The effects of citrate dialysate in hemodialysis on polymorphonuclear elastase interaction with tissue factor and its inhibitor

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Background: This study aimed to investigate whether hemodialysis (HD) affects tissue factor (TF), tissue factor pathway inhibitor (TFPI), and polymorphonuclear elastase (PMNE) in endstage renal disease (ESRD) patients when eliminating the effects of heparin. Also, to explore the interaction of TF, TFPI, and PMNE throughout a single HD session.

Methods: We enrolled 57 ESRD patients who had undergone hemodialysis for >3 months as an experimental group. Plasma levels of TF, TFPI and PMNE were measured by ELISA in 24 ESRD patients on intermittent HD using low-molecular-weight heparin (LMWH) as anticoagulation (LMWH group) and 33 ESRD patients using citrate as anticoagulation (citrate group) at the start and at 1, 2 and 5 h of the HD session. Meanwhile, 28 ESRD patients not on dialysis were enrolled as a control group and fasting venous blood samples were taken in the morning.

Results: Compared with the control group, the plasma TFPI levels of the LMWH group and the citrate group were significantly higher (P=0.001, P=0.02, respectively) under baseline conditions as well as the plasma PMNE levels (P=0.001, P=0.02, respectively), whereas TF showed no difference (P=0.186). During HD with citrate, plasma TFPI decreased slightly (P=0.012) and PMNE increased significantly (P=0.008) at 1 h. The plasma TFPI levels of the citrate group correlate with PMNE at 2 and 5 h (P=0.001, P=0.008, respectively).

Conclusions: ESRD patients on HD have significantly higher TFPI and PMNE levels compared to patients not on HD under baseline conditions, while TF levels were similar between the three groups. TFPI and PMNE are differently regulated, but the plasma levels correlated during HD in the citrate group. It might be possible that PMNE plays a role in anticoagulative activity through TFPI.

Keywords: Hemodialysis (HD); tissue factor (TF); tissue factor pathway inhibitor (TFPI); polymorphonuclear elastase (PMNE)
Introduction

As the incidence of chronic kidney disease (CKD) steadily increase worldwide, hemodialysis (HD) is widely used for treating end-stage renal disease (ESRD). Adequate anticoagulation is a prerequisite for HD both to prevent clotting in the extracorporeal circuit (1), influencing the safety and efficacy of HD therapy. In China, there are three common anticoagulation means clinically, including unfractionated heparin (UFH), low-molecular-weight heparin (LMWH), and ‘no heparin,’ where LMWH is the prioritized option in maintenance HD. However, CKD patients already have an increased risk of excess bleeding, such as platelet dysfunction caused by uremia (2,3), let alone the potentially fatal heparin-induced thrombocytopenia type II and ‘no heparin’-induced consumption of platelet when receiving dialysis (4,5). Regional citrate anticoagulation has been tested on dialysis, but it might lead to potentially disastrous complications, such as citrate accumulation presenting metabolic acidosis, hypernatremia, and citrate net overload presenting metabolic alkalosis (6). The use of anticoagulation during HD remains a problem for ESRD patients with a high risk of bleeding complications. A new, safer, and more effective anticoagulation mean in HD is required.

Recently, many scholars have focused on tissue factor (TF), which initiates the extrinsic pathway of blood coagulation (4) and plays an important role in atherosclerosis (7,8). It was recently reported that adherent cells from the dialysis membrane were mainly constituted by activated polymorphonuclear neutrophils (PMNs), which triggered thrombin generation in a TF-dependent manner (9). The tissue factor pathway inhibitor (TFPI) is the only endogenous protein that effectively inhibits TF-factor VIIa in a factor Xa-dependent manner (10,11). The use of anticoagulation during HD remains a problem for ESRD patients with a high risk of bleeding complications. A new, safer, and more effective anticoagulation mean in HD is required.

Methods

Study population

Thirty-three selected ESRD patients (CKD5d), who had already undergone hemodialysis for >3 months and used citrate for anticoagulation for >3 days, were enrolled in the study (citrate group). They were compared with age, sex and etiology-matched ESRD patients who had undergone hemodialysis for >3 months using LMWH as anticoagulation (LMWH group) (n=24) and those who had never received dialysis (control group) (n=28). Each case of ESRD was diagnosed with the calculated estimated glomerular filtration rate (eGFR) <10 mL/min/1.73 m^2 using the simplified Modification of Diet in Renal Disease (MDRD) formula and was present for >3 months (17). Subjects were excluded from the study if they presented with any of the following diseases: malignancy, hyperlipidemia (cholesterol concentrations >6.21 mmol/L), advanced liver disease, contraceptive, or blood transfusion within three days before the study. The etiology of ESRD patients are displayed in Table 1.

The study was approved by the Research Ethics Committee of West China Hospital of Sichuan University (2016-188), and informed consent was obtained from all participants.

Dialysis details

Hemodialysis was applied using a temporary femoral venous catheter (n=36) or a permanent internal jugular venous cuff catheter (n=21). All these patients used polyacrylonitrile (AN69) as the dialysis membrane and were maintained on low-flux (130–200 mL/min) intermittent renal replacement therapy. For the citrate group, anticoagulation was carried out using sodium citrate (4.0%) continuously infused at a speed of 130–200 mL/h and extra sodium bicarbonate (5%) at 100–170 mL/h. For the LMWH group, a single i.v. bolus of LMWH at the start of hemodialysis (range, 1,000–2,500 IU) was followed by a constant infusion (range, 100–250 IU) and extra sodium bicarbonate (5%) at 190–260 mL/h. There were two dialysis patterns: day-time continuous veno-venous hemodialysis (CVVHD) and day-time continuous veno-venous hemodialysis filtration (CVVHDF) (Table 1). Correspondingly, the displacement velocity was 1,500/1,500 mL/h and 3,000 mL/h, and all adopted post-dilution.
Blood samples from hemodialysis patients were collected before a single dialysis session and at hours (h) 1, 2, and 5 during the hemodialysis. Residual containing fluid was removed from catheters before collection of the first sample. When collecting blood samples during the hemodialysis, we shut off the peripheral pump of the dialyzer for 3 min except for the blood pump and then collected blood samples directly from the arterial line on the machine. Fasting venous blood samples were taken from the antecubital vein in the morning from the control of ESRD patients. Citrated-plasma samples were prepared conventionally, centrifuged (15 minutes, 3,000 ×g), aliquoted and stored at −80 °C until the assay.

### Analytical procedures

Plasma levels of TF and TFPI were measured in duplicate by enzyme-linked immunosorbent assays (ELISA) using commercially available standard kits supplied by R&D Systems, Inc., USA. The plasma level of PMNE was measured by ELISA kit purchased from Life Technologies, Carlsbad, USA.

### Statistical analysis

Quantitative data followed the normal distribution and were expressed as the mean ± SD (standard deviation) in tables or mean ± SEM (standard error of the mean) in figures. Characteristics of the participants were first presented as numbers and percentages. The data obtained at different time points were corrected by hematocrit. Chi-square test, one-way analysis of variance (one-way ANOVA) or Mann Whitney U test were used to adjust models controlling for sex, etiology, and age. Comparisons of the TF, TFPI, and PMNE plasma levels between the three groups and the levels at different time points were performed using one-way analysis of variance or Kruskal-Wallis H test. Bivariate correlation analysis was applied to explore the association between measured plasma constituents before hemodialysis and clinical indexes and the correlation of TF, TFPI, and PMNE plasma levels at each time point during hemodialysis. All statistical analyses were performed using IBM SPSS software SPSS 24. P<0.05 was considered statistically significant.
Results

ESRD patients’ demographic characteristics did not differ significantly between the three groups (Table 1). Compared with the control subjects, patients on hemodialysis with LMWH and citrate showed significant higher plasma concentrations of TFPI (P=0.000, P=0.002, respectively) and PMNE (P=0.001, P=0.02, respectively) under baseline conditions (Figure 1). However, TF plasma levels (P=0.186) between the three groups were similar (Figure 1A). Also, we found under baseline conditions that the cholesterol of ESRD patients in the citrate group was lower than that of the control group, including total cholesterol (P=0.002), high-density lipoprotein cholesterol (HDL-C) (P=0.009), and low-density lipoprotein-cholesterol (LDL-C) (P=0.000). The clinical indexes of the studied groups are shown in Table 2.

The results of factors correlating with plasma TF, TFPI, and PMNE levels in ESRD patients before hemodialysis are presented in Tables 3,4. We did not find any correlation between the pre-dialysis TF, TFPI, PMNE levels, and clinical indexes in the LMWH group (Table 3). However, in the citrate group, the pre-dialysis TF level was significantly correlated to glucose and monocyte % (r=0.345 and r=0.355, respectively; P<0.05). Also, the pre-dialysis TFPI level was positively associated with cholesterol (r=0.406, P=0.019) and LDL-C (r=0.490, P=0.004), whereas we did not find any clinical index that was correlated to the PMNE level before hemodialysis (Table 4).

As the different time points were measured, presented in Table 5, showed that the plasma levels of TFPI and PMNE fluctuated obviously (P=0.000, P=0.000, respectively) during HD with LMWH, but TF did not change significantly (P=0.935). Further analysis showed that plasma TF levels did not vary obviously (Figure 2A); plasma TFPI levels ascended significantly at 1 h (P=0.000), descended slightly at 2 h (P=0.364) and significantly at 5 h (P=0.000) (Figure 2B); the plasma levels of PMNE increased significantly at 1 h (P=0.000) and continued to rise at 2 h but did not reach a significant level (P=1.000) (Figure 2C).

Figure 1  The measured plasma constituents of ESRD patients with hemodialysis were obtained before a single dialysis session. (A) Comparison of plasma TF between three groups shows no difference; (B) Plasma TFPI in hemodialysis patients is higher than that of patients without hemodialysis; (C) Plasma PMNE in patients with hemodialysis with citrate is higher than that of patients without hemodialysis. Values are expressed as mean ± SEM. *P<0.05, **P<0.01, ***P<0.001; a comparison is made by one-way analysis of variance or Kruskal-Wallis H test. ESRD, endstage renal disease; TF, tissue factor; TFPI, tissue factor pathway inhibitor; PMNE, polymorphonuclear elastase.
The trends of TF, TFPI and PMNE in the LMWH group were shown in Figure 2D. For the citrate group, we found that the plasma levels of PMNE had changed significantly throughout hemodialysis (P=0.000), but the overall changes of TF (P=0.986) and TFPI (P=0.088) did not reach statistical significance (Table 6). Further analysis found that the change of plasma TF levels was not obvious, either (Figure 3A); plasma TFPI levels declined modestly at 1 h (P=0.012) (Figure 3B); the plasma levels of PMNE increased significantly at 1 h (P=0.008) and continued to rise at 2 h.

Table 2 Laboratory data

<table>
<thead>
<tr>
<th>Measured Plasma Constituents</th>
<th>ESRD patients without hemodialysis</th>
<th>ESRD patients with hemodialysis</th>
<th>P</th>
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<tbody>
<tr>
<td></td>
<td></td>
<td>Hemodialysis with LMWH</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Hemodialysis with citrate</td>
<td></td>
</tr>
<tr>
<td>TF (pg/mL)</td>
<td>82.96±18.24</td>
<td>88.84±16.89</td>
<td>0.186††</td>
</tr>
<tr>
<td>TFPI (ng/mL)</td>
<td>26.41±12.79</td>
<td>119.79±21.04</td>
<td>0.000††</td>
</tr>
<tr>
<td>PMNE (ng/mL)</td>
<td>33.10±20.87</td>
<td>50.69±18.98</td>
<td>0.001††</td>
</tr>
<tr>
<td>Creatinine (µmol/L)</td>
<td>721.39±162.2</td>
<td>493.96±286.03</td>
<td></td>
</tr>
<tr>
<td>Cystatin C (mg/L)</td>
<td>4.40±0.95</td>
<td>4.63±2.02</td>
<td>0.450**</td>
</tr>
<tr>
<td>MONO%</td>
<td>6.38±2.09</td>
<td>6.64±3.82</td>
<td>0.826**</td>
</tr>
<tr>
<td>Triglyceride (mmol/L)</td>
<td>1.83±1.11</td>
<td>1.99±1.34</td>
<td>0.464††</td>
</tr>
<tr>
<td>Cholesterol (mmol/L)</td>
<td>3.84±1.29</td>
<td>3.41±0.99</td>
<td>0.002††</td>
</tr>
<tr>
<td>HDL-C (mmol/L)</td>
<td>1.10±0.37</td>
<td>0.90±0.44</td>
<td>0.012††</td>
</tr>
<tr>
<td>LDL-C (mmol/L)</td>
<td>2.17±1.06</td>
<td>1.67±0.86</td>
<td>0.000**</td>
</tr>
</tbody>
</table>

The values of ESRD patients with hemodialysis were obtained before a single dialysis session. Values are expressed as mean ± SD; a comparison is made by one-way analysis of variance** or Kruskal-Wallis H test††. *, P<0.05 compared with the control group; †, P<0.05 compared with the LMWH group. ESRD, end-stage renal disease; LMWH, low-molecular-weight heparin; TF, tissue factor; TFPI, tissue factor pathway inhibitor; PMNE, polymorphonuclear elastase; MONO%, the count of monocyte/the count of white blood cell%; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol.

Table 3 Correlation of clinical indexes and TF, TFPI and PMNE levels of ESRD patients on hemodialysis with LMWH

<table>
<thead>
<tr>
<th>Laboratory results</th>
<th>Pre-dialysis TF</th>
<th>Pre-dialysis TFPI</th>
<th>Pre-dialysis PMNE</th>
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<tbody>
<tr>
<td></td>
<td>r</td>
<td>P</td>
<td>r</td>
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<tr>
<td>Glucose (mmol/L)</td>
<td>0.215</td>
<td>0.313</td>
<td>0.105</td>
</tr>
<tr>
<td>Albumin (g/L)</td>
<td>−0.051</td>
<td>0.814</td>
<td>0.214</td>
</tr>
<tr>
<td>MONO%</td>
<td>0.044</td>
<td>0.840</td>
<td>0.030</td>
</tr>
<tr>
<td>Creatinine (µmol/L)</td>
<td>−0.153</td>
<td>0.476</td>
<td>0.142</td>
</tr>
<tr>
<td>Cystatin C (mg/L)</td>
<td>−0.139</td>
<td>0.516</td>
<td>0.100</td>
</tr>
<tr>
<td>BUN (mmol/L)</td>
<td>0.074</td>
<td>0.733</td>
<td>0.126</td>
</tr>
<tr>
<td>Triglyceride (mmol/L)</td>
<td>0.214</td>
<td>0.315</td>
<td>−0.096</td>
</tr>
<tr>
<td>Cholesterol (mmol/L)</td>
<td>−0.248</td>
<td>0.243</td>
<td>0.129</td>
</tr>
<tr>
<td>HDL-C (mmol/L)</td>
<td>−0.086</td>
<td>0.691</td>
<td>0.084</td>
</tr>
<tr>
<td>LDL-C (mmol/L)</td>
<td>−0.348</td>
<td>0.096</td>
<td>0.229</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SD. A correlation is made by bivariate correlation analysis. TF, tissue factor; TFPI, tissue factor pathway inhibitor; PMNE, polymorphonuclear elastase; MONO%, the count of monocyte/the count of white blood cell%; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol.
but did not reach statistical significance (P=1.000), then descended slightly at 5 h (P=1.000) (Figure 3C). The trends of TF, TFPI, and PMNE in the citrate group were shown in Figure 3D.

We used bivariate correlation analysis to explore the correlation of TF, TFPI, and PMNE plasma levels at each time point during hemodialysis. There was a possible correlation between TFPI levels and PMNE levels throughout the hemodialysis session with citrate, especially at 2 h (r=0.534, P=0.001) and 5 h (r=0.456, P=0.008), as shown in Figure 4. Furthermore, the change of the plasma PMNE level between 0 and 1 h was associated with the change of the plasma TFPI level between 0 and 1 h (r=0.454, P=0.008) (Figure 4C). However, there was no such change and correlation for TF and the LMWH group.

Also, we divided the experimental group into CVVHD group and CVVHDF group according to the different HD modes and analyzed whether there were differences in TF, TFPI and PMNE plasma levels at different time points in the dialysis process between the two groups. No significant difference was found between CVVHD and CVVHDF groups.

**Discussion**

The major finding of the present study is that hemodialysis affected the plasma levels of TFPI and PMNE of ESRD patients but had minimal effect on plasma TF. Moreover, we found that, excluding the effect of heparin on TFPI and PMNs, PMNE might play a role in anticoagulative activity through TFPI during hemodialysis.

Numerous studies indicate that, compared to healthy
volunteers, plasma levels of TF and TFPI are higher in patients undergoing maintenance hemodialysis therapy (18-20). However, the variation of TF plasma levels during hemodialysis is inconsistent, as TF levels at times increase (21) or do not (18,19). It is contentious whether dialysis affects TF. Our study demonstrated that there was no difference in TF levels between ESRD patients with or without hemodialysis and that the TF plasma levels didn’t change during HD no matter what category of anticoagulation was adopted. These data suggest that a decreased kidney function is associated with increased TF levels, but not HD. TF is associated with enhanced oxidative stress (22,23), endothelial injury (24) and uremic solutes (25,26) in chronic kidney disease. A recent article reported that advanced oxidation protein products (AOPPs) and serum TF correlate with kidney function. ESRD patients who are on HD had significantly higher serum levels of AOPPs and TF compared to patients after kidney transplantation with a decent GFR (27). The study indicated that TF was still strongly associated with oxidative stress caused by renal insufficiency in HD patients. It was reported that the dialyzer membrane triggered TF

Table 6 Variation of plasma TF, TFPI and PMNE levels during hemodialysis with citrate

<table>
<thead>
<tr>
<th>Variables</th>
<th>Measured Plasma Constituents During Experiments</th>
<th>0 h</th>
<th>1 h</th>
<th>2 h</th>
<th>5 h</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>TF (pg/mL)</td>
<td>91.58±18.51</td>
<td>92.26±19.05</td>
<td>92.99±18.78</td>
<td>93.13±19.55</td>
<td>0.986*</td>
<td></td>
</tr>
<tr>
<td>TFPI (ng/mL)</td>
<td>44.80±10.22</td>
<td>38.13±9.45</td>
<td>41.51±10.52</td>
<td>42.46±12.20</td>
<td>0.088*</td>
<td></td>
</tr>
<tr>
<td>PMNE (ng/mL)</td>
<td>47.01±23.31</td>
<td>85.98±54.66</td>
<td>100.15±68.95</td>
<td>88.00±60.41</td>
<td>0.000†</td>
<td></td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SD. All the data are corrected by hematocrit. The analysis is done by one-way analysis of variance or Kruskal-Wallis H test. TF, tissue factor; TFPI, tissue factor pathway inhibitor; PMNE, polymorphonuclear elastase.

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expression of activated PMNs (9), but, in our study, the plasma levels of TF were not correlated with PMNE, which was a marker of activated PMNs. A possible reason for this phenomenon could be that TF was expressed richly enough because of renal insufficiency; consequently, the effect of dialysis on the tissue factor is weak. Thus, we speculate that the change in plasma TF level may be associated with chronic renal failure superior to HD. Also, the TF level of pre-dialysis was correlated with glucose and MONO%, which is similar to that of previous studies (10).

Several studies found that TFPI levels tended to rise significantly from minute 15 and minute 60 onward during hemodialysis (21,28,29), which is similar to the trend of the LMWH group in our study. There is no doubt that the use of heparin can significantly increase the TFPI level in hemodialysis patients. Also, the TF level of pre-dialysis was correlated with glucose and MONO%, which is similar to that of previous studies (10).

Figure 3 Variation of plasma TF, TFPI and PMNE levels at different points during hemodialysis with citrate. (A) Variation of plasma TF levels at different time points during hemodialysis with citrate shows no difference; (B) Plasma TFPI declines lightly at 1 h; (C) Plasma PMNE increases significantly at 1 h; (D) Variation trends of TF, TFPI, PMNE during hemodialysis. Values are expressed as mean ± SEM. All the data are corrected by hematocrit. *, P<0.05, **, P<0.01; a comparison is made by one-way analysis of variance or Kruskal-Wallis H test. TF, tissue factor; TFPI, tissue factor pathway inhibitor; PMNE, polymorphonuclear elastase.

during hemodialysis when using citrate for anticoagulation. Our study indicated that the pre-dialysis plasma level of TFPI was slightly higher in ESRD patients of the citrate group than that in controls. This may be due to the coagulation disorders in patients of the citrate group after receiving long-term hemodialysis. During hemodialysis, the anticoagulant effect of heparin is approximately 30%, mediated by the TFPI released from its endothelial stores into the circulating blood, which will lead to untoward depletion of TFPI (30,31). Repeated and continuous heparin administration will lead to excessive depletion of intravascular pools of TFPI, thereby resulting in attenuation of its contribution to the antithrombotic action of heparin (32) and protective effect on atherosclerosis. However, the results should be viewed with caution because the effect of dialysis cannot be ruled out.

The hemodialyzer membrane activates PMNs and promotes the release of PMNE (33,34). PMNE is a marker of the biological incompatibility of the dialyzer. Heparin is reported to stimulate PMNs in synergy with the stimulatory effect of dialysis membranes in vivo (35), which is similar to the PMNE changes in the LMWH group in our research.
Meanwhile, the PMNE changes of the citrate group in our study reconfirmed that when eliminating the effect of heparin, hemodialysis itself also stimulated PMNs and elevated the PMNE level. PMNE was considered to reflect adequate anticoagulative activity, as when anticoagulation was inadequate, the elastase values rose consistently (36). TFPI, as the unique inhibitor of the extrinsic coagulation pathway, plays an important part in defending thrombosis during hemodialysis (37). Recently, Massberg et al. bred mice with injured carotid arteries that were deficient in PMNE and cathepsin G (Elane−/−; Ctsg−/− mice), attesting that PMNE inactivated TFPI by involving local proteolysis and then sequentially promoted intravascular thrombus growth in vivo (12). In our study, we also found a possible correlation between the plasma level of TFPI and PMNE during a heparin-free hemodialysis session. We didn’t find such correlation in the LMWH group probably because heparin has a great influence on TFPI but a little influence on PMNE, which is mainly affected by hemodialyzer membrane. The present study suggests that when eliminating the effect of heparin on TFPI and PMNs, PMNE might play a role in anticoagulative activity through TFPI during hemodialysis.

The study, however, also bears some limitations. Although plasma levels of TF and TFPI can represent for their content in the body, alternatively spliced isoforms of TF and TFPI are differentially expressed by endothelial cells and human platelets (10,38). We only measured the plasma level of TF and TFPI, and it exists with some limitations. A prospective cohort study with a large number of hemodialysis subjects and direct biochemical research is required to confirm our results.

In conclusion, the change in the plasma TF level during HD is associated with chronic renal failure superior to HD. When eliminating the effect of heparin, hemodialysis can reduce the plasma concentration of TFPI, and PMNE might play a role in anticoagulative activity through TFPI during HD. Increasing or maintaining TFPI concentration may be a new target for anticoagulation and improving the biocompatibility of the hemodialysis membrane during hemodialysis, especially in ESRD patients with a high risk of bleeding complications.
Acknowledgments

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was approved by the Research Ethics Committee of West China Hospital of Sichuan University (2016-188), and informed consent was obtained from all participants. The authors had full access to all of the data in this study and take complete responsibility for the integrity of the data and the accuracy of the data analysis.

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