Adult mesenchymal stem cells for the treatment in patients with rotator cuff disease: present and future direction

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Rotator cuff tendon tear (RCTT) is reported to be one of the most common causes of chronic shoulder pain in the United States with more than 250,000 RCTT repairs each year (1). Individuals under 60 years of age show a 6% incidence of full-thickness RCTT, while those over 60 show an incidence of 30%, as determined by cadaveric studies (2). RCTT results in considerable disability and diminished quality of life for many patients (3).

Chronic tendon degeneration is currently considered a major contributor to the pathology of RCTT (4). Treatment for RCTT includes conservative treatments such as exercise, electrotherapy, acupuncture, manual treatment, taping, steroid and platelet-rich plasma injections and surgical treatment (5). Surgical repair for tendon tear is one of the most common orthopedic procedures undertaken today (6).

The classification of RCTT using a reliable classification method is critical to improve the treatment of rotator cuff disease. This classification has been used to determine the appropriate treatments to achieve the best results for each type of tendon tear. Many factors such as tear size, shape, retraction, and fatty infiltration have been identified as important factors when assessing a patient with a symptomatic RCTT. RCTT are classified as either partial- or full-thickness. Partial-thickness tears are focal defects in the rotator cuff tendon and involve only bursal or articular surface or intratendinous. Full-thickness tears extend from the articular surface to the bursal surface of the tendon.

Ellman (7) proposed a classification system for partial- thickness RCTT based on intra-operative findings according to the location (bursal-side, articular-side, and intratendinous), grade, and tear area (mm²). In his classification, a grade 3 tear is >6 mm deep (>50%), a grade 2 tear is 3–6 mm in depth but not exceeding one-half of the tendon thickness (25–50%), and a grade 1 tear is considered as <3 mm deep (<25%). DeOrio and Cofield (8) developed classification systems for full-thickness RCTT, which involved measuring the anterior-posterior length of the tendon that was torn off of the humeral head at the time of surgery. Their system classified tendon tear size as massive if the tear was greater than 5 cm, large if the tear was 3–5 cm, medium if the tear was 1–3 cm, and small if the tear was less than 1 cm in length.

For acute tears, and in patients younger than 65 years, in the case of chronic, reparable tears larger than 1 cm and without significant chronic muscle changes (such as atrophy, fatty infiltration, and impairment of the biomechanical properties of the repair sites), early surgical repair can be considered. Early surgical repair can also be considered in large bursal-sided partial thickness tears (larger than 6 mm or more than 50% of the tendon thickness) because the risk of tear progression and rate of failure of conservative treatment is high (9).

However, the failure rate for rotator cuff tendon repair is 20–90% (10) and the outcome can vary depending on the patient’s age, tear size, chronicity, muscle atrophy, fatty degeneration, tendon quality, operative technique, and postoperative rehabilitation treatment (11). Revision rotator
cuff surgery has been less successful than primary repair (12). Current therapeutic approaches result in suboptimal quality at the distal end of the RCTT and a lack of physiological restoration; thus, a new biological treatment, which creates an optimized environment for the restoration of the tendon, is needed to regenerate the tendon and reduce scar formation (13).

The paper being referenced in this editorial [published in the Stem Cells on May 11, 2018 (14)] shows that injecting autologous adipose tissue-derived mesenchymal stem cells (ADMSCs) into the tendon without surgery for the treatment of rotator cuff disease is feasible, safe, and effective to regenerate of tendon defect being seen.

Tissue engineering approaches are now able to target and generate almost any cell of the body. Stem-cell-based research has grown exponentially over the past few years, drawing much attention from scientific and public, with various types of stem cells being considered for clinical applications. Embryonic stem cells and induced pluripotent stem cells have nearly infinite potential to differentiate in vitro and in vivo into specific progenitor cells and into mature and specialized cell lineages of all 3 embryonic germ layers and are very promising their (15). However, the clinical use of these cells is limited by scientific and clinical issues of safety, such as teratoma formation (16).

Mesenchymal stem cells (MSCs) are a very attractive source of cells for several reasons for clinical treatment. MSCs can be isolated with relative ease from multiple sources, including the bone marrow (BM), adipose tissue (AT), and umbilical cord blood (UCB). MSCs can self-renew for several passages before reaching a state of senescence. They have been used to treat immune diseases because of their inherent immunomodulatory characteristics. MSCs have been shown to produce anti-inflammatory cytokines and suppress the proliferation, differentiation, and function of immune cells in vitro (17). In vivo, transplanted MSCs do not elicit an immune response, allowing their use in allogenic stem cell therapy (18,19). MSCs can be induced to directly differentiate into musculoskeletal cell types. They do not form teratomas when implanted in vivo, making them safe for use in clinical applications (20).

The BM was the first reported source of MSCs. Although BM remains the major source of multipotent MSCs, harvesting is a very invasive procedure. In addition, the number, the potential for differentiation, and the maximum life span of BM-MSCs decrease with increasing age (21). Therefore, intensive investigation is underway to isolate alternative sources of MSCs.

One such alternative source is AT, which can be obtained by a less invasive method and in larger quantities than BM. AT contains stem cells similar to BM-MSCs, called processed lipoaspirate (PLA) cells (22). These cells can be separated from a large number of cosmetic liposuctions, easily grown under standard tissue culture conditions, and show multi-lineage differentiation capacity (22).

UCB is another alternative source that can be obtained by a minimally invasive method without harming the mother or the baby (23). UCB-MSCs have the low frequency, however it has the highest expansion potential among all the cell sources. Considering the clinical applications, the resulting cell numbers may be similar to both BM and AT with higher frequencies.

Of the various MSCs, human UCB-MSCs have the greatest therapeutic potential due to various attributes, such as the ability to home in on injured tissue, low immunogenicity, multi-directional differentiation, and extensive secretion profiles (23). Allogenic UCB-MSCs can be particularly beneficial to elderly patients, or ones with significant comorbidities, in whom the function of autologous MSCs is impaired (24). In addition, UCB-MSCs can be produced commercially in larger quantities without compromising on quality.

The study by Jo et al. (14) was performed to assess the safety and feasibility of autologous ADMSCs for the treatment of rotator cuff disease. Intratendinous administration of a single dose of ADMSCs of up to 1.0×10⁸ cells/3 mL of saline was well tolerated in 19 patients with RCTT, with no evidence of ADMSCs injection-associated serious or serious adverse events at any of the three doses tested. The most common adverse event during the study period was back pain in three patients. Back pain was considered to be unrelated to the ADMSCs by the clinical investigators and was managed with appropriate treatment. Moreover, there was also a follow-up scheduled for up to 6 months after ADMSCs injection to confirm the long-term safety of the treatment. As far as we know, this study documents the first clinical trial of ADMSCs for the treatment of rotator cuff disease. The results showed significant improvement in SPADI scores, shoulder pain, MRI examination, and arthroscopic examination in the mid- and high-dose groups.

Before application of MSCs can be considered, it should be clarified whether there is a critical optimum dose (including repeated doses) of cells for RCTT for safety and therapeutic benefit (25). It is reasonable that the optimal number of cells might vary depending on the size of the
rotator cuff tendon defect, the cell type used, and the particular region in need of regeneration. The dose–efficacy relationship has not been well studied. Only one study so far has assessed the dose-dependent response to allogeneic ADMSCs used to treat lateral epicondylitis in humans, with participants receiving either $10^6$ cells or $10^7$ cells (26). The authors of the study reported faster pain improvement and earlier plateauing of performance scores in the group that received the higher dose, but there were no significant differences in elbow pain and performance changes between the 2 groups at any of the follow-ups. However, too small participants can make it difficult to conclude that the dose-dependent response to the MSCs was significant. In the study by Jo et al. (14), the authors established an optimum ADMSC dose that would show regenerative effects in human patients with rotator cuff disease.

Choosing an effective delivery method in MSCs therapies is a crucial step. The two main methods of delivery are direct injection of a cell suspension alone and implantation of cells that are placed in a matrix carrier vehicle. Direct delivery of MSCs seems to be a more plausible approach for tendons. In their study, Jo et al. (14) injected ADMSCs through the lateral aspect of the shoulder under ultrasonographic guidance. In previous studies, one-third of the blind shoulder injections, performed by experienced physician, were shown to be inaccurate (27).

Polydeoxyribonucleotide (PDRN) is a biologic adjuvant that has the same advantages as UCB-MSCs in terms of commercial mass production. PDRN is a mixture of DNA polymers with chain lengths ranging from 50 to 2,000 bp. It is extracted from trout sperm and purified as a preparation containing a high percentage of DNA. PDRN is a source of pyrimidines and purines, and stimulates nucleic acid synthesis through the salvage pathway (28). PDRN can induce angiogenesis and collagen synthesis and exhibits anti-inflammatory activity (29). Recently, an animal study (18) showed using histological and motion analysis that co-injection of UCB-MSCs and PDRN was more effective than UCB-MSCs injection alone in a rabbit model of chronic traumatic full-thickness RCTT. One published clinical study reported the effectiveness of PDRN in the treatment of chronic rotator cuff disease (30). Therefore, PDRN is a candidate that can enhance the effectiveness of MSCs.

In this study, the shoulder into which the ADMSCs were injected intratendinously was immobilized for 4 weeks using an abduction brace. Deconditioning and reduced muscle activity is estimated to be the most important factors leading to muscle atrophy and loss of function in patients (31). In particular, the treatment of soft tissue and bone injuries by immobilization may result in skeletal muscle atrophy and decreased functional performance (32).

Microcurrent therapy (MT) involves the therapeutic application of very low electric current (less than 1 mA), which is usually sub-sensory to the body. One study demonstrated that MT may prevent progression of GCM muscle atrophy and facilitate the regeneration of muscle cells (32). These effects may increase protein synthesis and stimulate satellite cell proliferation. Therefore, MT has the potential to become an effective therapeutic intervention for the recovery of muscle atrophy due to immobilization.

The intensity of MT plays an important role in the effective treatment of muscle damage. Studies have shown that when an electric current with an intensity of less than 1,000 μA was applied for treating muscle damage which increases the adenosine triphosphate (ATP) production in the tissue and stimulates the amino acids incorporated into the protein in the tissue by 30–40% above the control level along with the activation of myogenic precursor cells (satellite cells) (32,33). On the other hand, when the intensity exceeded 1,000 μA, these bio-stimulatory effects were reversed. One study using 25 μA to treat muscle atrophy reported good regeneration in vivo and in vitro (32). Therefore, it is suggested that a combination of microcurrent treatments may be helpful to prevent muscle atrophy caused by immobilization.

The role of imaging study in the patients with RCTT is to guide treatment decisions whether the patients will be managed conservatively or surgically. Various imaging methods have been used to evaluate the RCTT including arthrography, ultrasound, sonoelastography, arthrosonography, MRI, and MR arthrography. Single-contrast arthrography of the shoulder is a valuable diagnostic method and can produce a high accuracy for the detection of full-thickness tendon tears of the rotator cuff. It can be helpful in assessing deep-surface, incomplete tendon tears of the rotator cuff but may fail to detect bursal-side partial-thickness tendon tears (34).

Jo et al. (14) conducted serial arthroscopy to evaluate the gross findings but not the mechanical properties of regenerated RCTT.

Sonoelastography is a recently developed technique based on ultrasound that assesses tissue elasticity in real-time. It is based on the principle that pressure acting on tissue induces changes in the tissue depending on its elastic properties and tissue compression produces strain (displacement),
which is higher in soft tissue and lower in hard tissue (35). This technique has numerous advantages including its low cost, short examination time, noninvasiveness, and simple clinical accessibility compared to using magnetic resonance elastography to estimate tissue elasticity. Shear wave sonoelastography using acoustic radiation force impulse imaging technology measure shear wave velocity that calculate the mechanical stiffness property of tissue objectively and directly (Figure 1). Sonoelastography may be used to assess the mechanical properties of rotator cuff tendon under clinical conditions and may be more effective than ultrasound and MRI in detecting subclinical tendon disorder (36).

Therefore, sonoelastography can be applied for the early diagnostics and monitoring of treatment by assessing biochemical and pathophysiological changes in the course of RCTT (37).

Arthrosonography involves a combination of ultrasound and single-contrast arthrography and provides more information about the size of RCTT and synovial proliferation in the long biceps tendon sheath or subacromial bursa for treatment planning (38). Additionally, it is an effective imaging method for assessing the postoperative integrity of the repaired rotator cuff tendon and appears to be an appropriate imaging method for replacing ultrasound (39).

MR arthrography is more sensitive and specific than either MRI or ultrasound in the diagnosis of both partial- and full-thickness RCTT. Therefore, MR arthrography can be used when ultrasound and MRI diagnosis do not provide definitive results (40).

Jo et al. (14) concluded that intratendinous injection of autologous ADMSCs for the treatment of rotator cuff disease is feasible, safe, and effective and provide evidence of regeneration of tendon defect without surgery. However, they did not provide any data on the size of rotator cuff tear, which is important for determining the treatment method.

In conclusion, this study provides a foundation for future stem cell-based treatment approaches in humans, which can be used to promote the clinical application of various therapies combining stem cells and chemical agents and/or physical methods to facilitate recovery from RCT.
Additionally, the use of sonoelastography, an imaging method that can assess the quality of the regenerated rotator cuff, may further aid clinical management.

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**Footnote**

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