Bruix *et al.* reported that the 5-year post-transplantation overall survival rate could reach 75% for patients with HCC

meeting the criteria (10). However, organ shortage and recurrence limited the selection of LT treatment in patients with HCC. Prognoses of tumor recurrence based on CTCs are meaningful for the selection of LT patients and follow-up observations. This retrospective study aims to investigate the predictive value of testing for CTCs for early recurrence of patients with HCC after LT. We present the following article in accordance with the STROBE reporting checklist (available at <http://dx.doi.org/10.21037/atm-20-2751>).

Methods

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and approved by the Institutional Ethics Committee for Clinical Research and Animal Trials of the First Affiliated Hospital of Sun Yat-sen University and informed consent waiver was granted by the IEC given the retrospective, minimal risk nature of the study [Approval ID: [2020]088].

General information

Three hundred and twenty-eight patients were diagnosed and then underwent LT in our department from January 1st, 2016 to February 28th, 2019. Among them, a total of 94 patients who were tested for CTCs were enrolled. The inclusion criteria were as follows: 18 to 75 years of age, a diagnosis of HCC confirmed by postoperative pathological examination, and follow-up of more than 1 year. The exclusion criteria were as follows: patients with perioperative or nonrecurrence-related mortality, a diagnosis of other types of tumors, and follow-up of less than 1 year. Three patients' postoperative pathological diagnoses were without tumors, 9 patients died within 30 days of operation and 32 patients' follow-up periods were less than 1 years. Afterwards, 50 patients were included in this study. The immunosuppressive regimen was tacrolimus (Tac) + mycophenolate mofetil (MMF). Corticosteroid was not included in routine regimen for its side effects.

CTCs detection

The samples for CTCs analysis were collected at pre-LT. Negative enrichment and imFISH methods were introduced to detect CTCs. Peripheral blood (3.2 mL) was processed by a Human Blood Cell Deletion Kit (Cytel). First, red blood cells were lysed and depleted. The residual cell pellet was suspended again and incubated with anti-

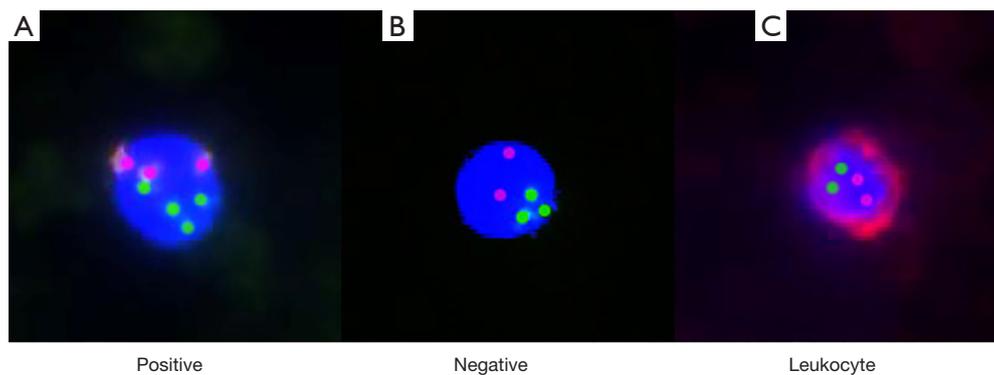


Figure 1 Preoperative CTC fluorescence imaging in patients with HCC. The phenotype of CTCs was CEP8+/DAPI+/CD45- (DAPI: blue; CEP8: orange; CD45: red). When the CEP8 signal was ≥ 3 , it was considered positive (A). When the CEP8 signal was ≤ 2 , it was considered negative (B). The phenotype of leukocytes was CEP8+/CD45+/DAPI+ (C).

CD45 monoclonal antibody-coated magnetic beads for 20–30 minutes. Then, A magnetic stand (Promega, Madison, WI) was used to separate these beads. The supernatant was subsequently used for identification.

The identification of enriched CTCs was performed by imFISH, which combined the FISH probes with chromosome 8 (orange) centromere probes (Abbott Molecular Diagnostics, Des Plaines, IL, USA) and anti-CD45 monoclonal antibodies (Red, Cytel). First, the CEP8 probe and specimen were hybridized at 37 °C for 20–90 minutes in a hybridizer (DAKO). Subsequently, they were washed in 50% formamide at 43 °C for 15 minutes and then subjected to gradient alcohol. Finally, the specimens were incubated with anti-CD45 antibodies conjugated to Alexa Flour 594 (Invitrogen) for 1 hour. Afterwards, they were washed again with 0.2% BSA. Finally, the specimens were stained with DAPI, which contained Vectashield mounting medium. To be considered positive, CTCs needed to be hyperdiploid and have the phenotype CEP8+/DAPI+/CD45-. The CTC test results are shown in *Figure 1*.

Follow-up

The patients were followed up for at least 1 year. For deceased patients/patients with recurrences, the date of death/recurrence was used as the last follow-up. Postoperative visits were performed on postoperative day (POD) 1-7, POD14 and each postoperative month (POM). Biomedical values, imaging examinations and tumor markers were documented. Routine Doppler ultrasound of the liver graft blood flow and biliary tract was performed once every 2 days for 7 days. Afterward, imaging studies

were performed based on patients' clinical status or laboratory findings. The follow-up deadline was February 29th, 2020.

Statistical analysis

All statistical analyses of the data were performed by SPSS version 26.0. All data are expressed as the mean \pm standard deviation or the number and percentage of patients. For comparison between groups, the chi-square and Fisher's exact tests were performed for frequencies and continuous data, respectively. Cox proportional hazards model was performed for multivariate analysis. Overall and disease-free survival were compared using the Kaplan–Meier method with a log-rank test. A P value <0.05 was considered statistically significant.

Result

Patient characteristics

To eliminate selection bias, we compared baseline data between CTC-test group and no CTC-test group (*Table 1*) and there were no significant differences between groups ($P>0.05$). A total of 50 patients who were tested for CTCs and then underwent LT were analyzed, and their characteristics are shown in *Tables 2* and *3*. The male-to-female ratio was 24 to 1. The mean age was 52.88 ± 1.43 years old. Forty-seven (94%) patients had cirrhosis, and 38 (76%) patients had hepatitis B virus (HBV) infection. Nineteen (38%) patients had larger tumors (>6.5 cm) and 5 (10%), 24 (48%), and 21 (42%) patients

Table 1 Comparison of baseline data between CTC-test group and no CTC-test group

Variables	Total (n=328) (%)	CTC-test (n=50) (%)	No CTC-test (n=278) (%)	χ^2	P value
Age, years				0.12	0.728
<50	137 (41.8)	22 (6.7)	115 (35.1)		
≥50	191 (58.2)	28 (8.5)	163 (49.7)		
Gender				2.220	0.36
Male	296 (80.2)	48 (14.6)	248 (75.6)		
Female	32 (9.8)	2 (0.6)	30 (9.1)		
AFP (μg/L)				0.150	0.699
≤400	237 (72.3)	35 (10.7)	202 (61.6)		
>400	91 (27.7)	15 (4.6)	76 (23.2)		
Cirrhosis				0.005	0.946
No	19 (5.8)	3 (0.9)	16 (4.9)		
Yes	309 (94.2)	47 (14.3)	262 (79.9)		
TNM staging				2.275	0.517
I	29 (8.8)	4 (1.2)	25 (7.6)		
II	103 (31.4)	16 (4.9)	87 (26.5)		
III	174 (53.0)	29 (8.8)	145 (44.2)		
IV	22 (6.7)	1 (0.3)	21 (6.4)		

CTC, circulating tumor cells; AFP, preoperative alpha-fetoprotein.

Table 2 Association between peripheral blood CTCs count and baseline data of 50 patients

Variables	Count (%)	Peripheral blood CTCs count (%)		χ^2	P value
		>1	0		
Age, years				0.102	0.746
<50	22 (6.7)	12 (3.7)	10 (3.0)		
≥50	28 (8.5)	14 (4.3)	14 (4.3)		
Gender				0.003	0.954
Male	48 (14.6)	25 (7.6)	23 (7.0)		
Female	2 (0.6)	1 (0.3)	1 (0.3)		
Tumor size (cm)				5.773	0.016
≤6.5	31 (9.5)	12 (3.7)	19 (5.8)		
>6.5	19 (5.8)	14 (4.3)	5 (1.5)		
Tumor amount				0.765	0.382
≤3	39 (11.9)	19 (5.8)	20 (6.1)		
>3	11 (3.4)	7 (2.1)	4 (1.2)		
PVT				0.056	0.814
No	41 (12.5)	21 (6.4)	20 (6.1)		
Yes	9 (2.7)	5 (1.5)	4 (1.2)		

Table 2 (continued)

Table 2 (continued)

Variables	Count (%)	Peripheral blood CTCs count (%)		χ^2	P value
		>1	0		
Milan criteria				2.826	0.093
No	27 (8.2)	17 (5.2)	10 (3.0)		
Yes	23 (7.0)	9 (2.7)	14 (4.3)		
UCSF criteria				1.282	0.258
No	22 (6.7)	15 (4.6)	7 (2.1)		
Yes	28 (8.5)	11 (3.4)	17 (5.2)		
Pre-transplantation treatment				3.994	0.550
TACE	10 (3.0)	7 (2.1)	3 (0.9)		
Resection	2 (0.6)	1 (0.3)	1 (0.3)		
RFA	14 (4.3)	6 (1.8)	8 (2.4)		
Resection with TACE	1 (0.3)	0	1 (0.3)		
Resection with TACE and RFA	1 (0.3)	0	1 (0.3)		
None	22 (6.7)	12 (3.7)	10 (3.0)		
AFP ($\mu\text{g/L}$)				5.454	0.020
≤ 27	25 (7.6)	11 (3.4)	14 (4.3)		
> 27	25 (7.6)	15 (4.6)	10 (3.0)		
HBsAg				0.025	0.874
Negative	12 (3.7)	6 (1.8)	6 (1.8)		
Positive	38 (11.6)	20 (6.1)	18 (5.5)		
Cirrhosis				2.946	0.086
No	3 (0.9)	3 (0.9)	0 (0)		
Yes	47 (14.3)	23 (7.0)	24 (7.3)		
Tumor grade*				6.478	0.039
Grade I	5 (1.5)	0 (0)	5 (1.5)		
Grade II	24 (7.3)	15 (4.6)	9 (2.7)		
Grade III	21 (6.4)	11 (3.4)	10 (3.0)		
TNM staging				1.483	0.686
I	4 (1.2)	2 (0.6)	2 (0.6)		
II	16 (4.9)	7 (2.1)	9 (2.7)		
III	29 (8.8)	16 (4.9)	13 (4.0)		
IV	1 (0.3)	1 (0.3)	0 (0)		
MVI				0.056	0.814
Yes	9 (2.7)	5 (1.5)	4 (1.2)		
No	41 (12.5)	21 (6.4)	20 (6.1)		
Recurrence				6.211	0.013
Yes	12 (3.7)	10 (3.0)	2 (0.6)		
No	38 (11.6)	16 (4.9)	22 (6.4)		
Death				0.588	0.443
Yes	6 (1.8)	4 (1.2)	2 (0.6)		
No	44 (13.4)	22 (6.7)	22 (6.7)		

*, HCC cellular differentiation. CTC, circulating tumor cells; PVT, portal vein thrombosis; TACE, transarterial chemoembolization; RFA, radiofrequency ablation; AFP, preoperative alpha-fetoproteins; HBsAg, hepatitis B surface antigen; MVI, microvascular invasions.

Table 3 Analysis of relevant factors for recurrence of HCC in 50 patients

Variables	Count (%)	Recurrence (%)		χ^2	P value
		Yes	No		
Age, years				1.317	0.251
<50	22 (6.7)	7 (2.1)	15 (4.6)		
≥50	28 (8.5)	5 (1.5)	23 (7.0)		
Gender				0.658	0.417
Male	48 (14.6)	12 (3.7)	36 (11.0)		
Female	2 (0.6)	0 (0)	2 (0.6)		
Tumor size (cm)				13.773	0.000
≤6.5	31 (9.5)	2 (0.6)	29 (8.8)		
>6.5	19 (5.8)	10 (3.0)	9 (2.7)		
Tumor amount				3.559	0.059
≤3	39 (11.9)	7 (2.1)	32 (9.8)		
>3	11 (3.4)	5 (1.5)	6 (1.8)		
PVT				2.515	0.113
No	41 (12.5)	8 (2.4)	33 (10.1)		
Yes	9 (2.7)	4 (1.2)	5 (1.5)		
Milan criteria				5.469	0.019
No	27 (8.2)	10 (3.0)	17 (5.2)		
Yes	23 (7.0)	2 (0.6)	21 (6.4)		
UCSF criteria				9.914	0.002
No	22 (6.7)	10 (3.0)	12 (3.7)		
Yes	28 (8.5)	2 (0.6)	26 (7.9)		
Pre-transplantation treatment				4.488	0.481
TACE	10 (3.0)	3 (0.9)	7 (2.1)		
Resection	2 (0.6)	1 (0.3)	1 (0.3)		
RFA	14 (4.3)	1 (20.3)	13 (4.0)		
Resection with TACE	1 (0.3)	0	1 (0.3)		
Resection with TACE and RFA	1 (0.3)	0	1 (0.3)		
None	22 (6.7)	7 (2.1)	15 (4.6)		
AFP (μg/L)				1.754	0.185
≤27	25 (7.6)	4 (1.2)	21 (6.4)		
>27	25 (7.6)	8 (2.4)	17 (5.2)		
HBsAg				0.010	0.921
Negative	12 (3.7)	3 (0.9)	9 (2.7)		
Positive	38 (11.6)	9 (2.7)	25 (7.6)		

Table 3 (continued)

Table 3 (continued)

Variables	Count (%)	Recurrence (%)		χ^2	P value
		Yes	No		
Cirrhosis				3.185	0.074
No	3 (0.3)	2 (0.6)	1 (0.3)		
Yes	47 (14.3)	10 (3.0)	37 (11.3)		
Tumor grade*				1.709	0.426
Grade I	5 (1.5)	0 (0)	5 (1.5)		
Grade II	24 (7.3)	6 (1.8)	18 (5.5)		
Grade III	21 (6.4)	6 (1.8)	15 (4.6)		
TNM staging				4.486	0.214
I	4 (1.2)	0	4 (1.2)		
II	16 (4.9)	2 (0.6)	14 (4.3)		
III	29 (8.8)	10 (3.0)	19 (5.8)		
IV	1 (0.3)	0	1 (0.3)		
CTCs				6.211	0.013
>1	26 (7.9)	10 (3.0)	16 (4.9)		
0	24 (7.3)	2 (0.6)	22 (6.7)		
MVI				2.515	0.113
Yes	9 (2.7)	4 (1.2)	5 (1.5)		
No	41 (12.5)	8 (2.4)	33 (10.0)		

*, HCC cellular differentiation. HCC, hepatocellular carcinoma; PVT, portal vein thrombosis; TACE, transarterial chemoembolization; RFA, radiofrequency ablation; AFP, preoperative alpha-fetoproteins; HBsAg, hepatitis B surface antigen; CTC, circulating tumor cells.

had tumors classified as grade I, grade II and grade III, respectively (11). The median alpha fetoprotein (AFP) levels in these 50 patients was 27 $\mu\text{g/L}$. Four, 16, 29 and 1 patients had tumors classified as stage I, II, II and IV, respectively. The median follow-up time was 16.4 months (IQR, 15.4–25.55) for recurrence and 20.0 months (IQR, 12.28–24.07) for survival, respectively.

CTCs result

The data of 50 patients were analyzed and are shown in Table 2. Twenty-six (52%) patients were CTC-positive (CTCs >1), and 24 (48%) patients were CTC-negative (CTCs \leq 1). The mean ages of the CTC-positive group and CTC-negative group were 52.31 \pm 1.97 years old and 53.50 \pm 2.12 years old, respectively. The median preoperative AFP levels of the CTC-positive group and CTC-negative group were 109.0 and 15.9 $\mu\text{g/L}$, respectively. The results

showed that CTCs result was correlated with tumor size ($\chi^2=5.773$, $P=0.016$), AFP level ($\chi^2=5.454$, $P=0.020$), tumor grade ($\chi^2=6.478$, $P=0.039$) and Recurrence ($\chi^2=6.211$, $P=0.013$).

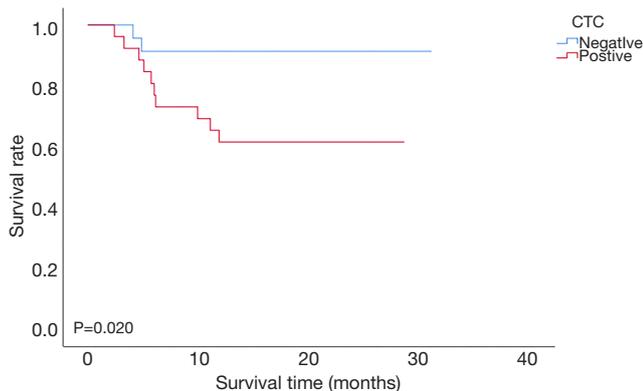
Analysis of recurrence factors after LT

Analysis of the 50 patients revealed that 12 (24%) patients had a recurrence after LT (Table 3). All 12 patients were male, and the mean age was 50.67 \pm 3.39 years old. The mean preoperative AFP levels of the recurrence group and non-recurrence group were 115,840.99 \pm 111,306.46 and 945.82 \pm 573.35 $\mu\text{g/L}$, respectively. Ten of these 12 patients had cirrhosis. Ten of these 12 patients were positive for CTCs. The results showed that recurrence was correlated with tumor size ($\chi^2=13.733$, $P<0.001$), compliance with Milan criteria ($\chi^2=5.469$, $P=0.019$), compliance with USCF criteria ($\chi^2=9.914$, $P=0.002$), and the CTCs result ($\chi^2=6.211$,

Table 4 Multivariate analysis of relevant factors for recurrence of HCC in 50 patients

Variable	Multivariate analysis	
	RR (95% CI)	P value
AFP	2.018 (0.591–6.891)	0.263
CTCs	5.411 (1.132–25.874)	0.034
Pre-transplantation intervention	0.505 (0.154–1.660)	0.261
Tumor grade*	1.395 (0.450–4.329)	0.564
MVI	1.769 (0.745–4.200)	0.196

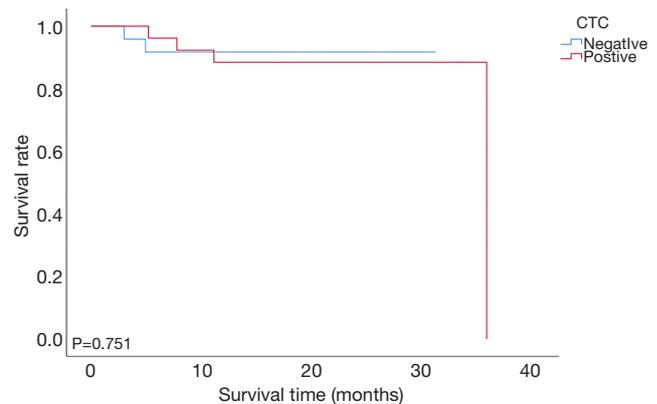
*, HCC cellular differentiation. HCC, hepatocellular carcinoma; AFP, preoperative alpha-fetoproteins; CTCs, circulating tumor cells; MVI, microvascular invasions.

**Figure 2** Comparison of disease-free survival between the CTC-positive group and the CTC-negative group in patients with HCC after LT. CTC, circulating tumor cell; HCC, hepatocellular carcinoma.

P=0.013), while there were no significant differences in other groups like preoperative AFP ($\chi^2=1.754$, P=0.185). In multivariate analysis, CTCs result was the only independent impact factor [CTCs: P=0.034; preoperative AFP: P=0.263; Pre-transplantation intervention: P=0.261; Tumor grade: P=0.564; microvascular invasions (MVI): P=0.196] (Table 4).

Analysis of disease-free survival and overall survival after LT

During the 12-month follow-up period, two of 24 CTC-negative patients and 10 of 26 CTC-positive patients experienced recurrence. The mean disease-free survival times of CTC-negative and CTC-positive patients were 28.75 ± 1.56 and 20.14 ± 2.13 months, respectively. The

**Figure 3** Comparison of overall survival between the CTC-positive group and the CTC-negative group in patients with HCC after LT. CTC, circulating tumor cell; HCC, hepatocellular carcinoma.

1-year disease-free survival rates of CTC-negative and CTC-positive patients were 91.6% and 61.5%, respectively (91.6% vs. 61.5%, P=0.020, Figure 2). The mean overall survival times of CTC-negative and CTC-positive patients were 28.81 ± 1.53 and 32.50 ± 2.0 months, respectively. The 1-year overall survival rate of patients with CTC-positive was similar to those with CTC-negative (88.5% vs. 91.7%, P=0.751, Figure 3). Thirty-two patients whose follow up periods were less than 1 year did not suffer from deaths or recurrences.

Discussion

Hepatectomy was once the standard treatment to cure HCC worldwide. However, even with complete resection, the recurrence after hepatectomy remains high. LT is an attractive option to cure HCC completely. It can offer R0 resection and achieve the widest negative incisional margin (12). Early studies revealed poor outcomes after LT for HCC related to tumor pathological staging (13). Several advances have been made in the field of LT over decades. With the development of selection criteria such as the Milan criteria and the UCSF criteria (14), technical advances, and enhanced knowledge of HCC staging and biology, the therapeutic efficacy of LT for HCC has achieved an improvement in the 5-year survival rate (15). Approximately 40% of LT recipients are HCC patients, which means that HCC has been a frequent indication for LT (16,17). However, obstacles exist and affect the prognosis of LT for HCC. Organ shortage has become a worldwide issue, with many patients dying on the waiting list (18,19). For patients

with HCC, finding a method to predict the prognosis after LT is of great significance. In this study, we aimed to investigate the predictive value of testing for CTCs for the early recurrence of patients with HCC after LT.

First, we analyzed the baseline data of 50 patients. The patients were mostly male and had been diagnosed in their 50s. A large proportion of the tumors developed because of liver cirrhosis with chronic HBV infection and were in an advanced stage when discovered. The reason for these findings may be that in China, patients tend to undergo surgical resection or conservative treatment due to economic or ideological reasons, even if tumors are detected early. LT would be considered only when other treatments were ineffective or if the tumor progressed.

Testing for CTCs is a new noninvasive examination method for early diagnosis, determination of treatment options and prognostic evaluation in HCC due to the continuous improvements in methods related to cell separation and identification (20). Immunoaffinity-based positive enrichment (as also called positive enrichment) and immunoaffinity-based negative enrichment (as also called negative enrichment) are commonly used to detect CTCs. Positive enrichment uses antibodies to target tumor cell-associated antigens, which then adhere to magnetic particles, so tumor cells are captured. Epithelial cell adhesion molecule (EpCAM) is usually used as target antigen to capture CTCs, like the processed used by the Cell Search System (21). This system was approved by the FDA to monitor patients with metastatic solid cancers. However, the lower expression of EpCAM in HCC than in other solid tumors, such as breast cancer and colorectal cancer (22), results in false negative outcomes. Yamashita *et al.* reported that only 35% of HCC cells are positive for EpCAM (23). A low positive expression rate hampers its use in monitoring patients with tumor metastasis after LT. Negative enrichment, in contrast with positive enrichment, uses antibodies to target unwanted background cells such as leukocytes, which then adhere to magnetic particles, so tumor cells are retained. It has advantages in that it is simple, provides easy-to-use batch separation and allows for high-throughput processing (24). Compared with positive enrichment, it can enrich for all subtypes of CTCs. In addition, the size, density, and charge of CTCs will change due to cell damage and immune attack, and the accuracy of biophysical-related separation methods may be affected. Negative enrichment does not depend on any physical properties of tumor cells, so it can eliminate this effect. ImFISH further improves the sensitivity and

signal intensity. In our study, by using this method, CTCs were detected in 26 of 50 (52%) patients before LT. The sensitivity and specificity of CTCs detection in our study were 83.3% and 62.5%, respectively. The results showed that this strategy had good sensitivity and specificity so it could be helpful to detect tumors. Schulze *et al.* revealed that CTCs that were EpCAM (+) had a positive correlation with HCC staging and AFP level (25). In our study, the results showed that CTCs result was correlated with tumor size ($\chi^2=5.773$, $P=0.016$), AFP level ($\chi^2=5.454$, $P=0.020$) and tumor grade ($\chi^2=6.478$, $P=0.039$). Our study results supported Schulze's report.

With improvements in CTCs detection methods, most studies have revealed that CTCs are related to HCC recurrence, metastasis, disease-free survival (DFS) and overall survival (OS). Cheng *et al.* (26) revealed that total CTCs are more effective compared with AFP in the diagnosis of HCC. Our study showed that the preoperative positive rate of CTCs and the postoperative recurrence rate in patients with HCC were 52% and 24%, respectively. The recurrent rate in patient meeting Milan criteria and UCSF criteria was 8.7% and 7.1%, respectively. The recurrence time after LT was earlier in CTC-positive patients than in the CTC-negative group (20.14 ± 2.13 and 28.75 ± 1.56 months, $P=0.020$). The 1-year overall survival rate of patients with CTC-positive was smaller than those with CTC-negative, but without significantly difference (88.5% *vs.* 91.7%, $P=0.751$). In multivariate analysis, we found that CTCs result was the only independent impact factor affecting recurrence and preoperative AFP was not significantly different. Sun *et al.* (27) reported that patients with more CTCs had a worse prognosis after surgery than patients with less CTCs. However, Wang (28) suggested that CTCs levels and subtype were not effective in monitoring HCC recurrence after LT. Our study supported Sun's conclusion. Some reports concluded that LT survival within one year can reach 80% to 90% (29). In our study, 1-year overall survival rate was 88% (44/50), and it was consistent with the current epidemiological results. We didn't observe the effect of CTCs result on survival rate, and the reason would be small sample size and short follow-up period. Residual CTCs may still cause metastasis or recurrence even after removal of the primary cancer or transplantation, and changes in the number of CTCs during therapy may predict prognosis and recurrence in patients with cancer (30). Therefore, further studies are needed to investigate the predictive value of postoperative CTCs in patients with HCC.

Our study is limited by its small sample size and by being from a single-center institution. Larger multicenter studies are needed to determine whether CTCs result can predict recurrence in patients with HCC after LT. Second, other tumor types, such as cholangiocarcinoma, were not investigated in this study. In addition, postoperative CTCs were not included in this study. The association between postoperative CTCs and early recurrence of patients with HCC after LT should have been analyzed in the study. For future studies, the 3- and 5-year DFS and OS values should be calculated to obtain more convincing conclusions.

Conclusions

In conclusion, CTCs result was closely related to the early recurrence of patients with HCC after LT, and patients in the CTC-positive group had a worse prognosis than those in the CTC-negative group. Larger multicenter studies are needed, and longer follow-up outcomes should be used to obtain more convincing results.

Acknowledgments

Funding: This work was supported by the National Natural Science Foundation of China (81401324 and 81770410), Science and Technology Planning Project of Guangdong Province (2016A020215048), Guangdong Provincial Key Laboratory of Organ Donation and Transplant Immunology (2013A061401007), Guangdong Basic and Applied Basic Research Foundation (2020A1515011557), Guangdong Provincial International Cooperation Base of Science and Technology (Organ Transplantation) (2015B050501002), and Scientific Program for Young Teacher of Sun Yat-sen University (16ykpy05), China.

Footnote

Reporting Checklist: The authors have completed the STROBE reporting checklist. Available at <http://dx.doi.org/10.21037/atm-20-2751>

Data Sharing Statement: Available at <http://dx.doi.org/10.21037/atm-20-2751>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/atm-20-2751>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and approved by the Institutional Ethics Committee for Clinical Research and Animal Trials of the First Affiliated Hospital of Sun Yat-sen University and informed consent waiver was granted by the IEC given the retrospective, minimal risk nature of the study {Approval ID: [2020]088}.

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References

1. Chen W, Zheng R, Baade PD, et al. Cancer statistics in China, 2015. *CA Cancer J Clin* 2016;66:115-32.
2. Wang JH, Wang CC, Hung CH, et al. Survival comparison between surgical resection and radiofrequency ablation for patients in BCLC very early/early stage hepatocellular carcinoma. *J Hepatol* 2012;56:412-8.
3. Ferreira MM, Ramani VC, Jeffrey SS. Circulating tumor cell technologies. *Mol Oncol* 2016;10:374-94.
4. Racila E, Euhus D, Weiss AJ, et al. Detection and characterization of carcinoma cells in the blood. *Proc Natl Acad Sci U S A* 1998;95:4589-94.
5. Vona G, Sabile A, Louha M, et al. Isolation by size of epithelial tumor cells : a new method for the immunomorphological and molecular characterization of circulating tumor cells. *Am J Pathol* 2000;156:57-63.
6. Cristofanilli M, Budd GT, Ellis MJ, et al. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med* 2004;351:781-91.
7. Lara O, Tong X, Zborowski M, et al. Enrichment of rare cancer cells through depletion of normal cells using density and flow-through, immunomagnetic cell separation. *Exp Hematol* 2004;32:891-904.
8. Harouaka RA, Zhou MD, Yeh YT, et al. Flexible micro spring array device for high-throughput enrichment of

- viable circulating tumor cells. *Clin Chem* 2014;60:323-33.
9. Sangiovanni A, Colombo M. Treatment of hepatocellular carcinoma: beyond international guidelines. *Liver Int* 2016;36 Suppl 1:124-9.
 10. Sapisochin G, Bruix J. Liver transplantation for hepatocellular carcinoma: outcomes and novel surgical approaches. *Nat Rev Gastroenterol Hepatol* 2017;14:203-17.
 11. Quaglia A. Hepatocellular carcinoma: a review of diagnostic challenges for the pathologist. *J Hepatocell Carcinoma* 2018;5:99-108.
 12. Gorgen A, Muaddi H, Zhang W, et al. The New Era of Transplant Oncology: Liver Transplantation for Nonresectable Colorectal Cancer Liver Metastases. *Can J Gastroenterol Hepatol* 2018;2018:9531925.
 13. Dageforde LA, Fowler KJ, Chapman WC. Liver transplantation for hepatocellular carcinoma: current update on treatment and allocation. *Curr Opin Organ Transplant* 2017;22:128-34.
 14. Yao FY, Ferrell L, Bass NM, et al. Liver transplantation for hepatocellular carcinoma: expansion of the tumor size limits does not adversely impact survival. *Hepatology* 2001;33:1394-403.
 15. Mazzaferro V, Regalia E, Doci R, et al. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. *N Engl J Med* 1996;334:693-9.
 16. Kim WR, Lake JR, Smith JM, et al. OPTN/SRTR 2015 Annual Data Report: Liver. *Am J Transplant* 2017;17 Suppl 1:174-251.
 17. Saidi RF, Hejazi Kenari SK. Liver transplantation for hepatocellular carcinoma: past, present and future. *Middle East J Dig Dis* 2013;5:181-92.
 18. Bhogal RH, Sutaria R, Gunson BK, et al. Similar liver transplantation survival with selected cardiac death donors and brain death donors (*Br J Surg* 2010; 97: 744-753). *Br J Surg* 2010;97:1310; author reply 1310-1.
 19. Wadei HM, Lee DD, Croome KP, et al. Early Allograft Dysfunction Is Associated With Higher Risk of Renal Nonrecovery After Liver Transplantation. *Transplant Direct* 2018;4:e352.
 20. Carr BI. Some new approaches to the management of hepatocellular carcinoma. *Semin Oncol* 2012;39:369-73.
 21. Cristofanilli M, Hayes DF, Budd GT, et al. Circulating tumor cells: a novel prognostic factor for newly diagnosed metastatic breast cancer. *J Clin Oncol* 2005;23:1420-30.
 22. Barbazan J, Dunkel Y, Li H, et al. Prognostic Impact of Modulators of G proteins in Circulating Tumor Cells from Patients with Metastatic Colorectal Cancer. *Sci Rep* 2016;6:22112.
 23. Yamashita T, Forgues M, Wang W, et al. EpCAM and alpha-fetoprotein expression defines novel prognostic subtypes of hepatocellular carcinoma. *Cancer Res* 2008;68:1451-61.
 24. Liu Z, Fusi A, Klopocki E, et al. Negative enrichment by immunomagnetic nanobeads for unbiased characterization of circulating tumor cells from peripheral blood of cancer patients. *J Transl Med* 2011;9:70.
 25. Schulze K, Gasch C, Staufer K, et al. Presence of EpCAM-positive circulating tumor cells as biomarker for systemic disease strongly correlates to survival in patients with hepatocellular carcinoma. *Int J Cancer* 2013;133:2165-71.
 26. Cheng Y, Luo L, Zhang J, et al. Diagnostic Value of Different Phenotype Circulating Tumor Cells in Hepatocellular Carcinoma. *J Gastrointest Surg* 2019;23:2354-61.
 27. Sun YF, Xu Y, Yang XR, et al. Circulating stem cell-like epithelial cell adhesion molecule-positive tumor cells indicate poor prognosis of hepatocellular carcinoma after curative resection. *Hepatology* 2013;57:1458-68.
 28. Wang S, Zheng Y, Liu J, et al. Analysis of circulating tumor cells in patients with hepatocellular carcinoma recurrence following liver transplantation. *J Investig Med* 2018;66:1-6.
 29. Meirelles Junior RF, Salvalaggio P, Rezende MB, et al. Liver transplantation: history, outcomes and perspectives. *Einstein (Sao Paulo)* 2015;13:149-52.
 30. Barreto G, Neve R, Shrikhande SV, et al. Role of circulating tumour cells in predicting recurrence after excision of primary colorectal carcinoma (*Br J Surg* 2007; 94: 96-105). *Br J Surg* 2007;94:644; author reply 644-5.

Cite this article as: Chen Z, Lin X, Chen C, Chen Y, Zhao Q, Wu L, Wang D, Ma Y, Ju W, Chen M, He X. Analysis of preoperative circulating tumor cells for recurrence in patients with hepatocellular carcinoma after liver transplantation. *Ann Transl Med* 2020;8(17):1067. doi: 10.21037/atm-20-2751