Epidemiological, biological and clinical update on exercise-induced hemolysis

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Abstract: Exercise-induced hemolysis can be conventionally defined as rupture and destruction of erythrocytes during physical exercise. The currently available epidemiologic information attests that a substantial degree of exercise-induced hemolysis is commonplace after short-, medium-, long- and ultra-long distance running, as reflected by significant decrease of serum or plasma haptoglobin and significant increase of plasma concentration (or overall blood content) of free hemoglobin. This paraphysiological intravascular hemolysis is typically mild (average variations of hemolysis biomarkers are usually comprised between 1.2- and 1.8-fold), almost self-limiting (completely resolving within 24–48 hours), with severity depending on athlete population, analytical technique used for detecting intravascular hemolysis, as well as on number, frequency and intensity of ground contacts, but not on running technique. Additional lines of evidence support the notion that both osmotic fragility and membrane structure of erythrocytes are considerably modified during endurance exercise. This fact goes hand in hand with findings that erythrocyte lifespan in runners is approximately 40% shorter than in sedentary controls. Direct mechanical injury caused by forceful ground contacts, repeated muscle contractile activity or vasoconstriction in internal organs are three potential sources of exercise-induced hemolysis, whilst metabolic abnormalities developing while exercising (e.g., hyperthermia, dehydration, hypotonic shock, hypoxia, lactic acidosis, shear stress, oxidative damage, proteolysis, increased concentration of catecholamines and lyssolecithin) may actively contribute to trigger, accelerate or amplify this phenomenon. Although no systematic evidence is available, it seems also reasonable to hypothesize that patients bearing erythrocyte disorders may be particularly vulnerable to developing exercise-induced hemolysis.

Keywords: Hemolysis; red blood cells (RBC); hemoglobin; sports, physical exercise

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Introduction

Hemolysis is conventionally defined as a process of rupture and destruction of erythrocytes occurring either within (e.g., in vivo, also known as “intravascular hemolysis” or “hemolytic anemia”) or outside (i.e., in vitro, also known as “spurious hemolysis”) the circulation (1). Although the “normal” (i.e., physiological) concentration of free hemoglobin is typically comprised within 0.22–0.25 g/L in serum and between 0.10–0.13 g/L in plasma (2), the definition of hemolysis is usually reserved to conditions when serum or plasma samples contain a cell-free hemoglobin concentration >0.5 g/L and display a pink to slightly red color. When the concentration of cell-free
hemoglobin exceeds 2–3 g/L, it is frequently referred to as “gross” or “frank” hemolysis, and is accompanied by red to brown color of serum or plasma, depending on final amount of protein (1).

The potential causes of intravascular hemolysis are many and multifaceted, and can be basically differentiated between congenital (e.g., hemoglobinopathies, disorders of erythrocyte membrane or metabolism) or acquired (e.g., immune-mediated, severe trauma or burns, hypersplenism, infections, mechanical intravascular injuries, incompatible blood transfusion or toxic) (1). Laboratory testing plays a substantial role, wherein intravascular hemolysis is conventionally diagnosed by some simple but straightforward findings, encompassing increased concentration of free hemoglobin in serum or plasma (as a direct index of erythrocyte destruction) (3), reduced values of haptoglobin (haptoglobin binds hemoglobin into the circulation to generate haptoglobin-hemoglobin complexes, which are then cleared by binding to specific receptors on macrophage surface) (4). Additional tests can be used for supporting the diagnosis of hemolytic anemia, such as the measurement of biomarkers of cytolysis (i.e., lactate dehydrogenase, aspartate aminotransferase, unconjugated bilirubin, potassium) or bone marrow stimulation (i.e., reticulocytes) (1), but they are plagued by an expectedly lower specificity.

According to the most recent statistics of Statista, one of the widest online statistics portals (5), running is the most popular and practiced sports worldwide, with as many as 60 million people engaged in some forms of jogging or running in the US (~18.4% of the entire resident population), and nearly half of these covering over 42 km each week. There is now incontrovertible evidence that a physically active lifestyle determines immense health benefits by lowering the risk of severe and frequently disabling pathologies such as cardiovascular disease, diabetes, osteoporosis, sarcopenia, frailty, and cancer (6,7). On the other hand, the regular practice of physical exercise may also have some unfavorable consequences such as sports injuries involving muscles, tendons and ligaments, accidental falls, bone fractures, cardiac arrhythmias, up to sudden death (8-10). Not only the musculoskeletal system may be injured by unaccustomed, incorrectly performed or extreme exercising, whereby several other organs and tissues may be affected, especially kidneys (11), liver (12) and bladder (13). Alongside the identification of possible unfavorable biological consequences, the main purposes of this article are: (I) establishing whether endurance exercise may be associated with intravascular hemolysis; (II) defining the possible determinants of exercise-induced hemolysis; and (III) reviewing the possible physiopathological mechanisms by which endurance exercise may promote, trigger or amplify intravascular hemolysis. In order to respond to these important questions, we have carried out a large free literature search for identifying all relevant studies that have previously addressed this problem and have summarized them in the following section of this narrative review.

Epidemiologic association between endurance exercise and intravascular hemolysis

The first case of probable exercise-induced hemolysis was described in 1884 by Kast (14). The German physician reported the case of a 19-year-old patient who developed gross hemoglobinuria after prolonged marches. Notably, he also observed that the lower were the periods of rest between consecutive marches, the shorter was the distance necessary to trigger hemoglobinuria. More than 30 years later, in 1916, the English Captain Macmanus described three cases of young men in early stage of military training who also developed gross hemoglobinuria after marching (15). The urine of the three young men appeared dark brown, with only few red blood cells (RBC). Spectroscopic analysis revealed that the color of the urine was attributable to large presence of methemoglobin. Notably, Macmanus attributed that unusual phenomenon to a combined effect of exercise and low temperature to which all the three cadets had been exposed. These two original reports then paved the way to the publication of a very large series of studies aimed at investigating the real burden of exercise-induced intravascular hemolysis.

The very first structured investigation on the relationship between exercise and hemolysis was published in 1943 by Gilligan and colleagues (16). The authors studied 11 male subjects who completed a cross-country run of 4.2–4.5 km, 11 male subjects who completed a cross-country run of 7.2–8.2 km and 22 male subjects who completed a long distance run of 42.2 km. An additional subject underwent a marathon, a 5-hour exercise on a cycle ergometer and a 421.6-km walk with a 24 kg pack on his back. Overall, 5/11 subjects (45.4%) displayed clearly reddish plasma after the 4.2–4.5 km (mean value, 0.14±0.13 g/L) and 7.2–8.2 km cross-country runs (mean value, 0.14±0.13 g/L), whilst this percentage increased to 81.8% (18/22) after the 421.6-km walk. A large series of studies showed that the percentage of subjects with hemoglobinuria increased significantly after exercise exceeding 8 km/h (17,18). Notably, the results of this series of investigations allowed to distinguish between two types of hemolysis depending on duration and intensity of the exercise: (I) low intensity exercise-induced hemolysis; and (II) high intensity exercise-induced hemolysis. The potential causes of intravascular hemolysis are many and multifaceted, and can be basically differentiated between congenital (e.g., hemoglobinopathies, disorders of erythrocyte membrane or metabolism) or acquired (e.g., immune-mediated, severe trauma or burns, hypersplenism, infections, mechanical intravascular injuries, incompatible blood transfusion or toxic) (1). Laboratory testing plays a substantial role, wherein intravascular hemolysis is conventionally diagnosed by some simple but straightforward findings, encompassing increased concentration of free hemoglobin in serum or plasma (as a direct index of erythrocyte destruction) (3), reduced values of haptoglobin (haptoglobin binds hemoglobin into the circulation to generate haptoglobin-hemoglobin complexes, which are then cleared by binding to specific receptors on macrophage surface) (4). Additional tests can be used for supporting the diagnosis of hemolytic anemia, such as the measurement of biomarkers of cytolysis (i.e., lactate dehydrogenase, aspartate aminotransferase, unconjugated bilirubin, potassium) or bone marrow stimulation (i.e., reticulocytes) (1), but they are plagued by an expectedly lower specificity.

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the long-distance run (mean plasma hemoglobin value, 0.14±0.09 g/L). No correlation was found between plasma hemoglobin and age or training status. The concentration of hemoglobinuria, measured with benzidine reaction, was negative in all post-exercise urine samples after the 4.2–4.5 km cross-country run, was positive in 1 of the 8 subjects (12.5%) who provided urine after the 7.2–8.2 km cross-country run, and was positive in 4 of the 22 (18.2%) of those who completed the long-distance run.

In a subsequent investigation, Flatmark studied a 54-year-old man with previous history of exercise-induced hemoglobinuria (17). He was asked to exercise on a treadmill at 2 km/h for 180 min, at 4 km/h for 180 min or on a cycle ergometer for 120 min (17). Notably, plasma hemoglobin very modestly increased after exercising on treadmill at 2 km/h for 180 min or on the cycle ergometer, whilst it increased by over 6-fold after exercising on treadmill at 4 km/h.

An interesting study was published in 1964 by Davidson (18). Four subjects were asked to complete a 4.8 km run on two different grounds, i.e., grass or road. The concentration of plasma hemoglobin increased significantly after running on both types of grounds, but the relative increase was much higher after running on road (i.e., 6.0±4.8 folds) than on grass (i.e., 1.4±0.2 folds). Hemoglobinuria was absent in all four urine samples after grass running, whilst it was present in 1/4 (25%) urine samples after road running.

Some years afterwards, in 1979, Poortmans and Haralambie designed a comprehensive study based on assessment of a large number of biochemical parameters in 11 male athletes who were engaged in 100-km running (19). The values of serum hemoglobin consistently increased from 0.03±0.01 to 0.7±0.02 g/L at the end of the run. Serum haptoglobin values were instead found to be dramatically decreased by nearly 2.5-fold (i.e., from 1.19±0.65 to 0.47±0.45 g/L; P<0.001) immediately after the run, and remained significantly decreased compared to baseline (0.99±0.38 g/L; P<0.05) the day after the run.

Dufaux et al. carried out a cross-sectional study, including 62 male sedentary controls, 81 male professional rowers, 61 male middle- and long-distance runners, and 52 male professional cyclists (20). The concentration of serum haptoglobin was 1.0±0.3 g/L in the sedentary controls and was non-significantly different from that of the rowers (1.2±0.4 g/L) or professional cyclists (1.1±0.2 g/L), whilst that of the runners was significantly lower compared with the other groups (0.7±0.4 g/L; P<0.01).

Hunding and coworkers measured plasma hemoglobin and haptoglobin in 3 trained distance runners who performed a 25-km run (21). Plasma hemoglobin increased in all three subjects from 0.03±0.03 to 0.43±0.13 g/L (i.e., 12-fold increase), whilst plasma haptoglobin remained almost unchanged. All three subjects had plasma hemoglobin values over the upper reference limit (URL) (>0.2 g/L) at the end of the run.

Falsetti et al. carried out another interesting study, including 23 male marathon runners, who were randomly divided in two groups according to the type of shoes worn for running (i.e., 11 subjects wearing firm-sole and 12 subjects wearing soft air-sole shoes) (22). Plasma hemoglobin and haptoglobin concentration measured before and after a 24-km run were 0.05±0.04 vs. 0.08±0.08 g/L and 0.45±0.47 vs. 0.34±0.46 g/L in the firm-sole shoe group, thus exhibiting a 1.37-fold increase and a 1.34-fold decrease, respectively. Conversely, plasma hemoglobin and haptoglobin concentration measured before and after a 24-km run were 0.03±0.01 vs. 0.04±0.01 g/L and 0.50±0.24 vs. 0.47±0.28 g/L in the firm-sole shoe group, thus exhibiting a more limited 1.15-fold increase and 1.06-fold decrease, respectively.

Eichner performed an interesting experiment, in which a middle-aged man was regularly monitored after his training periods by means of plasma haptoglobin measurements (23). Notably, a strong inverse correlation was found between running distance and variation of this biomarker (r=−0.91).

Nyman studied 19 runners (16 men and 3 women) who ran on average 16.1 km/day each for covering a total distance of 1,207 km (24). Before the start of the trial, plasma haptoglobin levels were lower than the URL in 5/19 athletes (26.3%), but in 15 of them (78.9%) the values progressively decreased during the run.

Davidson et al. measured plasma haptoglobin in 115 runners (90 men and 25 women) before and immediately after the end of a competitive marathon (25). In both male (0.80±0.65 vs. 1.15±0.66 g/L; P<0.001) and female (0.62±0.51 vs. 1.14±0.56 g/L; P<0.001) athletes, plasma haptoglobin values significantly declined at the end of the run, by 1.4- and 1.8-fold, respectively.

A similar study was carried out by Egan et al., who measured serum haptoglobin values in 8 male athletes before and after a marathon race (26). As predictable, the mean serum haptoglobin values substantially decreased by approximately 1.3-fold immediately after the run (0.97±0.48 vs. 1.29±0.18 g/L; P<0.05), and were found to be further
Wolf and colleagues studied 11 male prominent middle-distance runners who completed a 5-km run at their maximal effort (27). The serum haptoglobin value was found to be reduced by approximately 4.7-fold at the end of the trial (0.20±0.02 vs. 0.95±0.08 g/L; P<0.03).

Lijnen et al. investigated the variation of cell breakdown biomarkers in the plasma of 23 male runners who were engaged in a marathon race (28). Plasma haptoglobin values decreased from 1.20±0.15 g/L before the run to approximately 0.75±0.15 g/L immediately after, remained decreased during 12 hours (0.85±0.15 g/L), and only returned to the baseline pre-run values 72 hours after the end of the marathon.

Miller et al. planned an interesting experiment for exploring the potential association between foot impact force and changes of hemolysis biomarkers in 14 male distance runners (29). The athletes completed two treadmill run at 12.9 km/h for a total number of 10,000 foot-strikes at two different elevations (i.e., -6% or +6%; force +11% higher running downhill vs. uphill). Plasma hemoglobin increased from 0.023±0.010 to 0.051±0.023 g/L (i.e., 2.2-fold increase) and from 0.037±0.023 to 0.086±0.051 g/L (i.e., 2.3-fold increase) after running uphill and downhill, respectively. Likewise, plasma haptoglobin decreased from 0.42±0.28 to 0.37±0.27 g/L (1.1-fold decrease) and from 0.42±0.30 to 0.33±0.29 g/L (i.e., 1.3-fold decrease), respectively.

O’Toole and colleagues carried out a large study based on 95 athletes who participated in two triathlon races of different distances (30). Thirty athletes (11 men and 19 women) first completed a 1.5-km swimming, 40-km cycling and 10-km running distance, whilst 65 other athletes (46 men and 19 women) completed a 3.9-km swimming, 180-km cycling and 42.2-km running distance. Overall, 95% of all runners displayed a decrease of serum haptoglobin immediately after the races, with a mean 1.28-fold decrease. Although the percentage of athletes with decreased haptoglobin values was nearly similar between the two races (i.e., 95% vs. 93%), the relative haptoglobin decrease was higher in athletes who completed the longer race (i.e., 32% vs. 20%). The percentage of athletes with occult blood in urine was also higher after the longer race (31% vs. 23%).

Seiler et al. studied 110 well-trained athletes (91 men and 19 women) who enrolled in a 1000-km running competition, lasting for 20 days (average running per day, 50 km) (31). The authors measured serum haptoglobin in 51 of these subjects at different time points and found that mean serum haptoglobin value was systematically lower than the baseline until the 11th running day (the lowest values were found after the first day with a nearly 1.5-fold decrease), whilst the concentration returned to pre-run values at the end of the trial. Even more importantly, although only 5% of urine samples were positive for blood before the start of the run, the percentage of positive samples for blood was always >25% throughout the study period (the peak was 34.5%, as recorded at the end of the competition).

Deitrick carried out a study including 15 male recreational runners in whom circulating haptoglobin and urobilinogen levels were measured before, immediately after, 1 day, 4 days, and 10 days after a 13-km run (32). Immediately after the run serum haptoglobin non-significantly decreased from 0.92±0.69 to 0.89±0.75 g/L, whilst a statistically significant reduction was then noted 24 hours after the end of the run (0.66±0.66 g/L; P<0.05), with values remaining significantly decreased up to 4 days after the run (0.59±0.53 g/L; P<0.05). The values of urobilinogen were also significantly increased at 1 and 24 hours after the run.

Dressendorfer et al. investigated the impact of 7 consecutive days of prolonged jogging (2 h per day at ~80% of maximal heart rates) in 10 moderately fit men, who finally covered a total distance of 129 km (33). The mean haptoglobin level decreased from 0.86 to 0.67 g/L on day 5 (P<0.05), and further decreased to 0.61 g/L (P<0.05) at day 8.

Weight et al. studied 20 male runners who completed a 42-km marathon (34). The plasma hemoglobin value was found to be substantially increased immediately after the run (0.113±0.074 vs. 0.077±0.050 g/L), whilst the serum concentration of haptoglobin significantly decreased (0.69±0.40 vs. 0.89±0.40 g/L). The values of both plasma hemoglobin and haptoglobin returned to the baseline after 24 hours. Interestingly, the authors also showed that the mean RBC lifespan in distance runners (i.e., 67 and 72 days in men and women, respectively) was considerably shorter than in sedentary controls (i.e., 113 and 114 days in men and women, respectively).

In another interesting investigation, Dressendorfer and colleagues (35) randomly divided a group of 14 male runners into those wearing firm-sole or soft-sole shoes. All athletes were then asked to undergo increased distance training, completing 430 km in 17 days. Plasma haptoglobin decreased more in the soft-sole (from 0.55±0.09 to 0.43±0.07 g/L; -27%) than in the firm-sole (from...
studied 13 male runners who engaged in a 100-km run (36). Serum haptoglobin considerably decreased from 0.66±0.18 g/L at baseline to 0.22±0.05 g/L immediately after the run.

Jordan et al. studied 13 male runners who engaged in a marathon race (37). Plasma haptoglobin decreased from approximately 1.2±0.8 g/L before the start of the marathon to 0.7±0.8 g/L immediately after. Another important aspect emerged from this study was that long-distance running induced significant RBC membrane skeleton modifications, probably due to a process of in vivo proteolysis.

An extreme exercise study was planned by Fallon et al. (38), who recruited 9 athletes (7 men and 2 women) participating in a 1,600-km ultramarathon. At variance with all the other studies, the authors found a substantial increase of serum haptoglobin from baseline (1.8±0.7 g/L) throughout (3.4±0.9 and 3.9±0.6 g/L) and after the run (4.1±1.2 g/L).

Schumacher et al. carried out another interesting study aimed at investigating several hematologic biomarkers in athletes of different sport disciplines (39). Interestingly, although no significant differences were observed in the concentration of serum haptoglobin between the whole cohorts of athletes (n=747; 0.67±0.37 g/L) and sedentary people (n=104; 0.67±0.40 g/L), runners (n=144; 0.50±0.35 g/L) displayed a substantially lower value than cyclists (n=272; 0.80±0.34).

Telford and coauthors published an interesting experiment, in which 10 male triathletes completed two different 1-hour sessions of cycling and running at 75% maximal oxygen consumption (40). Plasma hemoglobin values were found to be significantly increased after both the cycling (0.049±0.01 vs. 0.030±0.01) and running (0.120±0.02 vs. 0.037±0.01 g/L) sessions, but the relative increase was nearly double after running (3.2-fold vs. 1.6-fold). Haptoglobin values followed an inverse trend, with a much higher decrease after running than after cycling (the mean changes were 0.016 and 0.085 g/L after cycling and running, respectively).

Yusof et al. studied 6 male runners who engaged in a 216-km ultra-endurance race (41). Serum haptoglobin, which was measured at baseline (0.87±0.24 g/L) and at different time points throughout the run, showed a sharp decrease after 21 km (0.71±0.19 g/L) and 42 km (0.59±0.19 g/L), reaching its lowest value after 84 km (0.43±0.18 g/L). An opposite trend was then noted afterwards, with values slightly increasing after 126 km (0.46±0.27) and more markedly growing at the end of the run (0.64±0.34), thought remaining lower than at baseline. Interestingly, the authors also found that osmotic fragility was dramatically increased throughout the running distance.

Peeling et al. investigated the effect of two different training surfaces (i.e., grass or bitumen road) in 10 trained male runners, who performed two 10-km runs and ten 1-km interval running sessions (42). Interestingly, serum hemoglobin increased from 0.041±0.003 to 0.049±0.003 g/L after grass running (i.e., 19% increase) and from 0.032±0.003 to 0.04±0.002 g/L after road running (i.e., 40% increase), respectively. Plasma haptoglobin decreased from 0.58±0.12 to 0.51±0.11 g/L after grass running (i.e., 14% decrease) and from 0.66±0.13 to 0.58±0.12 g/L after road running (i.e., 14% decrease), respectively. The same group of authors also highlighted in another publication that the hemolysis degree was substantially similar after the 10-km run and the ten 1-km interval running sessions (both displaying a 37% increase of serum hemoglobin) (43).

Interesting findings have also been published by Sim et al., who measured serum hemoglobin and haptoglobin in 11 male athletes undergoing two 90-min running session at 75% of peak oxygen uptake, consuming either a 6% carbohydrate solution or placebo (44). The values of serum hemoglobin were found to be significantly increased decreased after both sessions, whilst those of haptoglobin were significantly decreased after both sessions. Notably, carbohydrate intake was effective to reduce the hemoglobin increase (1.60-fold vs. 1.86-fold for placebo) but not the haptoglobin decrease (1.27-fold versus 1.20-fold for placebo).

Gough et al. studied 18 male athletes who participated in an endurance triathlon race (3.8-km swimming, 180-km cycling and 42.2-km running distance) (45). Serum haptoglobin levels immediately after the race were substantially decreased compared to baseline (0.16 vs. 0.48 g/L).

Lippi and colleagues measured serum haptoglobin in 18 male runners who completed a 60-km ultramarathon (46), and found a substantial decrease of its values immediately after the end of the run (0.36 vs. 0.68 g/L).

Binnie et al. performed two separate investigations to assess the effect of different training surfaces on several biochemical parameters in 10 male athletes. In the first
Caulfield et al. studied 19 male runners who engaged in 8×3 min motorized treadmill exercises at 90% of maximal oxygen consumption (52). A significant reduction of serum haptoglobin was noted at the end of the trial, which was overall comparable among fore-foot (n=10; −9.3%) and rear-foot (n=9; −10.6%; P=0.902) athletes.

Finally, Liu et al. studied 19 male athletes who participated in a 24-hour ultramarathon (median distance covered, 154 km) (53). Unlike the vast majority of other studies, plasma hemoglobin concentration was found to be modestly decreased rather than increased at the end of the run (i.e., 0.2 vs. 0.3 g/L; P<0.001), a finding which was actually at odds with the remarkable decrease of serum haptoglobin contextually observed in the same cohort of athletes (0.19 vs. 0.77 g/L; P<0.001).

**Possible causes of exercise-induced hemolysis**

Once clearly established that exercise-induced hemolysis is a frequent phenomenon in athletes engaged in endurance exercise (especially runners), it seems reasonable to classify the potential (and often coexisting) causes into two leading categories, thus encompassing mechanical and metabolic factors (Figure 1). Indeed, a certain degree of direct erythrocyte injury cannot be denied as logic consequence of repeated foot impacts (especially heel strikes) on the ground. This is clearly deducible from studies showing that running on soft surfaces (i.e., on sand or grass) (18,42,47,48) or wearing cushioned shoe (i.e., soft air-sole shoes) (22) is sometimes effective to reduce but not completely neutralize exercise-induced hemolysis, as well as by other investigations demonstrating that the degree of hemolysis was dependent on intensity (29) or overall number (long distance or accelerated stepping cadence) (16,23,30) of ground impacts. On the other hand, reliable evidence attests that additional mechanisms other than foot strike may be involved in exercise-induced hemolysis. For example, Beneke et al. studied 10 male triathletes who engaged in two sessions of 35-min low- and high-intensity cycling, which does not involve any possible mechanical RBC trauma at foot strike (54). Interestingly, although low intensity cycling triggered a modest increase of plasma haptoglobin (from 1.30 to 1.49 g/L), high intensity cycling elicited a modest but significant reduction of this biomarker (from 1.10 to 1.01 g/L), confirming that even high intensity exercise per se is capable to trigger a certain degree of hemolysis independently from injuries caused by foot hitting the ground. Similar evidence has been...

experiment the athletes completed two interval training sessions, one on soft dry beach sand and the other on well-maintained sporting grass ground (47). Each session consisted of three interval sets separated by 5 min of rest of dynamic running (2×45:90 s, 3×20:60 s and 2×15:45 s). After both sessions, the concentration of serum haptoglobin was found to be significantly decreased, though the absorptive qualities of sand were effective to slightly attenuate haptoglobin reduction compared to the grass (1.08-fold decrease in sand vs. 1.12-fold decrease in grass, respectively). The design of the second part of the experiment was more or less similar to the former, with two repeated sessions of sprint bouts, agility and power drills involving rapid changes of direction and speed (48). As in the former experiment, one session was performed on soft dry beach sand and the other on well-maintained sporting grass ground. Unlike previous findings, and although the concentration of serum haptoglobin was found to be significantly decreased after both sessions, such decrease was also partially attenuated by the absorptive qualities of sand (1.14-fold decrease in sand vs. 1.20-fold decrease in grass, respectively).

In a subsequent investigation Christensen et al. studied 10 male runners who took part in a 78-km race developing at 2,400 m above the sea level (49). Plasma haptoglobin values dramatically decrease from 1.48±0.30 g/L pre-run, to 0.74±0.33 g/L immediately after the run, and remained so up to 6 hours afterwards. Haptoglobin concentration returned to pre-run values after 24 hours.

Another interesting study was published by Robach et al. (50), who examined biochemical changes in 22 male runners after a 166-km ultra-marathon with 9,500 m of altitude gain/loss. Plasma hemoglobin increased from 0.12±0.57 g/L before the run to 0.14±0.85 immediately after, whilst serum haptoglobin concentration was found to be significantly decreased immediately after the end of the run (0.74±033 vs. 0.87±0.34 g/L). Notably, the values of both plasma hemoglobin and serum haptoglobin normalized the following day after the run. Interestingly, the total plasma hemoglobin content of blood was also found to be increased by nearly 1.4-fold at the end of the run (570±363 vs. 395±187 mg).

Chiu et al. studied 25 male athletes who completed a 100-km ultramarathon (51). Plasma hemoglobin was found to be slightly increased after the end of the run (from 0.3±0.2 to 0.4±0.3 g/L), whilst serum haptoglobin was considerably decreased (from 0.64±0.31 to 0.30±0.30). Both values normalized 24 hours after the end of the run.

### Possible causes of exercise-induced hemolysis

Once clearly established that exercise-induced hemolysis is a frequent phenomenon in athletes engaged in endurance exercise (especially runners), it seems reasonable to classify the potential (and often coexisting) causes into two leading categories, thus encompassing mechanical and metabolic factors (Figure 1). Indeed, a certain degree of direct erythrocyte injury cannot be denied as logic consequence of repeated foot impacts (especially heel strikes) on the ground. This is clearly deducible from studies showing that running on soft surfaces (i.e., on sand or grass) (18,42,47,48) or wearing cushioned shoe (i.e., soft air-sole shoes) (22) is sometimes effective to reduce but not completely neutralize exercise-induced hemolysis, as well as by other investigations demonstrating that the degree of hemolysis was dependent on intensity (29) or overall number (long distance or accelerated stepping cadence) (16,23,30) of ground impacts. On the other hand, reliable evidence attests that additional mechanisms other than foot strike may be involved in exercise-induced hemolysis. For example, Beneke et al. studied 10 male triathletes who engaged in two sessions of 35-min low- and high-intensity cycling, which does not involve any possible mechanical RBC trauma at foot strike (54). Interestingly, although low intensity cycling triggered a modest increase of plasma haptoglobin (from 1.30 to 1.49 g/L), high intensity cycling elicited a modest but significant reduction of this biomarker (from 1.10 to 1.01 g/L), confirming that even high intensity exercise per se is capable to trigger a certain degree of hemolysis independently from injuries caused by foot hitting the ground. Similar evidence has been...
published in other nontraumatic sport disciplines such as endurance swimming (55). The repeated contractile activity of muscles during exercise is then another mechanism which may cause erythrocyte mechanical injury, whereby RBC compression during powerful and repeated muscle contractions (especially within the capillary network) may finally result in their rupture. Sustained vasoconstriction of internal organs (especially kidneys), a physiological process aimed at deviating large part of blood flow into the exercising muscles, is another frequent phenomenon in endurance running (21,56), which may contribute to generate RBC compression and injury in smaller arteries.

Beside direct mechanical injuries, a kaleidoscope of other metabolic causes and mechanisms occurring while exercising may promote enhanced erythrocyte fragility and/or impaired membrane structure, deformability or resistance (especially for older RBC), finally contributing to promote or increase the chance of intravascular destruction (Figure 1). Enhanced catecholamines values, as commonplace in sports, interplay with specific receptors at RBC surface, cause impaired membrane deformability and ultimately promote higher vulnerability to hemolysis (57). A parallel mechanism has been described by Yamada et al., who showed that exercise adaptation induces a significant change of lipid profile, accompanied by an increase of lysolecithin and a reduction of free cholesterol in the erythrocyte membrane, two converging phenomenon which contribute to enhance osmotic fragility (58). The hypothesis that RBC membrane may be altered during exercise has been confirmed by Beneke et al. (54), who identified abnormalities in alpha- and beta-spectrin comparable to those found in congenital erythrocyte disorders (e.g., spherocytosis). This hypothesis has been further confirmed by others (41). Hyperthermia, which is commonplace in athletes engaged in strenuous and/or prolonged exercise, significantly enhance erythrocyte fragility and thereby their propensity to rupture (59). Acidosis is another important mechanism which may at least partially explain the association between longer distance and higher propensity to develop hemolysis. An increased production of lactic acid is frequent in endurance
athletes, and the osmotic fragility of erythrocytes has been recently shown to increase in parallel with blood lactate concentration (60). Hypoxia, hypotonic shock and shear stress are additional conditions frequently developing in exercising muscles, and recent evidence has been provided that they may altogether contribute to strongly enhance ATP efflux from RBC, thus ultimately escalating their susceptibility to intravascular hemolysis (61). Reliable evidence has also been provided that exercise-induced dehydration may contribute to increase erythrocyte fragility in vivo, as demonstrated by Platt et al. (62). Smith et al. also showed that strenuous running is associated with a remarkable reduction of erythrocyte antioxidant capacity, making these cells much more vulnerable to lysis (63).

Although no systematic evidence is available in the scientific literature, it seems reasonable to hypothesize that patients bearing some underlying erythrocyte disorders causing increased osmotic fragility may be more vulnerable to developing exercise-induced hemolysis (Figure 1). Some paradigmatic cases have been described, such as those of patients with hereditary spherocytosis (64), sickle cell disease (65) or glucose-6-phosphate dehydrogenase (G6PD) deficiency (66), among others. Beside subjects carrying inherited or acquired erythrocyte disorders, which would increase per se the baseline risk of intravascular hemolysis, there are other conditions associated with enhanced erythrocyte fragility in vivo and may contribute to magnify the likelihood of exercise-induced hemolysis. These may actually include hyponatremia (67), diabetes (68) and chronic liver disease (69).

Conclusions

According to different pathogenetic mechanisms, the classical definition of foot-strike hemolysis seems now inappropriate for identifying all cases of intravascular hemolysis occurring while exercising. This is due to the fact that several studies showed that attenuating RBC injuries derived from ground contact is not completely effective for preventing intravascular hemolysis. Therefore, we suggest that foot-strike (or contact) hemolysis shall now be considered only a part of the more thoughtful and appropriate concept of exercise-induced hemolysis.

The literature data reviewed in this narrative review attests that a significant degree of exercise-induced hemolysis is commonplace after short-, medium-, long- and ultra-long distance running, as reflected by the significant decrease of serum or plasma haptoglobin combined with the significant increase of plasma hemoglobin concentration or overall blood content (Table 1). This paraphysiological intravascular hemolysis is typically mild (the average variations of hemolysis biomarkers are usually comprised between 1.2- and 1.8-fold), almost self-limiting (i.e., completely resolving within 24–48 hours), with relative extent depending on athlete population, analytical technique used for detecting intravascular hemolysis as well as on number, frequency and intensity of ground contacts, but not on running technique (i.e., fore-foot or rear-foot) (Table 1). Additional lines of evidence support the notion that both osmotic fragility and membrane structure of erythrocytes are considerably modified during endurance exercise (41), and this fact goes hand in hand with findings that erythrocyte lifespan in runners is approximately 40% shorter than in sedentary controls (e.g., 70 vs. 114 days) (34). Direct mechanical injury caused by forceful ground contacts, repeated muscle contractile activity or vasoconstriction in internal organs are three potential sources of exercise-induced hemolysis, whilst preexisting erythrocyte disorders and metabolic abnormalities developed while exercising (e.g., hyperthermia, dehydration, hypotonic shock, hypoxia, lactic acidosis, shear stress, oxidative damage, proteolysis, increased concentration of catecholamines and lysolcithin) may actively contribute to trigger, accelerate or amplify this phenomenon (Figure 1).

Regarding the possible clinical significance of mild and transitory intravascular hemolysis in athletes, an interesting study has revealed that cell-free hemoglobin levels are inversely correlated with mortality and hospital admissions for heart failure (i.e., each 0.1 g/L increase in plasma hemoglobin reduces the risk of mortality and heart failure-related hospital admissions of 40% and 21%, respectively) (71). Unlike severe hemolytic diseases, where massive release of cell–free hemoglobin overwhelms the potency of the homeostatic clearance systems and directly triggers a severe oxidative injury (72), the much lower concentration of cell–free hemoglobin generated from exercise-induced hemolysis is completely cleared by scavenger plasma proteins, especially haptoglobin. Within these complexes, the hemoglobin-related pseudoperoxidase activity is then transformed from detrimental into protective, ultimately contributing to safeguard cell integrity and offering protection to exercise-intensified oxidative stress (Figure 2) (73). This mechanism may hence be seen as a potentially beneficial pathway, which may ultimately contribute to amplify the many benefits of regular physical exercise in reducing the risk of morbidity, frailty and mortality (74–76).
Table 1 Summary of studies which have explored the frequently or extent of exercise-induced hemolysis in runners

<table>
<thead>
<tr>
<th>Authors</th>
<th>Study population</th>
<th>Type of study</th>
<th>Distance</th>
<th>Methods</th>
<th>Hemolysis after exercise</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gilligan et al., 1943</td>
<td>(I) 11 men; (II) 11 men; (III) 22 men</td>
<td>Prospective</td>
<td>(I) 4.2–4.5 km; (II) 7.2–8.2 km; (III) 42.2 km</td>
<td>Plasma hemoglobin</td>
<td>Hemoglobin &gt;URL in (I) 12.5%, (II) 12.5% and (III) 81.8%</td>
</tr>
<tr>
<td>Flatmark, 1963</td>
<td>1 man</td>
<td>Prospective</td>
<td>(I) Tread-mill at 2 km/h for 180 min; (II) treadmill at 4 km/h for 180 min; (III) cycle ergometer for 120 min</td>
<td>Plasma hemoglobin</td>
<td>Increase of hemoglobin by 6-fold only after treadmill at 4 km/h for 180 min</td>
</tr>
<tr>
<td>Davidson, 1964</td>
<td>4 men</td>
<td>Prospective</td>
<td>4.8 km in (I) grass or (II) road</td>
<td>Plasma hemoglobin</td>
<td>Increase of hemoglobin by 1.4- and 6-fold after grass and road running, respectively</td>
</tr>
<tr>
<td>Poortmans et al., 1979</td>
<td>11 men</td>
<td>Prospective</td>
<td>100 km</td>
<td>Serum hemoglobin and haptoglobin</td>
<td>Increase of hemoglobin by ~2.2-fold and decrease of haptoglobin by ~2.5-fold, respectively</td>
</tr>
<tr>
<td>Dufaux et al., 1981</td>
<td>62 male sedentary controls, 81 male professional rowers, 61 male middle- and long-distance runners, and 52 male professional cyclists</td>
<td>Cross-sectional</td>
<td>Not provided</td>
<td>Serum haptoglobin</td>
<td>Haptoglobin 1.4- to 1.7-fold lower in runners than in other groups</td>
</tr>
<tr>
<td>Hunding et al., 1981</td>
<td>3 men</td>
<td>Prospective</td>
<td>25 km</td>
<td>Plasma hemoglobin</td>
<td>Increase of hemoglobin by ~12-fold</td>
</tr>
<tr>
<td>Falsetti et al., 1983</td>
<td>23 men (11 subjects wearing firm-sole and 12 subjects wearing soft air-sole shoes)</td>
<td>Prospective</td>
<td>24 km</td>
<td>Plasma hemoglobin and haptoglobin</td>
<td>Increase of hemoglobin by ~1.4-fold and decrease of haptoglobin by ~1.3-fold wearing firm-sole shoes; Increase of hemoglobin by ~1.15-fold and decrease of haptoglobin by ~1.06-fold wearing soft air-sole shoes</td>
</tr>
<tr>
<td>Eichner, 1985</td>
<td>1 man</td>
<td>Prospective</td>
<td>Training at different running distances</td>
<td>Plasma haptoglobin</td>
<td>Inverse correlation between running distance and haptoglobin</td>
</tr>
<tr>
<td>Nyman, 1985</td>
<td>19 (16 men and 3 women)</td>
<td>Prospective</td>
<td>1,207 km</td>
<td>Plasma haptoglobin</td>
<td>Haptoglobin decreased in 79% of the subjects</td>
</tr>
<tr>
<td>Davidson et al., 1987</td>
<td>115 (90 men and 15 women)</td>
<td>Prospective</td>
<td>42.2 km</td>
<td>Plasma haptoglobin</td>
<td>Decrease of haptoglobin by 1.4- and 1.8-fold in men and women, respectively</td>
</tr>
<tr>
<td>Egan et al., 1987</td>
<td>8 men</td>
<td>Prospective</td>
<td>42.2 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.3-fold</td>
</tr>
<tr>
<td>Wolf et al., 1987</td>
<td>8 men</td>
<td>Prospective</td>
<td>5 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 4.7-fold</td>
</tr>
<tr>
<td>Lijnen et al., 1988</td>
<td>23 men</td>
<td>Prospective</td>
<td>42.2 km</td>
<td>Plasma haptoglobin</td>
<td>Decrease of haptoglobin by 1.6-fold</td>
</tr>
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</table>

Table 1 (continued)
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<table>
<thead>
<tr>
<th>Authors</th>
<th>Study population</th>
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<th>Methods</th>
<th>Hemolysis after exercise</th>
</tr>
</thead>
<tbody>
<tr>
<td>Miller et al., 1988 (29)</td>
<td>14 men</td>
<td>Prospective</td>
<td>10,000 foot-strokes at (I) −6% or (II) +6% elevation</td>
<td>Plasma hemoglobin and haptoglobin</td>
<td>Increase of hemoglobin by ~2.2-fold and decrease of haptoglobin by ~1.1-fold running uphill; Increase of hemoglobin by ~2.3-fold and decrease of haptoglobin by ~1.3-fold running downhill</td>
</tr>
<tr>
<td>O'Toole et al., 1988 (30)</td>
<td>95 (57 men and 38 women)</td>
<td>Prospective</td>
<td>(I) 1.5-km swimming, 40-km cycling and 10-km running; (II) 3.9-km swimming, 180-km cycling and 42.2-km running</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.28-fold (1.20-fold after shorter and 1.32-fold after longer distance)</td>
</tr>
<tr>
<td>Seiler et al., 1989 (31)</td>
<td>110 (91 men and 19 women)</td>
<td>Prospective</td>
<td>1,000 km</td>
<td>Plasma haptoglobin</td>
<td>Decrease of haptoglobin by ~1.5-fold</td>
</tr>
<tr>
<td>Deitrick, 1991 (32)</td>
<td>15 men</td>
<td>Prospective</td>
<td>15 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.6-fold</td>
</tr>
<tr>
<td>Dressendorfer et al., 1991 (33)</td>
<td>10 men</td>
<td>Prospective</td>
<td>129 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.4-fold</td>
</tr>
<tr>
<td>Weight et al., 1991 (34)</td>
<td>20 men</td>
<td>Prospective</td>
<td>42.2 km</td>
<td>Plasma hemoglobin and serum haptoglobin</td>
<td>Increase of hemoglobin by 1.5-fold and decrease of haptoglobin by 1.3-fold</td>
</tr>
<tr>
<td>Dressendorfer et al., 1992 (35)</td>
<td>14 men (7 with firm-sole and 7 with soft-sole shoes)</td>
<td>Prospective</td>
<td>430 km</td>
<td>Plasma hemoglobin and haptoglobin</td>
<td>Increase of hemoglobin by 1.7- and 1.6-fold in firm-sole and soft-sole shoes groups; decrease of haptoglobin by 1.27- and 1.15-fold in firm-sole and soft-sole shoes groups</td>
</tr>
<tr>
<td>De Paz et al., 1995 (36)</td>
<td>13 men</td>
<td>Prospective</td>
<td>100 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 3-fold</td>
</tr>
<tr>
<td>Jordan et al., 1998 (37)</td>
<td>13 men</td>
<td>Prospective</td>
<td>42.2 km</td>
<td>Plasma haptoglobin</td>
<td>Decrease of haptoglobin by 1.7-fold</td>
</tr>
<tr>
<td>Schumacher et al., 2002 (70)</td>
<td>416 men (144 runners and 272 cyclists)</td>
<td>Cross-sectional</td>
<td>–</td>
<td>Serum haptoglobin</td>
<td>Haptoglobin 1.6-fold lower in runners than in cyclists</td>
</tr>
<tr>
<td>Telford et al., 2003 (40)</td>
<td>10 men</td>
<td>Prospective</td>
<td>1-hour session at 75% maximal oxygen consumption of (I) cycling and (II) running</td>
<td>Plasma haptoglobin</td>
<td>Increase of hemoglobin by 1.6- and 3.2-fold after cycling or and running, respectively</td>
</tr>
<tr>
<td>Yusof et al., 2007 (41)</td>
<td>6 men</td>
<td>Prospective</td>
<td>216 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 2-fold</td>
</tr>
</tbody>
</table>
Table 1 (continued)

<table>
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<tr>
<th>Authors</th>
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<th>Methods</th>
<th>Hemolysis after exercise</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peeling et al., 2009 (42,43)</td>
<td>10 men</td>
<td>Prospective</td>
<td>Two 10-km runs and ten 1-km interval running sessions on (I) grass and haptoglobin and (II) road</td>
<td>Serum hemoglobin</td>
<td>Increase of hemoglobin by 1.19- and 1.40-fold after grass and road running, respectively; decrease of haptoglobin by 1.14-fold after both surfaces running</td>
</tr>
<tr>
<td>Sim et al., 2012 (44)</td>
<td>11 men</td>
<td>Prospective</td>
<td>90-min run at 75% of peak oxygen uptake</td>
<td>Serum hemoglobin and haptoglobin</td>
<td>Increase of serum hemoglobin by 1.6- to 1.9-fold and decrease of serum haptoglobin by 1.2- to 1.3-fold, respectively</td>
</tr>
<tr>
<td>Gough et al., 2012 (45)</td>
<td>18 men</td>
<td>Prospective</td>
<td>3.8-km swim, 180-km cycling and 42.2-km run</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 3-fold</td>
</tr>
<tr>
<td>Lippi G et al., 2012 (46)</td>
<td>18 men</td>
<td>Prospective</td>
<td>60 km</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.9-fold</td>
</tr>
<tr>
<td>Binnie et al., 2013 (47)</td>
<td>10 men</td>
<td>Prospective</td>
<td>Interval training sessions on sand or grass</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.08-fold in sand and 1.12-fold in grass, respectively</td>
</tr>
<tr>
<td>Binnie et al., 2013 (48)</td>
<td>10 men</td>
<td>Prospective</td>
<td>Sprint bouts, agility and power drills on sand or grass</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.14-fold in sand and 1.20-fold in grass, respectively</td>
</tr>
<tr>
<td>Christensen et al., 2014 (49)</td>
<td>10 men</td>
<td>Prospective</td>
<td>78 km</td>
<td>Plasma haptoglobin</td>
<td>Decrease of haptoglobin by 2-fold</td>
</tr>
<tr>
<td>Robach et al., 2014 (50)</td>
<td>22 men</td>
<td>Prospective</td>
<td>166 km</td>
<td>Plasma hemoglobin, plasma hemoglobin content, and serum haptoglobin</td>
<td>Increase of hemoglobin concentration and content by 1.2- and 1.4-fold, respectively; decrease of haptoglobin by 1.2-fold</td>
</tr>
<tr>
<td>Chiu et al., 2015 (51)</td>
<td>25 men</td>
<td>Prospective</td>
<td>100 km</td>
<td>Plasma hemoglobin and serum haptoglobin</td>
<td>Increase of hemoglobin by 1.3-fold and decrease of haptoglobin by 2.1-fold</td>
</tr>
<tr>
<td>Caulfield et al., 2016 (52)</td>
<td>19 men</td>
<td>Prospective</td>
<td>8x3 min motorized treadmill exercises</td>
<td>Serum haptoglobin</td>
<td>Decrease of haptoglobin by 1.09- and 1.11-fold in fore-foot and rear-foot runners</td>
</tr>
<tr>
<td>Liu et al., 2018 (53)</td>
<td>19 men</td>
<td>Prospective</td>
<td>154 km</td>
<td>Plasma hemoglobin and serum haptoglobin</td>
<td>Decrease of hemoglobin and haptoglobin by 1.5- and 4-fold, respectively</td>
</tr>
</tbody>
</table>

URL, upper reference limit.
**Figure 2** Different consequences of severe and mild intravascular hemolysis.

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**Footnote**

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

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